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**User Guide for**

# ***Kinetic Studio*** **5.x**

An Application for the Acquisition and Analysis of Kinetic Data

Revision 19.01  
19 October 2018



Suppliers of Hi-Tech Scientific Instruments

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# Contents

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<b>Contents</b> .....	<b>3</b>
<b>Introduction</b> .....	<b>9</b>
Health and Safety at Work Act, 1974 - UK .....	9
<b>Installing Kinetic Studio</b> .....	<b>10</b>
Driver Installation .....	14
USB / PCI Counter Timer Cards .....	14
Data Acquisition Card .....	15
Post Driver Installation .....	16
Initialising the Counter Timer Card Ready for Use .....	17
Upgrading Kinetic Studio .....	17
Starting Kinetic Studio .....	18
<b>Kinetic Studio Overview</b> .....	<b>19</b>
Introduction .....	19
KinetAsyst Stopped-Flow Photomultiplier Control Panel .....	20
Conductivity (Stopped-flow) Control Panel .....	20
T-Jump Control Panel .....	21
The Workspace .....	22
Navigating Kinetic Studio .....	23
Zoom .....	23
Scroll .....	23
Identify a Trace .....	23
Extract Trace Under Mouse .....	23
Available Chart Context Menu Features .....	23
Autoscale All .....	23
Fix X-Axis .....	23

Fix Y-Axis .....	24
Sticky Cursor .....	24
Find Nearest Trace .....	24
Extract Nearest Trace .....	24
Add Annotation .....	24
Properties .....	25
Graph Options .....	25
How Data is Displayed .....	26
Selecting a Single Thumbnail .....	27
Selecting Multiple Thumbnails .....	27
Displaying a Single Dataset .....	28
Overlaying Multiple Datasets .....	28
Data Manipulation – Analysis, Fitting, Management .....	29
Calculator .....	29
Analysis Control Panel .....	30
Dataset File Management .....	32
How to Change the Current Working Folder .....	32
How to Save Data .....	33
How to Export Data or Fit Results .....	34
How to Export to SPECFIT/32 .....	35
Importing into SPECFIT/32 Method 1 .....	37
Importing into SPECFIT/32 Method 2 .....	39
ReactLab .....	43
Exporting from Kinetic Studio into ReactLab .....	44
How to Load a Dataset .....	46
How to Import Data .....	47

---

<b>Stopped-Flow; Initial Setup and Getting Started</b> .....	<b>49</b>
<b>Photomultiplier</b> .....	<b>51</b>
How to perform a Scan Blank (Absorbance) .....	52
Scanning the Blank .....	56
How to set the Reference / Signal Levels at a Single Wavelength .....	59
General usage instructions for manual setup .....	61
Manual Setup for Absorbance .....	61
Manual Setup for Fluorescence .....	62
Fluorescence Excitation Scanning .....	64
Fluorescence Emission Scanning .....	65
Setting the Data Type and Dataset Parameters .....	66
Executing a Shot .....	67
Optimising the Dataset Parameters .....	68
Setting the Run Time .....	69
Specifying a file name .....	69
Adding Notes to a Dataset .....	70
<b>CCD Array</b> .....	<b>71</b>
Launching Kinetic Studio in CCD / Spectrometer Mode .....	71
Standalone CCD Mode - Set Up, Calibrate, Scan a Blank .....	73
CCD Hardware Options .....	78
Binning and Cropping .....	78
Trigger Modes .....	79
Slice .....	80
SVD Analysis .....	81
Spectrometer Mode .....	83
<b>How to use the Sequence Setup</b> .....	<b>85</b>

---

Average sequence .....	85
Time Delay Sequence .....	86
Wavelength Sequence .....	86
Age Time DX Sequence .....	86
In the Event a Sequence Stalls .....	86
<b>Double Mixing Stopped-Flow .....</b>	<b>89</b>
Enabling Double Mixing Modes .....	90
How to Reload the Drive Syringes During a Shot .....	92
How to Reload the Drive Syringes During a Single Mixing Shot .....	92
How to Reload the Drive Syringes During a Double Mixing Shot .....	93
Advanced Drive Positioning .....	94
<b>qPod Mode .....</b>	<b>96</b>
Executing a qPod Shot .....	99
<b>T-Jump Mode .....</b>	<b>101</b>
How to Setup for an Experiment in T-Jump Mode .....	102
Overview .....	102
Initial Setup .....	102
Setting the Data Type and Dataset Parameters .....	102
Data Trigger Offset .....	103
Time Constants .....	103
Recharge Time .....	104
Setting the References / Spectrometer Manual Setup .....	105
Manual Setup for Absorbance .....	106
Manual Setup for Fluorescence .....	107
How to perform a Scan Blank (Absorbance) .....	108
Scanning the Blank .....	112

---

T-Jump Shot Sequence .....	113
<b>Conductivity Mode .....</b>	<b>115</b>
Conductivity Description .....	116
How to Setup for an Experiment in Conductivity Mode .....	116
Overview .....	116
Initial Setup .....	117
Setting the Dataset Parameters .....	118
Setting the Run Time .....	118
Specifying a file name .....	119
Adding Notes to a Dataset .....	119
Adjusting Hardware Options and Signal Conditioning .....	120
How to Acquire a Single Conductivity Shot .....	122
How to Acquire a Conductivity Shot Series .....	123
<b>Data Manipulation .....</b>	<b>125</b>
Introduction .....	125
Rotating a Dataset .....	125
Extracting Traces from Datasets .....	126
Dataset Calculator .....	128
Previewing a Dataset .....	128
Entering Datasets or Traces into the Calculator .....	129
Calculator Functions .....	131
Copying a Dataset .....	131
Extracting a Trace .....	131
Convert a Dataset Type .....	131
Convert to Absorbance .....	131
Convert to Concentration .....	132

---

Convert to Conductivity .....	132
Convert to Fluorescence .....	132
Convert to Percentage .....	132
Combine .....	133
Rotate .....	133
Normalise .....	133
Smooth .....	134
Power .....	134
Derivative .....	134
Min .....	135
Max .....	135
Abs .....	135
Log .....	135
Ln .....	136
<b>Data Fitting .....</b>	<b>137</b>
Introduction .....	137
Fitting a Dataset with a Standard Model .....	137
Custom User Definable Equations .....	141
Entering a New Custom Model .....	142
Selecting a Custom Model .....	145
<b>Support, Updates and Suggestions .....</b>	<b>147</b>

# Introduction

The Kinetic Studio software package provides a wide range of facilities within a friendly, familiar format that is easy to use.

Kinetic Studio is primarily designed for the acquisition, analysis and management of kinetic data associated with the Hi-Tech Scientific instruments for transient kinetics. It is a feature rich package that provides an efficient and intuitive means of managing instruments in the laboratory and speeds data processing by providing a host of data handling and visualisation facilities. These include an array calculator, logarithmic time-base and high-speed graph plotting as well as offering automated modes of instrument operation for rapid and reproducible throughput of experiments. An oversampling function allows data points to be averaged, improving signal to noise and resolution.

Kinetic Studio also includes a data file converter which brings compatibility with other, third party instruments for spectroscopy and popular data processing packages.

## **Health and Safety at Work Act, 1974 - UK**

In accordance with the above Act, we ensure that all products manufactured or supplied by TgK Scientific Limited are safe and without risk to health when used by suitably trained personnel following our instructions.

# Installing Kinetic Studio

Kinetic Studio is made to run on computers running Windows 7 or Windows 10. The software may function on other versions of Windows however some functionality may be lost.

Installations involving instrument control or data acquisition managed by Kinetic Studio involves the installation of computer hardware devices. Please follow the guide relevant to the equipment supplied.

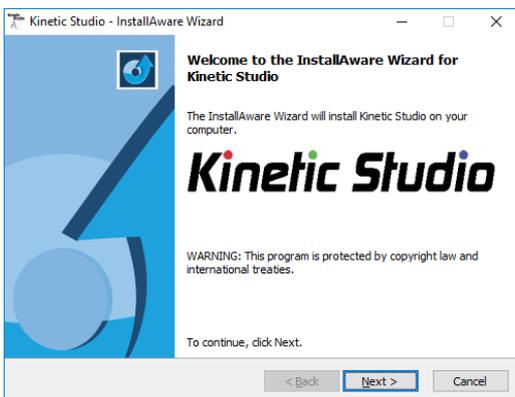
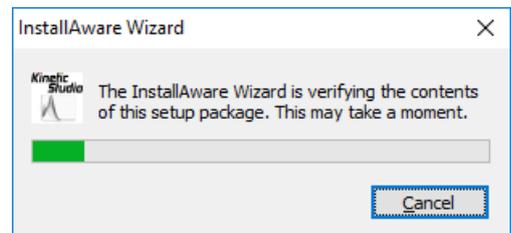
If a USB / PCI based Counter Timer card or Data Acquisition card has been supplied and is already fitted within the computer, Windows may display a 'New Hardware Found' message. Please cancel these messages.

The Kinetic Studio installer as part of the installation process will install the device drivers automatically.

If hardware devices cannot be found or do not install correctly, please check the Device Manager (accessible from the right click menu of the windows button). If there are any unknown devices, please remove them and reboot the computer.

If the supplied cards are not yet installed, please switch off the computer and install them now taking note to cancel any new devices found messages when the computer is started.

For single file installations, the setup program will unpack itself and prepare the PC for installation. For DVD installations, the installation process should start automatically. If not, please explore the DVD and run the kineticstudio\_#.###.exe executable.

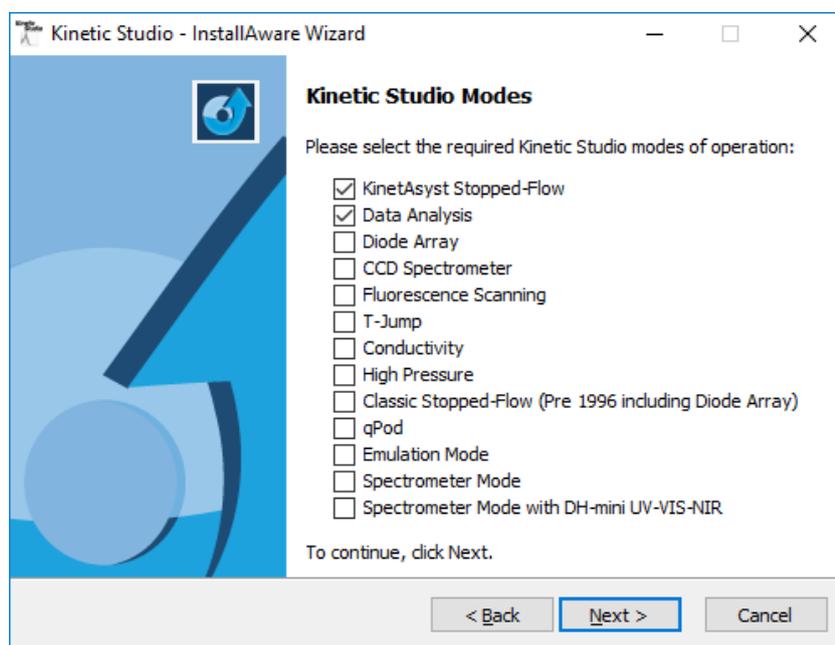


If there are any required components that need to be installed into the operating system, these will be shown as below. Click 'Next' to begin the installation.

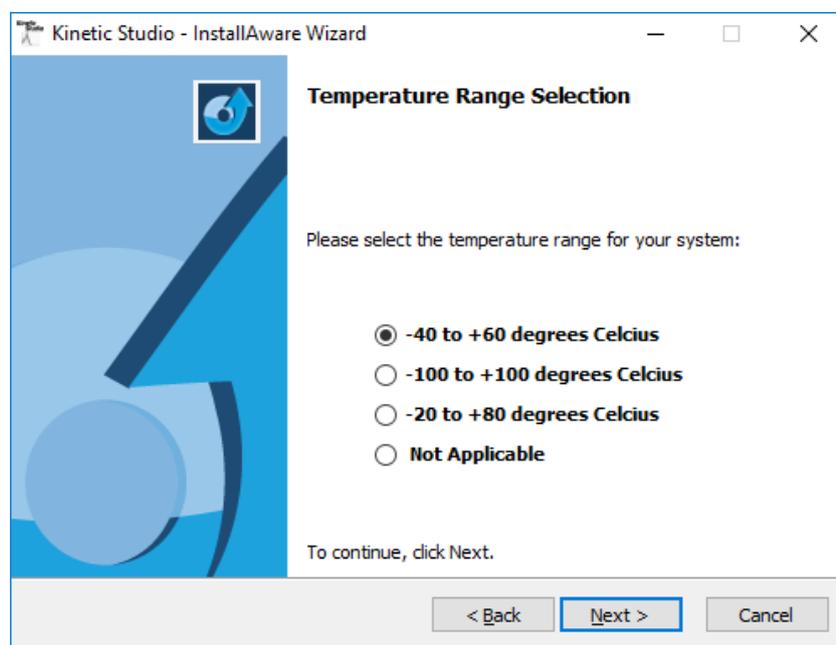
Once this has completed, installation of the main Kinetic Studio application and related drivers will commence. Click 'Next' to proceed.

Click 'Next' to begin the installation.

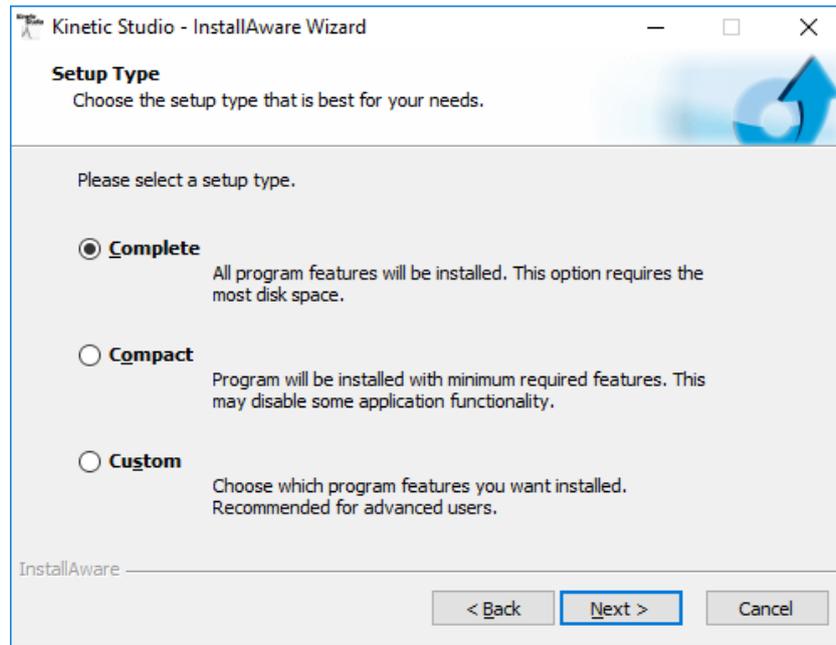
After agreeing to the terms and conditions, the installer will ask which modes of operation are required. Please select the required modes. SF-61DX2 users will need to select KinetAsyst Stopped-Flow as a minimum.



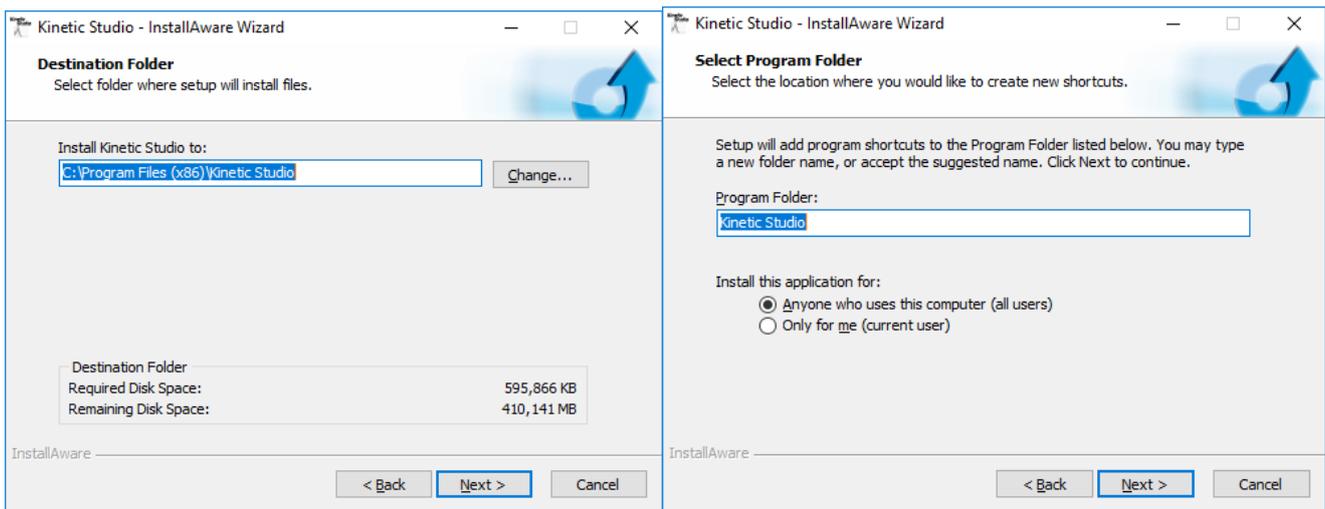
The Data Analysis option provides a quick shortcut to bypass any hardware scanning and go directly into the data analysis mode.



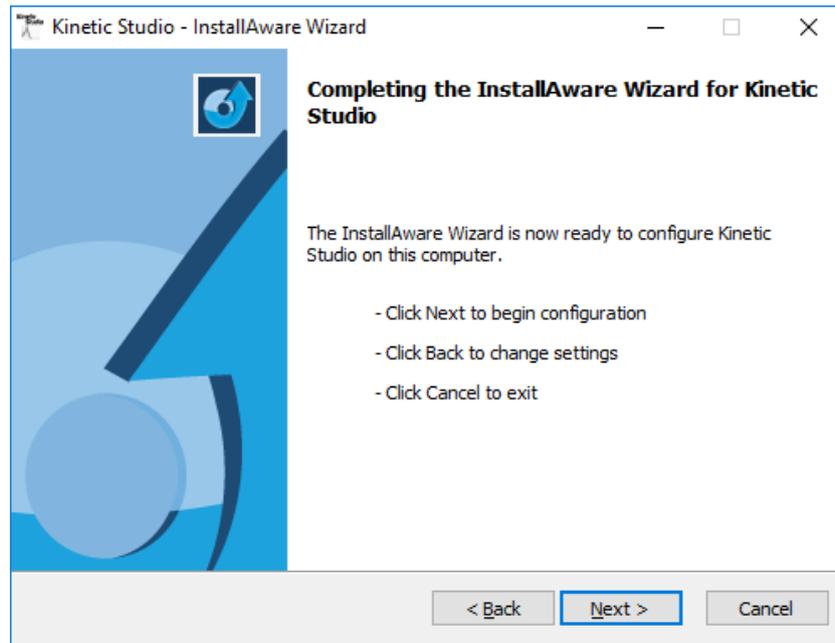
The next screen will prompt for the instrument temperature range. The default, -40 to +60 is suitable for most KinetAsyst Stopped-Flow instruments.



When prompted for the type of installation, please choose 'Complete'.



Generally it is recommended that Kinetic Studio is installed to the default directory.



The main Kinetic Studio program files will now be installed...

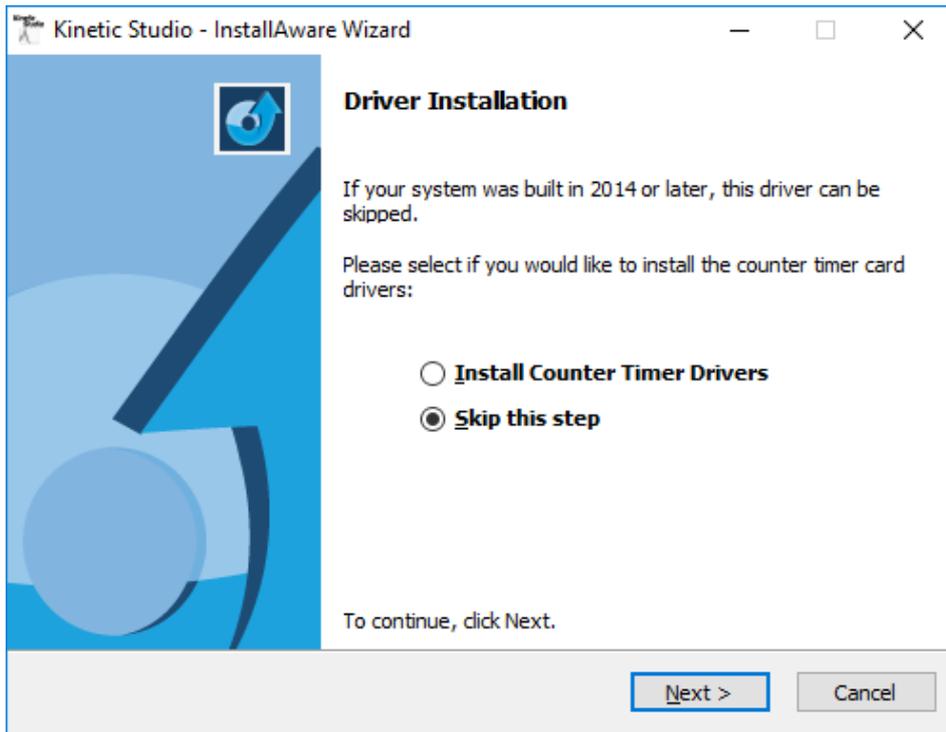
## Driver Installation

The next stage of the installation process allows the user to select required device drivers. These are required for controlling the instrument and acquiring data.

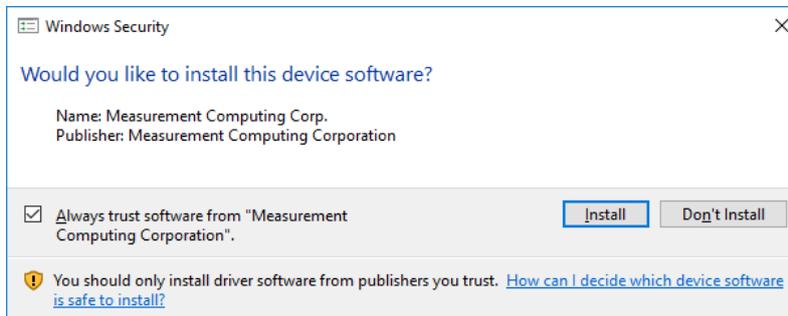
### USB / PCI Counter Timer Cards

Skip to the Data Acquisition driver section if there is not a Measurement Computing counter timer card present. Systems built in 2014 or later will not have a Counter Timer Card.

The first option is to install the Counter Timer card driver.



Please select the option and continue (if the counter timer card is present).



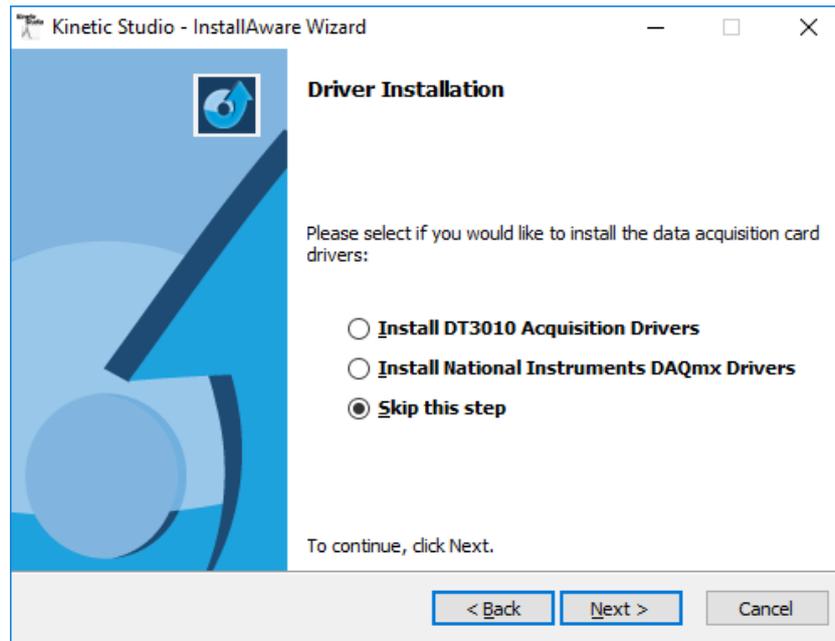
Depending on the operating system, the new driver dialog may be shown. Place a tick in the 'Always trust software from "Measurement Computing Corporation"' and 'Install' the device software.

## Data Acquisition Card

The next option is to install the high speed data acquisition drivers. If no hardware is present select 'Skip this step'.

For systems made before 2014 select 'Install DT3010 Acquisition Drivers'. For systems made in 2014 or later select 'Install National Instruments DAQmx Drivers'.

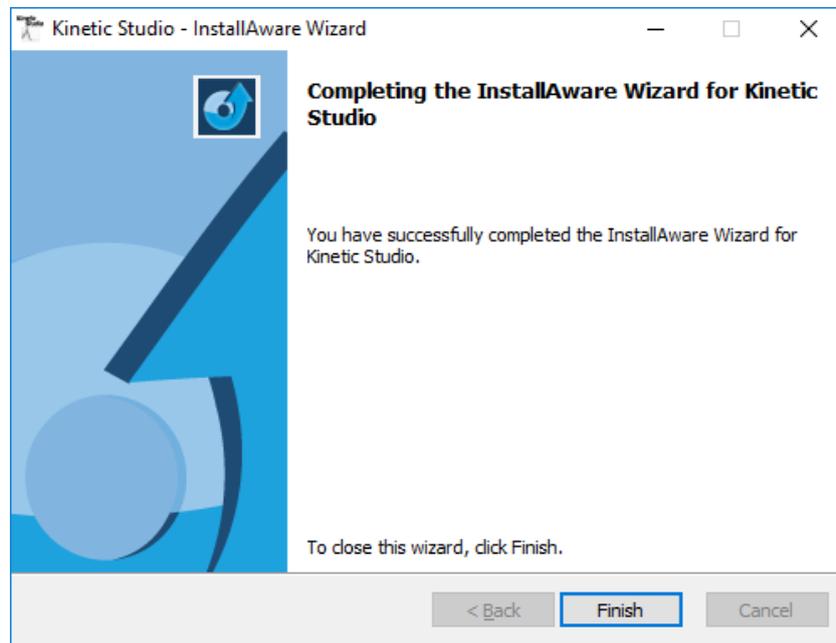
Press 'Next' to continue.



Depending on the operating system and the driver selected, the new driver dialog may be shown. Place a tick in the 'Always trust software from "data translation" and 'Install' the device software. On a 64bit version of Windows, both the 32bit and 64bit drivers need to be installed so the dialog may appear twice.

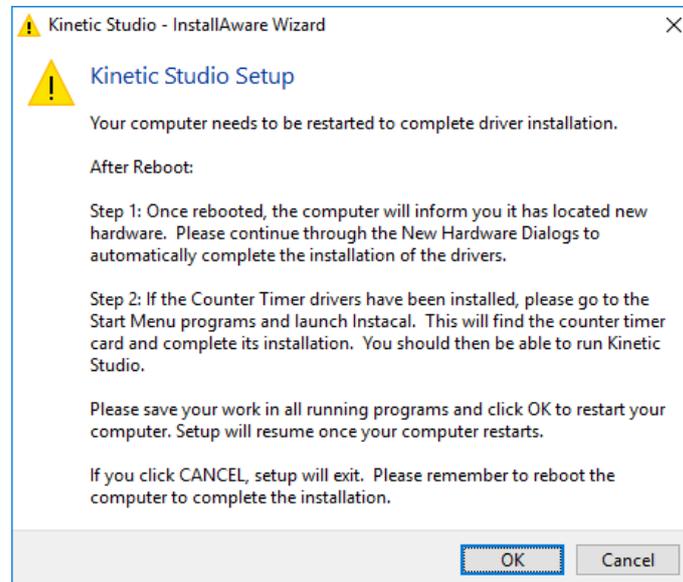
## Post Driver Installation

The installation is nearly ready.



Click 'Finish' to complete to installation.

The installer will display the following message if drivers have been installed.



Please read the message and make sure all open work is closed. Then click the [OK] button. This will reboot the computer.

When the computer has rebooted, new hardware may be found by the operating system (depends which version of Windows).

The drivers have been installed by the Kinetic Studio installer so Windows will find them automatically on the hard disk.

## **Initialising the Counter Timer Card Ready for Use**

If the counter timer card has been installed please run the Instacal program which will have been automatically installed.

Instacal initialises the Counter Timer card ready for use. The program can be found in the Start Menu under Measurement Computing. Once started, Instacal should automatically find the card. It will then display a summary, after which Instacal can be closed. The computer is ready to run Kinetic Studio.

## **Upgrading Kinetic Studio**

If an new version of the Kinetic Studio Installer is run while an old version of Kinetic Studio is installed on the PC, the old version will be uninstalled before the new version is installed.

Once the uninstall is complete follow the instructions for **Installing Kinetic Studio** on page 10.

## Starting Kinetic Studio

Launch **Kinetic Studio** from the icon on the desktop or the program icon in the Start Menu.



During software installation, a shortcut to a folder named '**SF Data**' is created which is located in the '**My Documents**' folder by default. The location of this file can be changed within Kinetic Studio.

To perform hardware troubleshooting, double-click the '**Engineers Toolkit**' icon.

Other shortcuts will have be created for other modes if selected during installation.

On launching the software, the main Kinetic Studio start-up screen will be displayed. This will display the initialisation steps (depending on the version of Kinetic Studio)

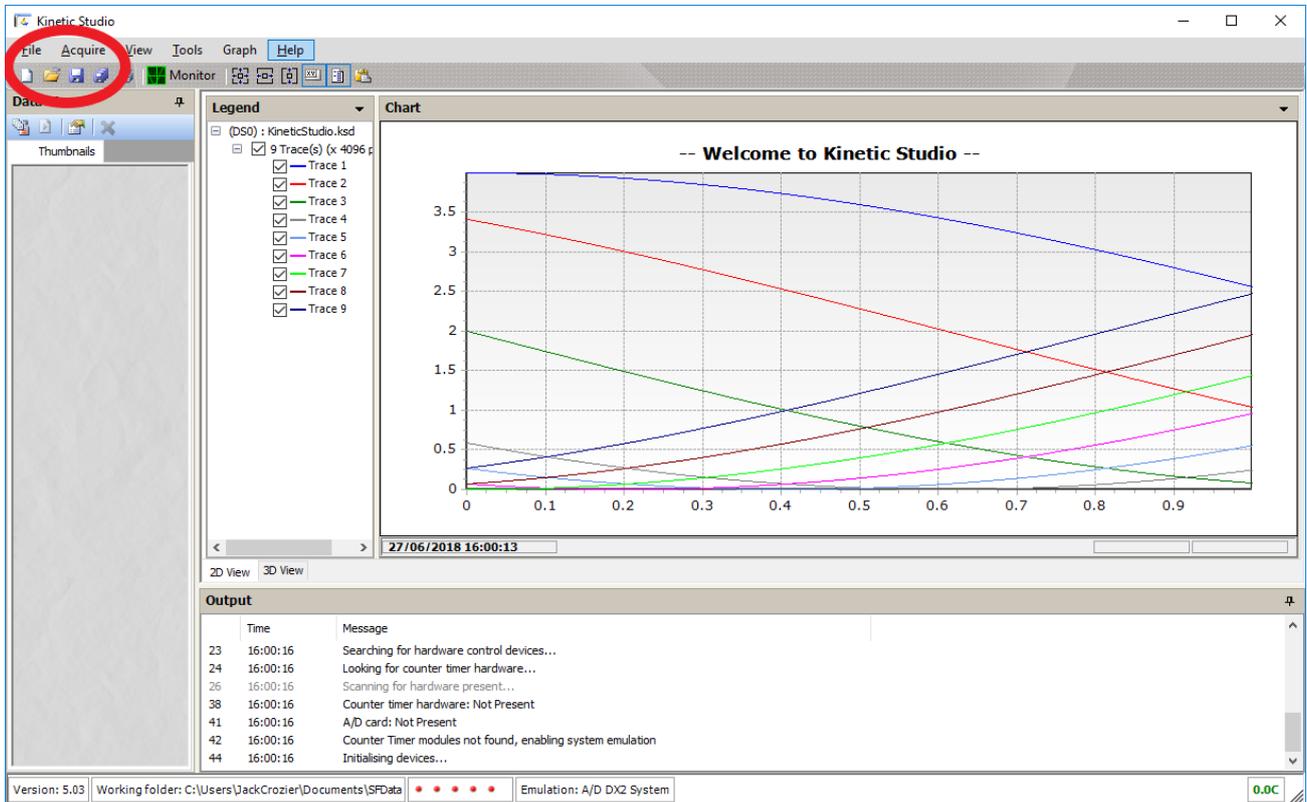


If Kinetic Studio is unable to verify the equipment is present or there is a communications problem, it will default to emulation mode. Emulation mode will allow the software to be used for data analysis.

# Kinetic Studio Overview

## Introduction

When Kinetic Studio has fully loaded, a screen similar to the following will be displayed:

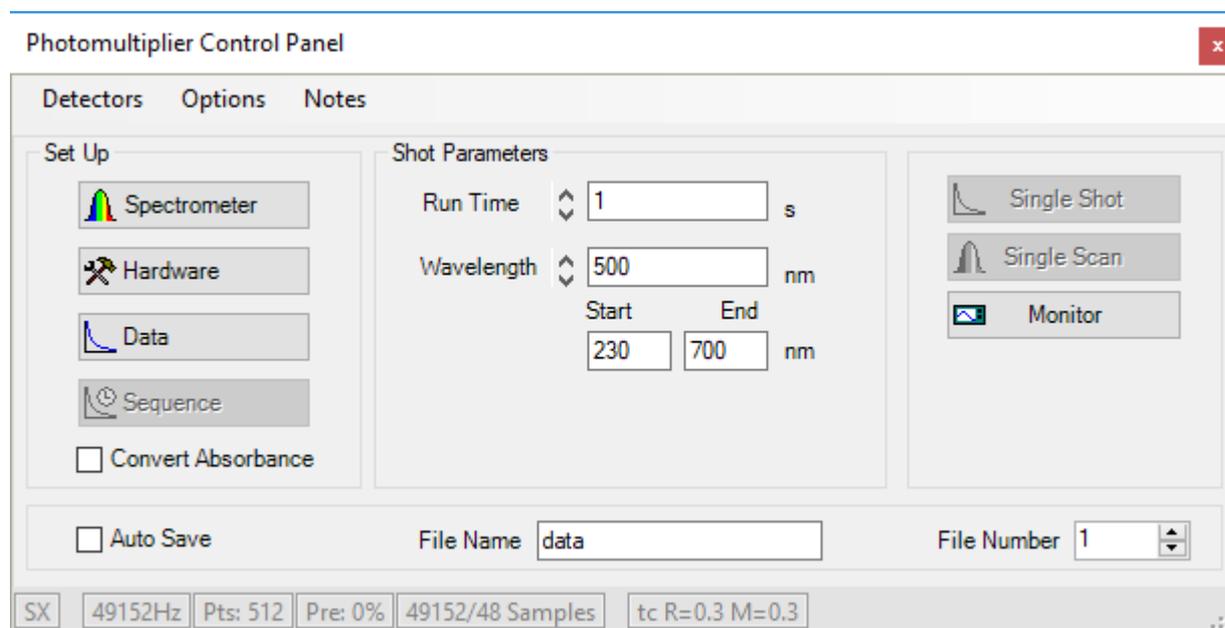


The red circle is highlighting the **'New Document'** icon  and the **'Acquire'** menu.

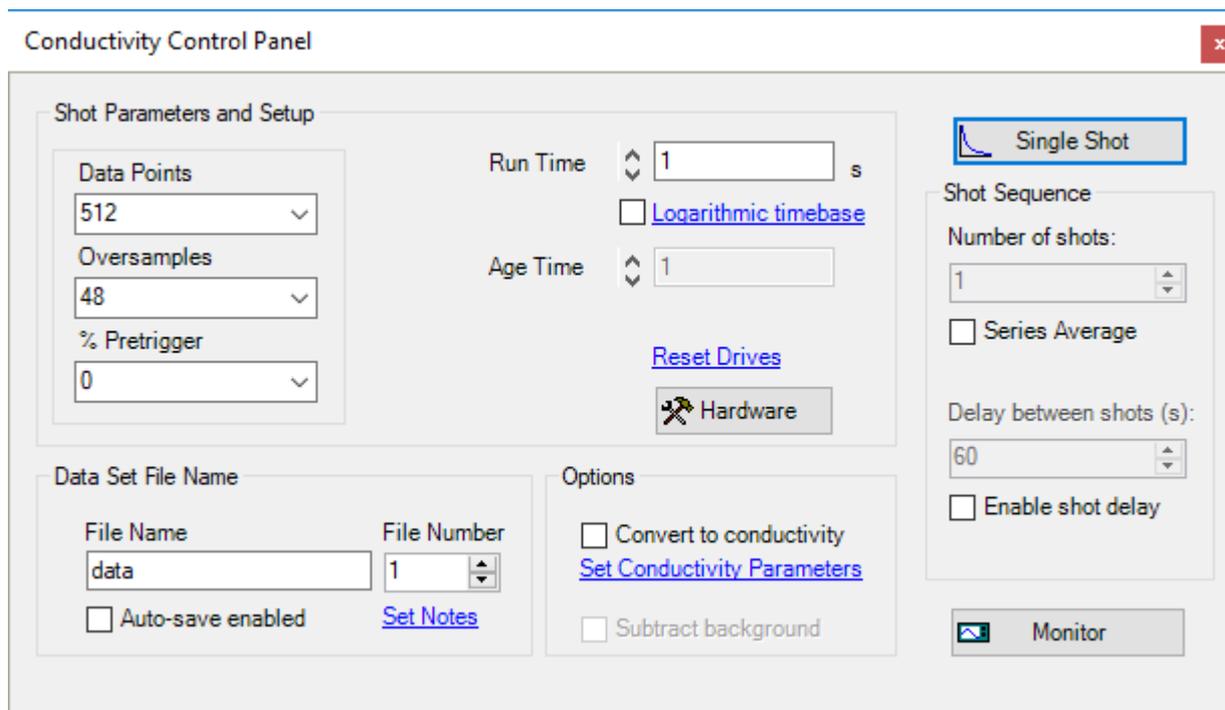
The **'New Document'** icon will display the control panel. The control panel can also be opened from the **'Acquire'** menu or by pressing **'F8'**.

The control panel that appears will depend on the installed option or options and modes of operation available for the available system. The following shows the most common examples:

## KinetAsyst Stopped-Flow Photomultiplier Control Panel



## Conductivity (Stopped-flow) Control Panel



## T-Jump Control Panel

Photomultiplier Control Panel - Temperature Jump

Detectors Options Notes

Set Up

- Spectrometer
- Hardware
- Data
- Sequence
- Convert Absorbance

Shot Parameters

Run Time  s [Fastest](#)

Wavelength  nm

Start  End  nm

Manual Jumps

Single Shot

Single Scan

Monitor

Auto Save

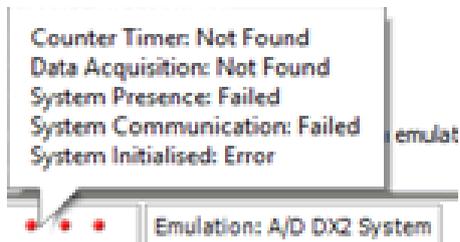
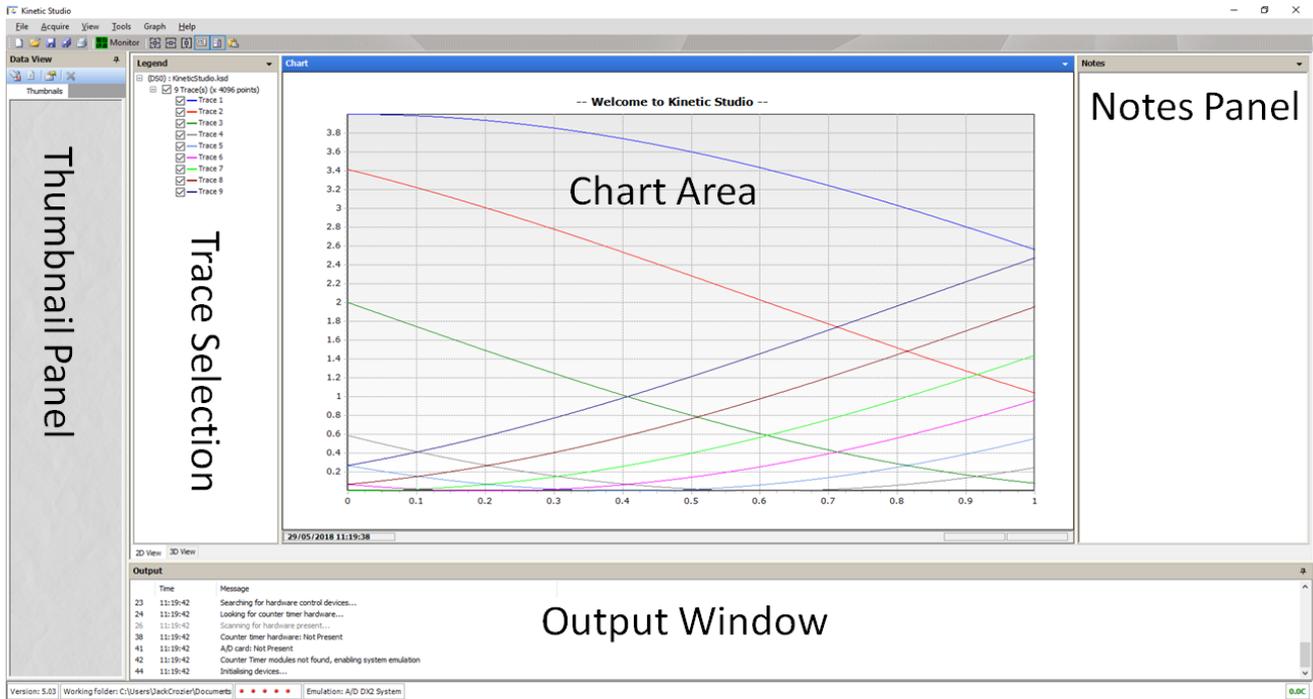
File Name

File Number

SX 24576Hz Pts: 512 Pre: 0% 24576/48 Samples tc M=0.3

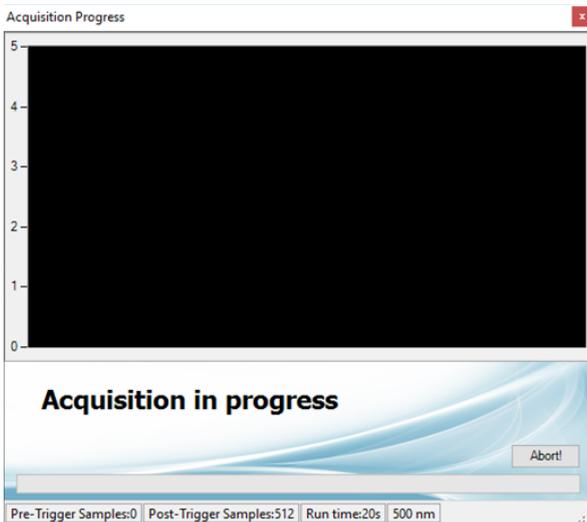
# The Workspace

An example workspace is shown below:



If there is a problem at start-up, red indicators at the bottom of the screen will be visible. Hover the mouse over the indicators to provide additional information.

Please Note: In '**Conductivity**' mode, certain of these indicators will be red since the instrument is not being controlled.



Kinetic Studio features a real-time progress dialog for longer run times.

Where using photomultiplier detection, linear time base, shots over ten seconds automatically enable the real-time display.

## Navigating Kinetic Studio

### Zoom

To zoom in on a Chart, press the left mouse button at the top left hand corner of the area to zoom in on and maintaining the mouse button pressed, drag out the rectangle to the bottom right-hand corner of the zoom area. Release the mouse button and the Chart will redraw the area selected.

To undo the zoom, press the left mouse button anywhere on the Chart area and drag up and left with the mouse button depressed. Release the button and the Chart will redraw to the originally defined Chart area. Alternatively use the context menu to select the '**Autoscale All...**' option.

### Scroll

To scroll a Chart across, press the right mouse button and maintaining the mouse button pressed, drag the mouse in the desired direction to scroll the Chart. After releasing the mouse button the Chart will remain at the new location.

To undo the scroll, press the left mouse button anywhere on the Chart area and drag up and left with the mouse button depressed. Release the button and the Chart will redraw to the originally defined Chart area.

### Identify a Trace

There are two ways to identify a given trace. A known trace can be selected from the legend which will highlight it in the graph area. Alternatively use the mouse pointer and then press the middle mouse button whilst hovering over the unknown trace. This will both temporarily highlight it and select the trace within the legend panel.

### Extract Trace Under Mouse

To extract a trace quickly and easily from the chart view, move the mouse cursor over the trace and then using the context menu (right mouse button), select '**Extract Nearest Trace...**'.

### Available Chart Context Menu Features

There are a number of facilities available via the chart context menu (right mouse button).

Most of these features are self-explanatory. Here is a quick summary of some of the key items:

#### **Autoscale All**

Reset the zoom to normal ensuring all of the data is visible on both the X and Y axis.

#### **Fix X-Axis**

This will prevent the X-Axis from changing. This applies to both new shots and viewing existing data. When enabled it will fix the X-Axis to the current settings.

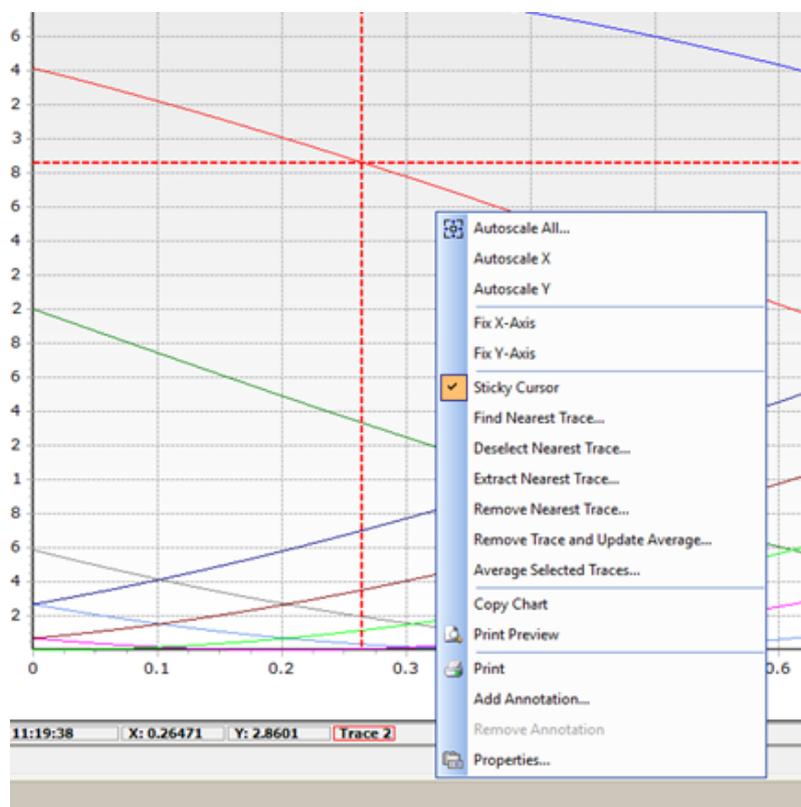
## Fix Y-Axis

This will prevent the Y-Axis from changing. This applies to both new shots and viewing existing data. When enabled it will fix the Y-Axis to the current settings.

## Sticky Cursor

Enabling this option will display a large cross-hair in the same colour as the currently selected trace. When enabling the option it will use the nearest trace to the mouse cursor. The trace will also be identified in the status bar at the bottom of the screen.

To move between traces, use the keyboard cursor up and down keys. This will scroll through the traces.



## Find Nearest Trace

The nearest trace to the mouse cursor will be identified and highlighted.

## Extract Nearest Trace

The nearest trace to the mouse cursor will be identified and extracted into a new dataset.

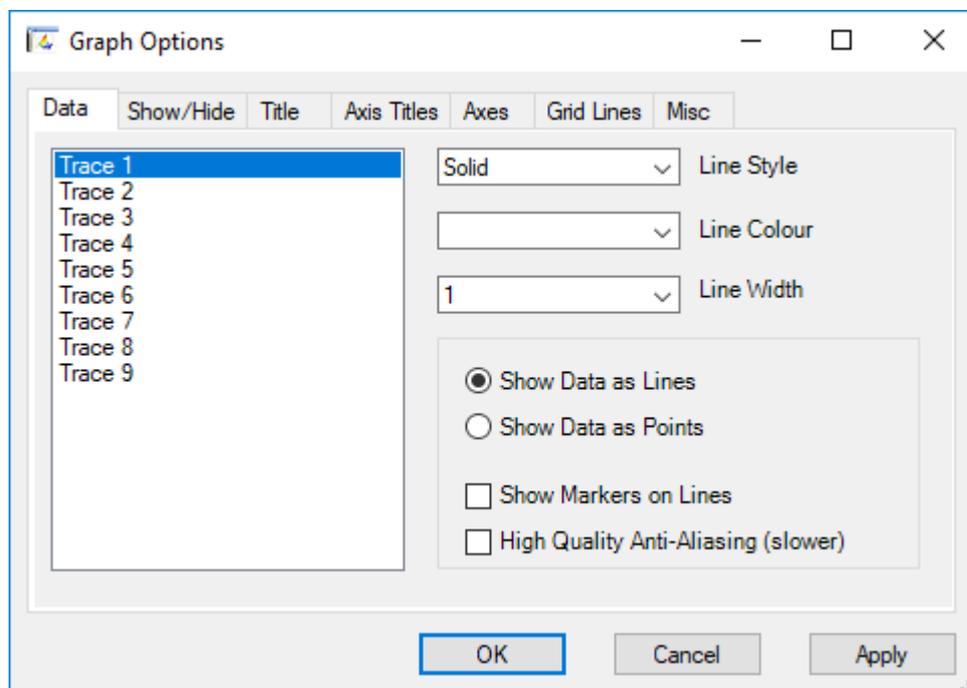
## Add Annotation

An annotation can be added to the current cursor position.

## Properties

The properties option will display the graph options.

## Graph Options



There are a number of graph options that allow the chart view to be customised. This includes trace colours, line width, graph title, axis titles and more.

The main options panel allows the trace to be displayed as a line, as points, or a combination of the two by enabling lines and markers.

If a higher quality view of the display is required, the high quality anti-aliasing option can be enabled. Please note that this will slow down the chart display - particularly with large datasets such as CCD data.

## How Data is Displayed

All data held in memory when initially created, whether it be from an acquisition, or loaded from disc, is given a small representation situated in the left portion of the Kinetic Studio window. This small window is called a thumbnail, and is an integral part of the software for the display and manipulation of data.



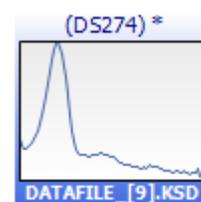
Taking a closer look at a thumbnail, it can be seen that it comprises of three elements.

The top title section provides information such as:

(DS274): dataset number.

\*: Indicates the dataset is currently unsaved.

Additionally if the dataset has been fitted, this will be indicated in the title.

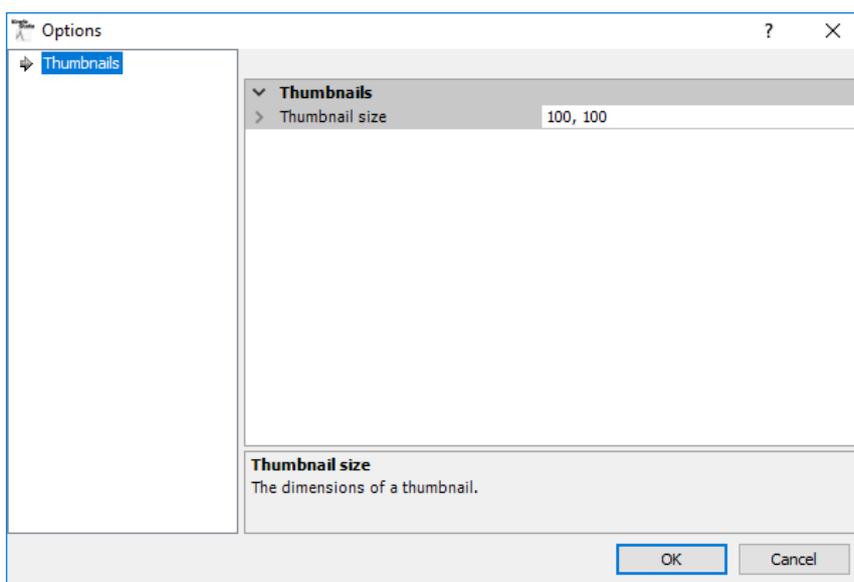


The graph represents the first few traces within a dataset.

The text at the bottom of the dataset is the filename.

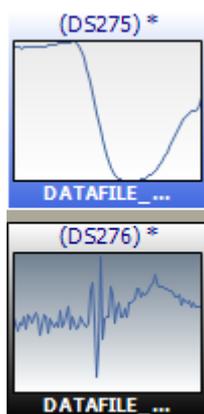
The thumbnails can be resized according to the users' preference.

Pressing the **'Options'** button at the top of the thumbnail panel will display the following dialog.



## Selecting a Single Thumbnail

A single thumbnail can be selected by using the mouse. Simply move the cursor over the thumbnail and click the left mouse button.



DS275 represents an unselected thumbnail.

DS276 is shaded in black representing a selected thumbnail.

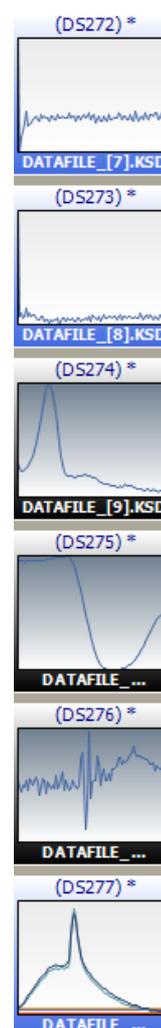
## Selecting Multiple Thumbnails

Multiple thumbnails can be selected with the mouse.

This is done in one of two ways:

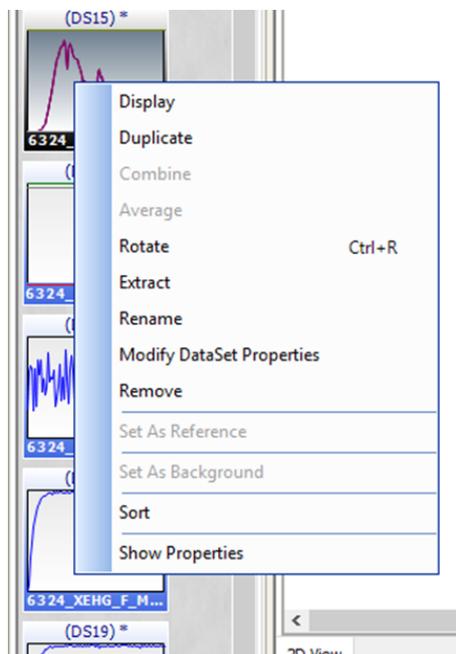
Selecting a group of thumbnails together can be achieved by clicking on the first thumbnail with the mouse and then clicking on the last thumbnail in the group whilst holding the **'SHIFT'** button on the keyboard.

Selecting individual thumbnails together can be achieved by clicking on each thumbnail with the left mouse button whilst holding the **'CTRL'** key down.



## Displaying a Single Dataset

A single dataset can be displayed by double clicking the thumbnail or by right clicking on a thumbnail and selecting the option to **'Display'**. A single trace, when displayed, will have its thumbnail turn red, or green if another thumbnail is selected.



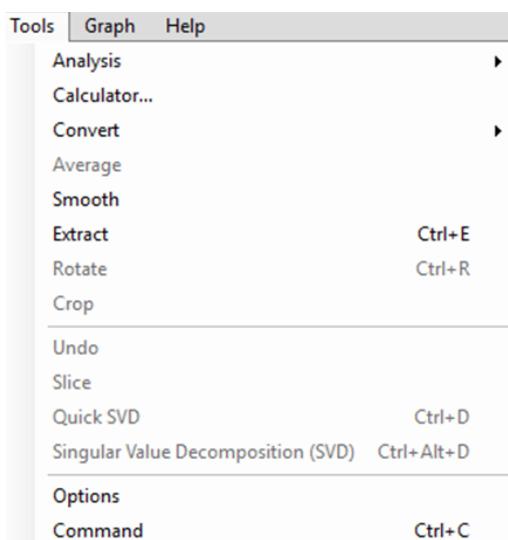
## Overlaying Multiple Datasets

To overlay multiple datasets, firstly please select multiple thumbnails using the method above.

Next, right click on one of the selected thumbnails to display the context menu. From there select the **'Display'** item.

## Data Manipulation – Analysis, Fitting, Management

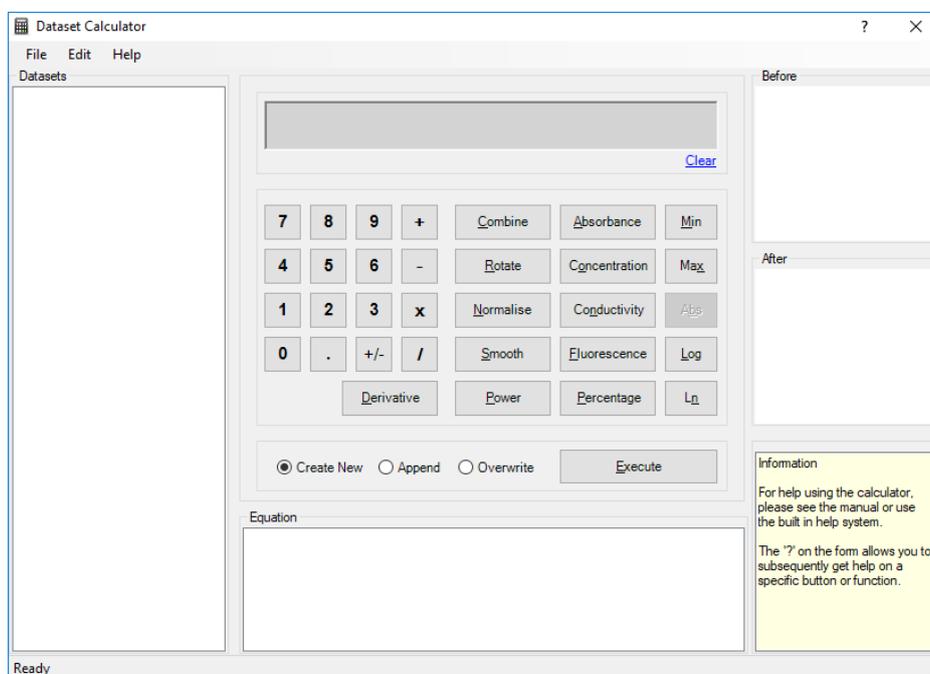
Data analysis functions can be found in the Tools menu.



The functions available will depend on the type of dataset.

### Calculator

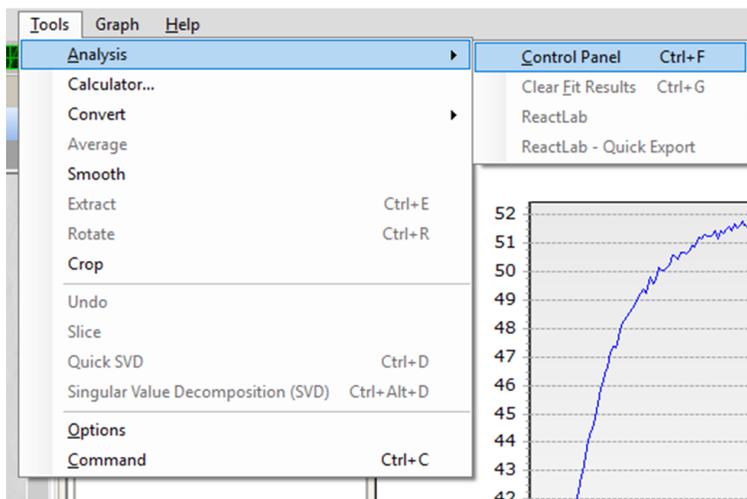
The dataset calculator allows traces or constants to be added, subtracted, multiplied or divided into a trace. The resulting trace can replace the original, be added to the dataset or added to a new dataset. More information can be found in **Dataset Calculator** on page 128.



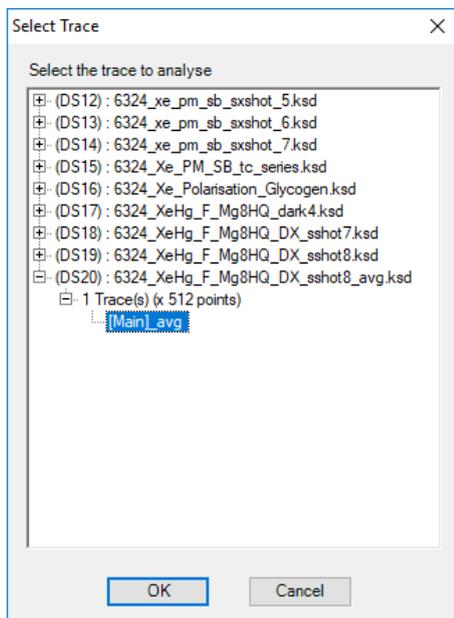
For convenience conversion operations can be quickly accessed via the '**Convert**' sub menu.

## Analysis Control Panel

Data analysis functions can be found in the **'Tools' -> 'Analysis'** menu, by pressing the key combination **'Ctrl' + 'F'** or by selecting the fitting icon from the toolbar:

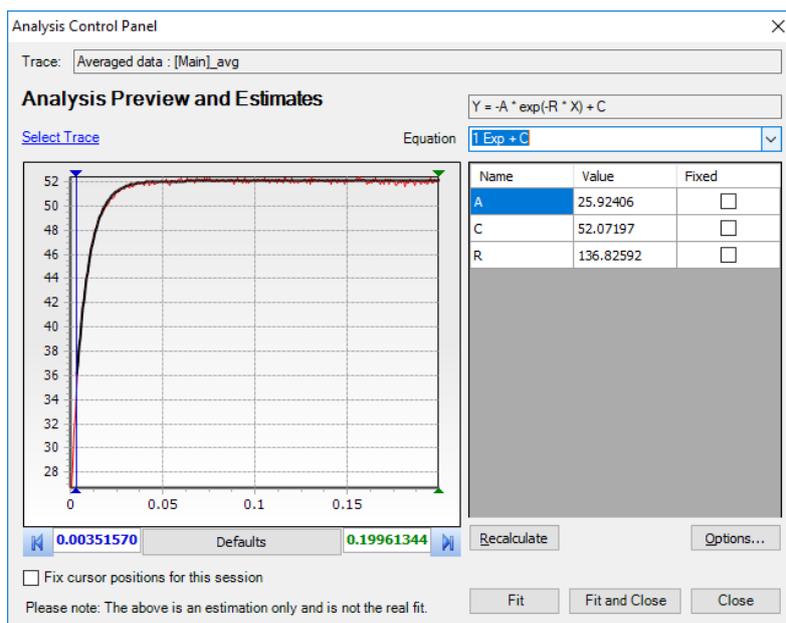


The 2D / 3D / Analysis results view can be selected at the lower part of the screen as highlighted above in red.



Kinetic Studio will ask which trace to fit. This is done by displaying a summary screen of all datasets in memory.

After selecting the trace, the Analysis Control Panel will be displayed.

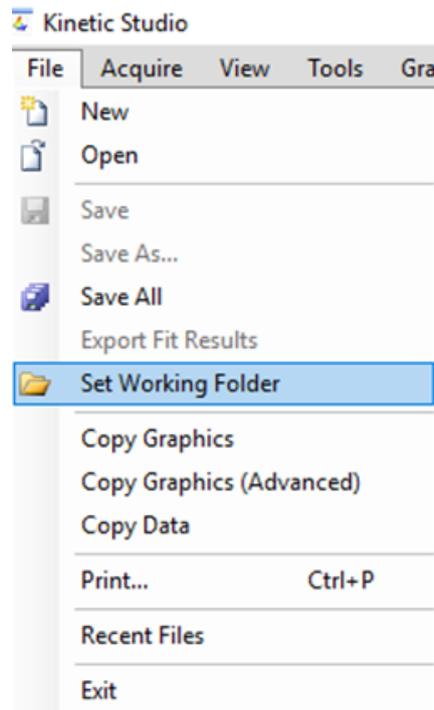


For further information about data fitting, please see **Data Fitting** on page 137.

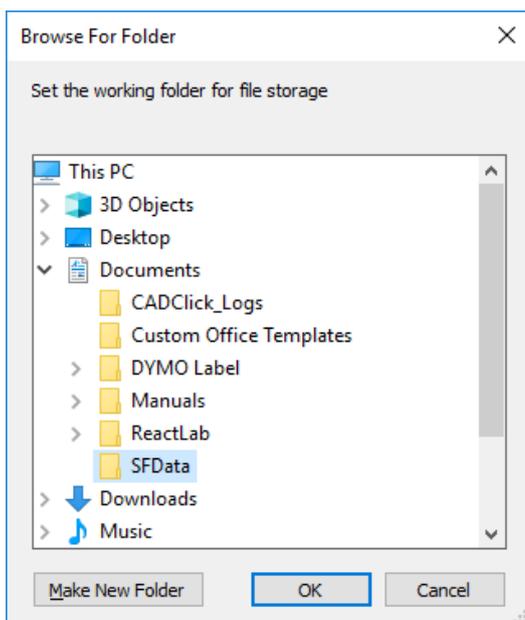
## Dataset File Management

### How to Change the Current Working Folder

To set the current working folder, i.e. the folder in which all subsequent files will be saved, go to the **'File'** menu and select the **'Set Working Folder'** option.



This will produce the following window:



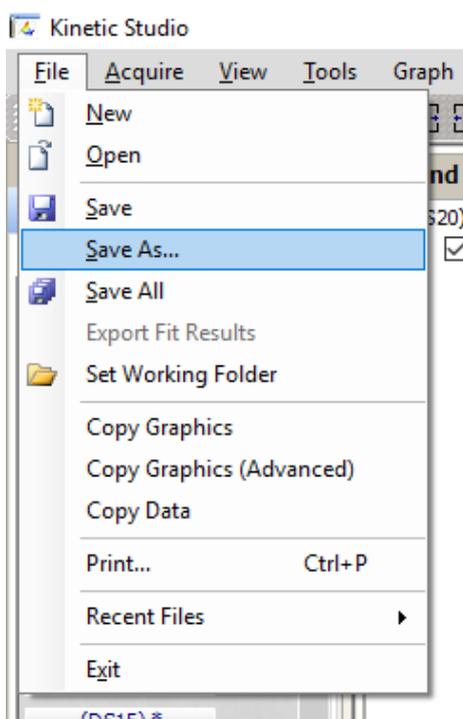
Choose the drive and folder to use, and select the **'OK'** option.

This folder will be used whenever the user wishes to load or save files and will be remembered when Kinetic Studio is next launched.

## How to Save Data

To save data items to disk, either use the **'Save'** option from the **'File'** menu, or use the toolbar button, which resembles a floppy disk. The status bar at the bottom of the main Kinetic Studio window will display the progress of the saving process.

To save all the datasets to disk, use the **'Save All'** button, the stack of disks, either on the toolbar or in the **'File'** menu.



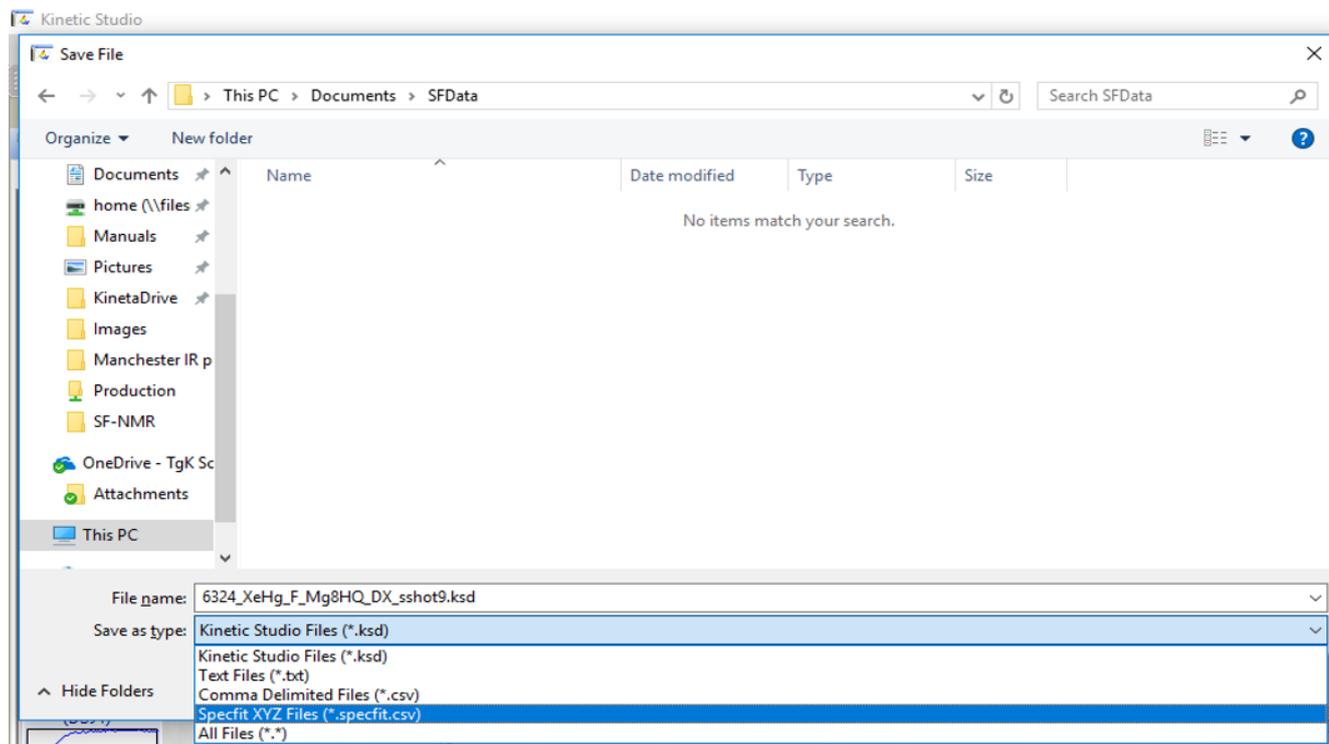
To save the file with a specific filename or to a specific location, use the **'Save As...'** menu item. This will display a standard save dialog allowing the data to be saved on the local machine, network or an external drive.

## How to Export Data or Fit Results

Exporting data is very similar to saving a normal dataset.

If fit results are required to be exported, there is a dedicated menu item entitled **'Export Fit Results'** within the **'File'** menu. The original trace, fitted trace and residuals are all saved.

Standard datasets can be exported to text files, comma delimited or SPECFIT XYZ files as standard.

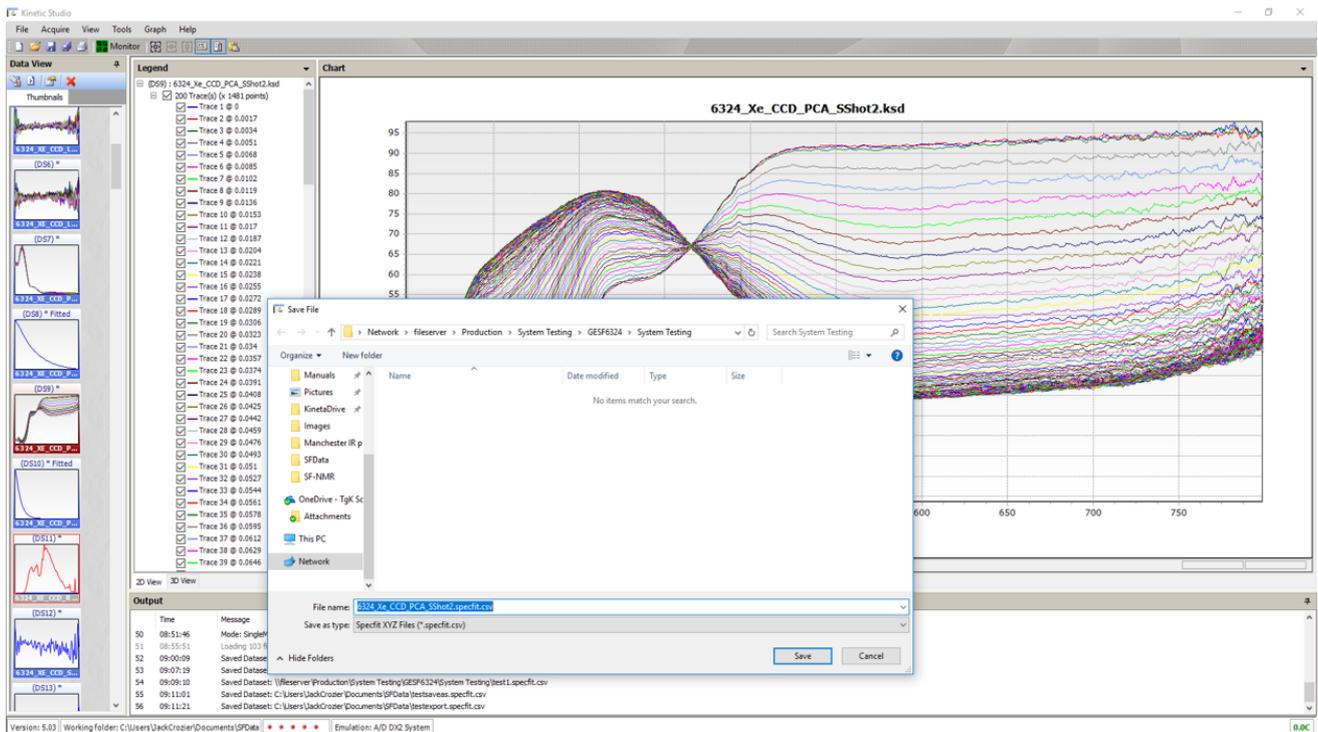


It is important to make sure the correct **'Save as type:'** option is selected. Next, enter the filename and click the **'Save'** button.

## How to Export to SPECFIT/32

Within Kinetic Studio a dataset can be saved as a number of different types. Convenient support for comma delimited SPECFIT/32 files has been included within the software.

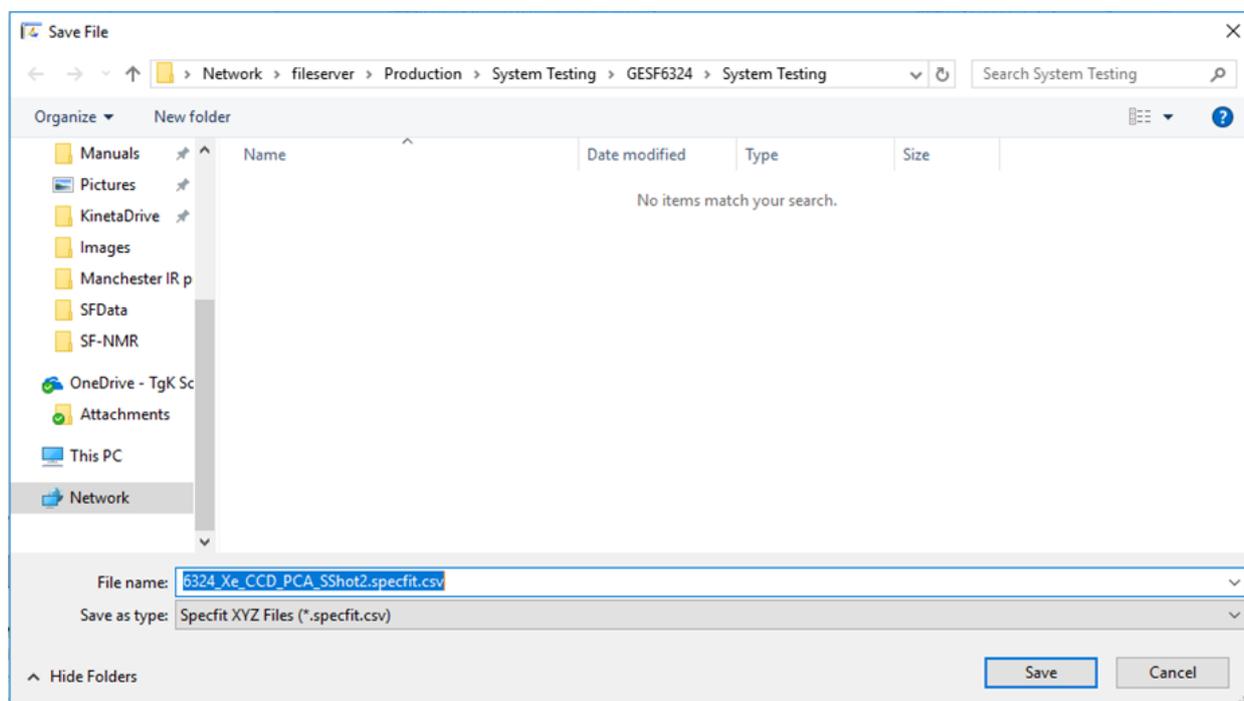
To Export a dataset ready for SPECFIT/32, go to the **'File'** menu and select **'Save As...'**.



The screenshot displays the Kinetic Studio interface. The main window shows a plot titled "6324\_Xe\_CCD\_PCA\_SShot2.ksd" with multiple colored traces. A "Save File" dialog box is open, showing the file path "Network > fileserver > Production > System Testing > GESF6324 > System Testing". The file name is "6324\_Xe\_CCD\_PCA\_SShot2.specfit.csv" and the save type is "Specfit XYZ Files (\*.specfit.csv)". The dialog box also shows a list of folders and files in the current directory.

The interface includes a menu bar (File, Acquire, View, Tools, Graph, Help), a toolbar, a Data View panel on the left with thumbnails, a Legend panel with a list of traces (Trace 1 to Trace 39), a Chart panel with the main plot, and an Output panel at the bottom showing a log of messages.

Within the dialog change the 'Save as type' to 'Specfit XYZ Files'.



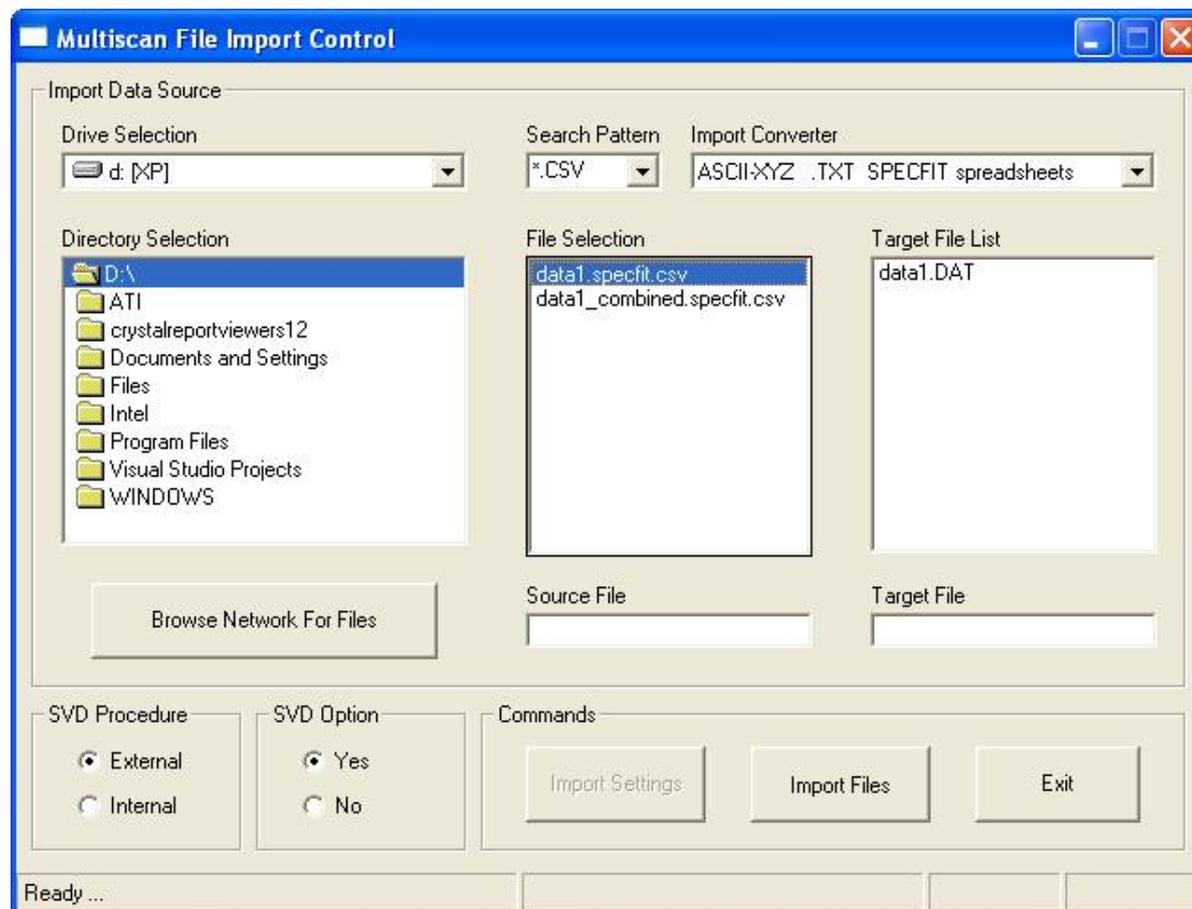
Finally set the 'File name' and click '**Save**'.

Within SPECFIT/32 there are two ways of loading the file that has just been saved. Both methods will be outlined below.

## Importing into SPECFIT/32 Method 1

SPECFIT/32 features an import system which can be used to quickly and easily load the data exported from Kinetic Studio.

To access this system, go to the **{Import}** menu and select **{3D Kinetics Files}**. This will display the following screen:



Navigate to the folder containing the exported Kinetic Studio SPECFIT XYZ file using a '\*.CSV' search pattern as above. Make sure the Import Converter is set to:

'ASCII-XYZ .TXT SPECFIT spreadsheets'

Press **'Import Files'** to begin the import process.

The following dialog will be displayed:

**Multiscan Kinetics File Information**

**Sample Preparation**

**Reactant A**

[A]<sub>o</sub> in flask (M) 1.000E+00  
 Dilution factor, A 2.0

**Reactant B**

[B]<sub>o</sub> in flask (M) 0.00  
 Dilution factor, B 2.0

**Reactant C**

[C]<sub>o</sub> in flask (M) 0.00  
 Dilution factor, C 1.0

Acetone

Thermostat (C) 25.00

Density 1 (g/mL) 0.784400  
 Density 2 (g/mL) 0.801180

Apply Density Corrections  
 Apply Reactant Dilutions

**Concentrations**

[A]<sub>o</sub> in cell (M) 1.000E+00  
 [B]<sub>o</sub> in cell (M) 0.00  
 [C]<sub>o</sub> in cell (M) 0.00

**Wavelength Limits**

Minimum (nm) 280.82  
 Maximum (nm) 700.04  
 Import steps 1

**Conditions**

Cell path length (cm) 1.000  
 Cell temperature (C) 10.00  
 Cell temperature (K) 283.15

**Scan Limits**

First time point (sec) 0.0050  
 Last time point (sec) 1.01  
 Import steps 1

DX2 Kinetics File: data1.ksd | 21/11/2008 15:29:05

**Timebase Options**

Set first time point to zero  
 Logarithmic compression  
 Compress file x 2

1 2 3 4  
 Log scale (decades)

**File Save Options**

Average as known spectrum  
 Over write any previous files  
 Use same settings for all files

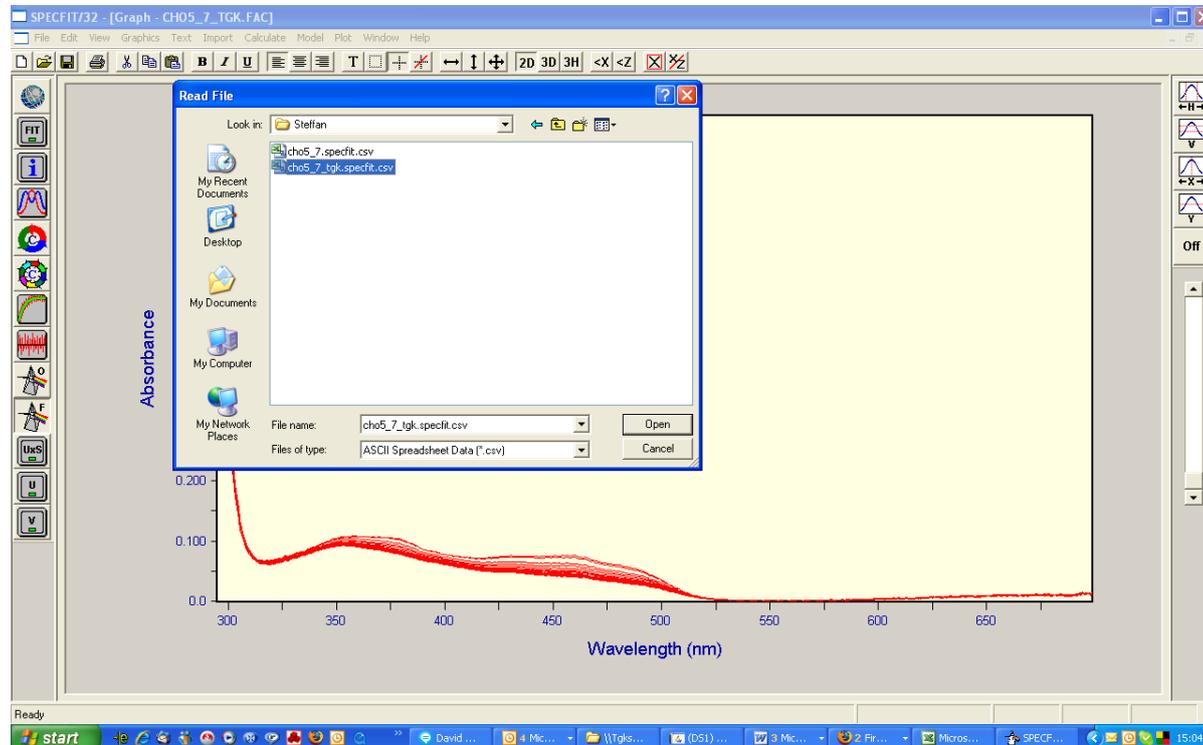
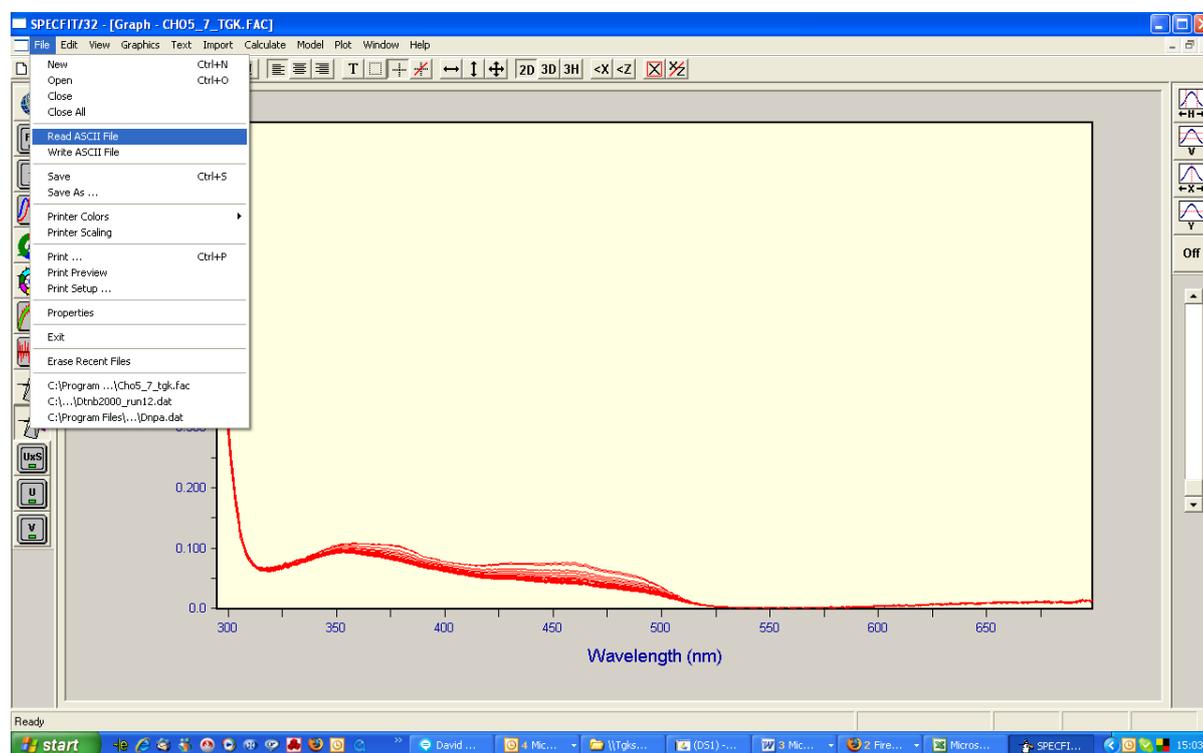
**Commands**

OK Cancel

Please review the parameters on this form. If Diode Array data is present, check the Wavelength Limits in the event the data should be windowed / cropped.

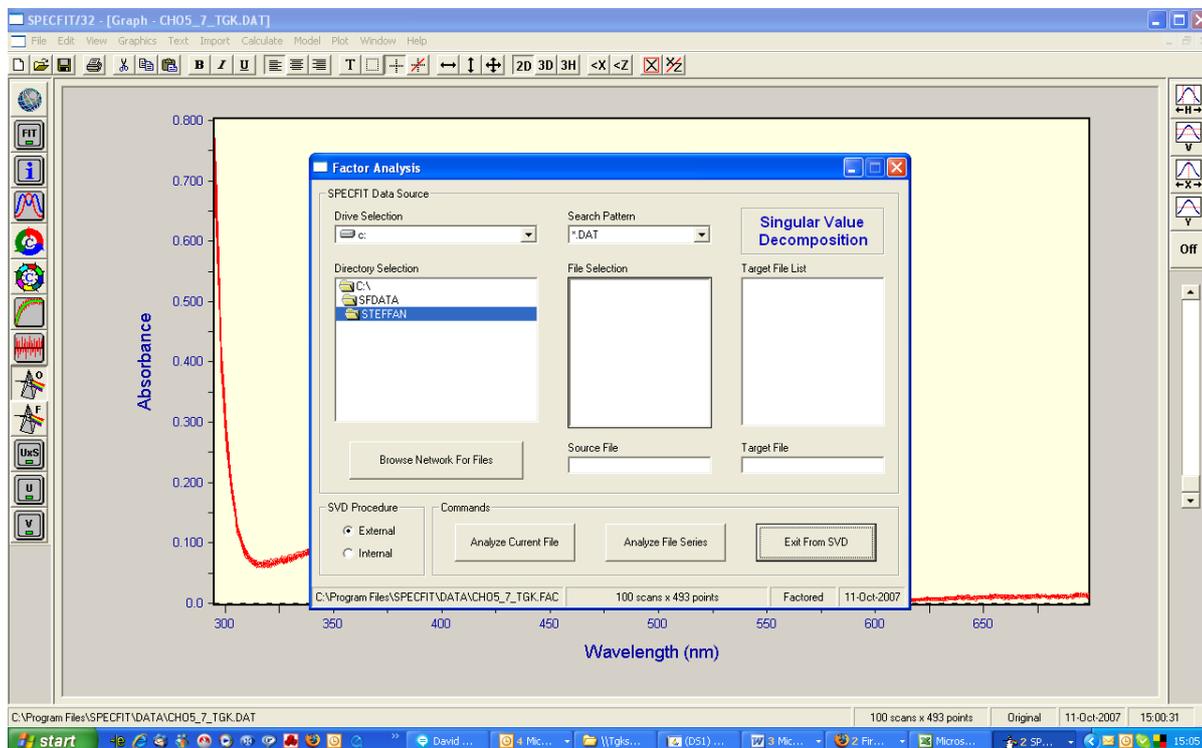
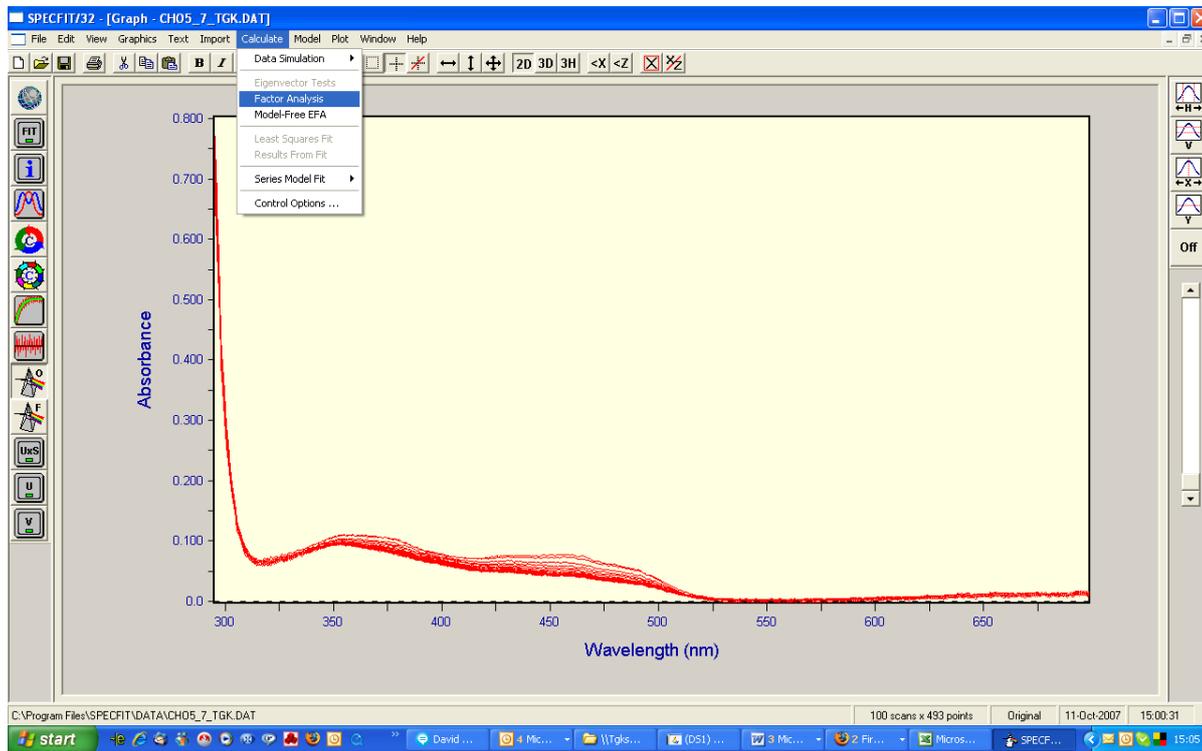
## Importing into SPECFIT/32 Method 2

Within SPECFIT/32, go to the **'File'** menu and select **'Read ASCII File'**.



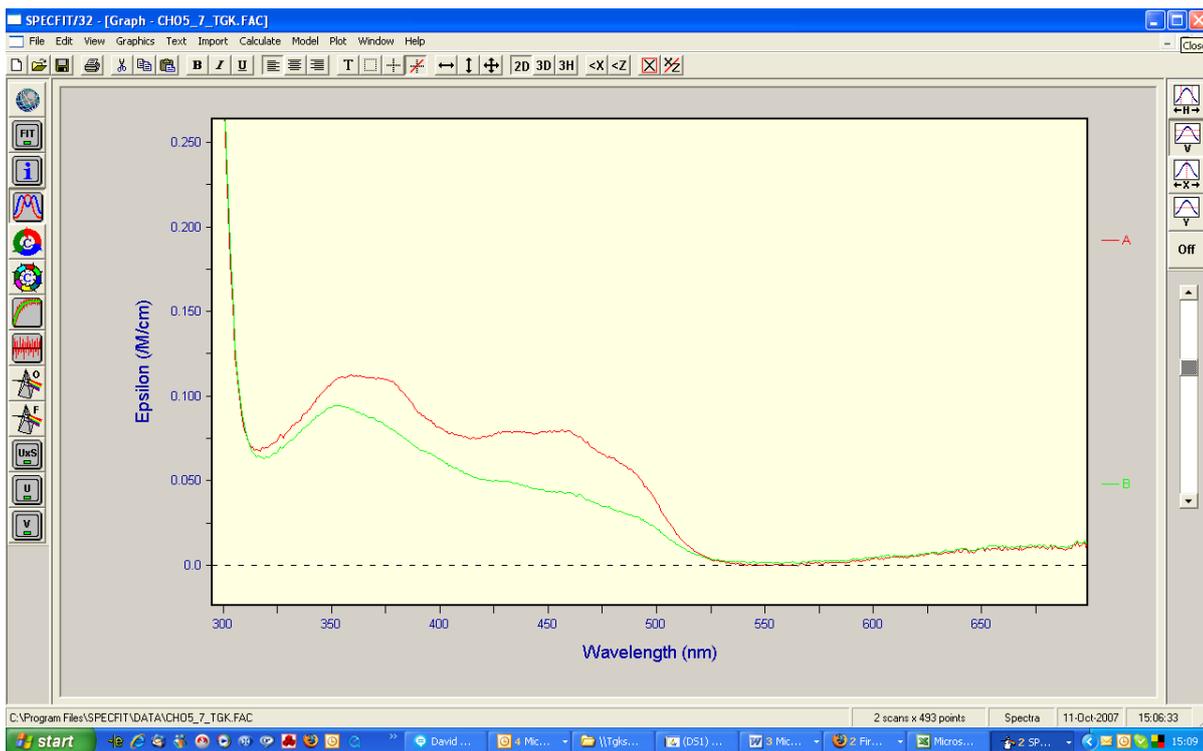
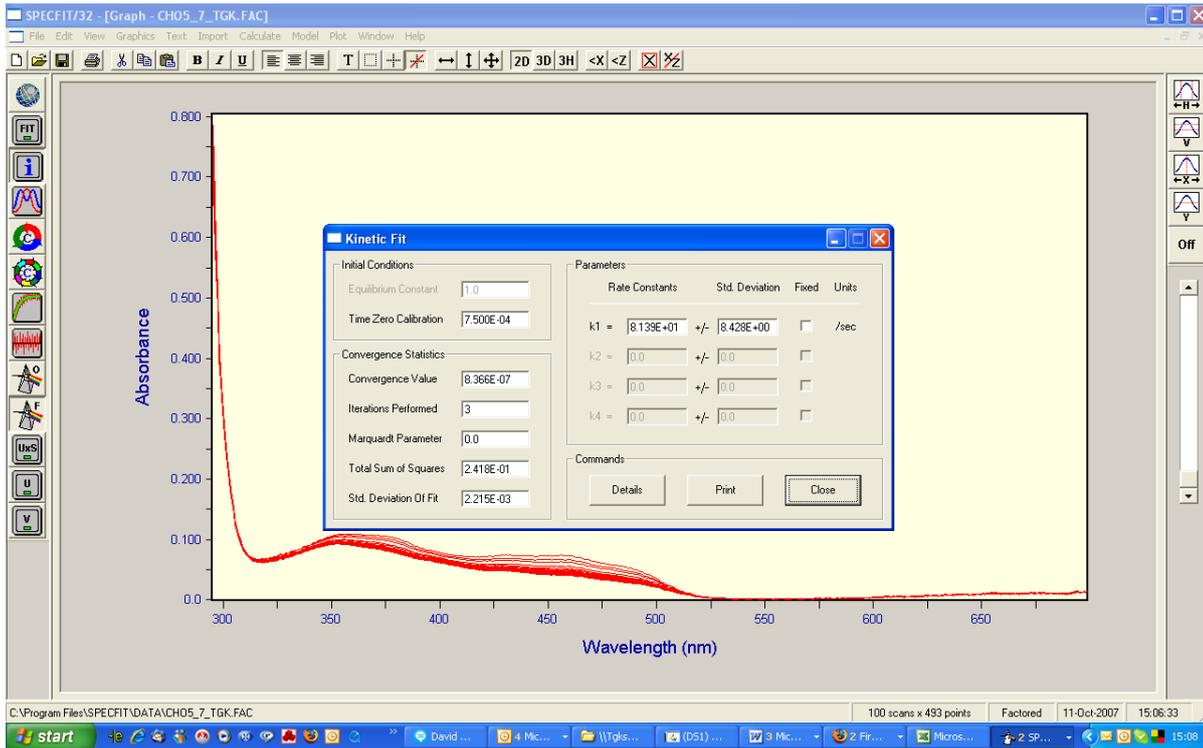
Browse to the exported Kinetic Studio SPECFIT XYZ data file and press **'Open'**.

The data file should now be loaded into SPECFIT/32. The next step typically involves Factor Analysis. To do this, select the **'Factor Analysis'** item from the **'Calculate'** menu.

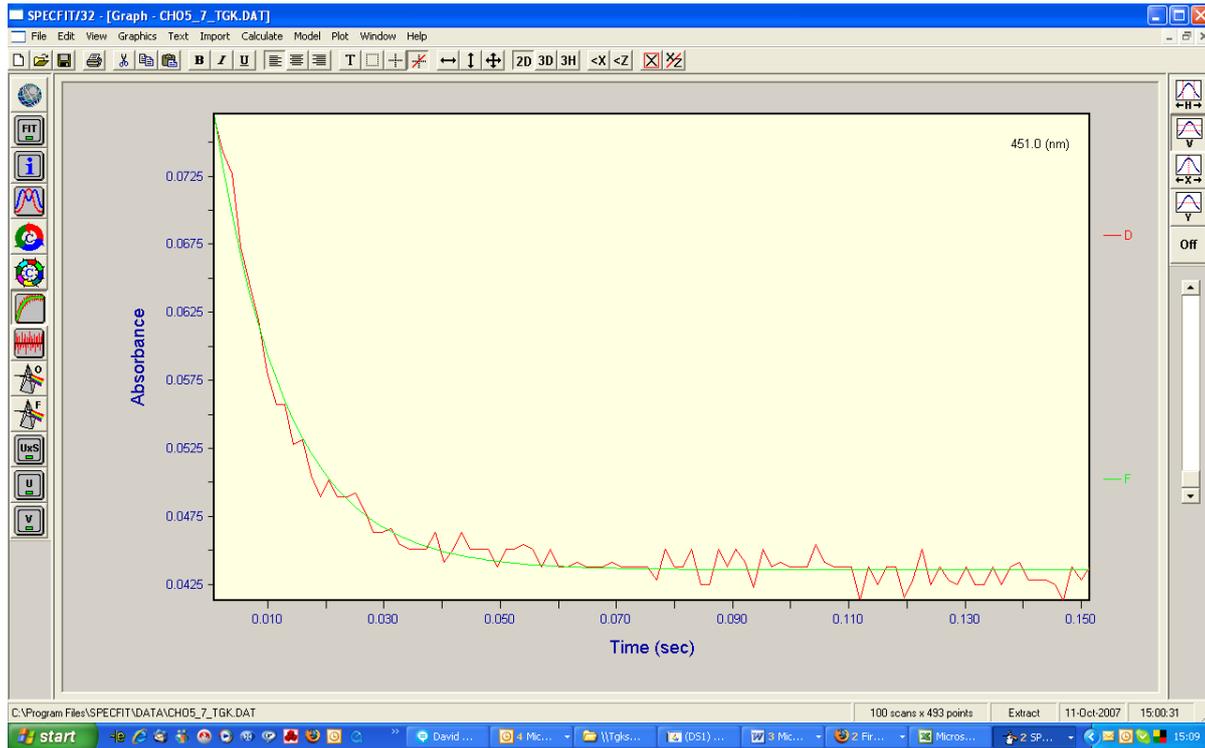


Select **'Analyse Current File'**.

Proceed to select Kinetic Fit from under the Model menu item.

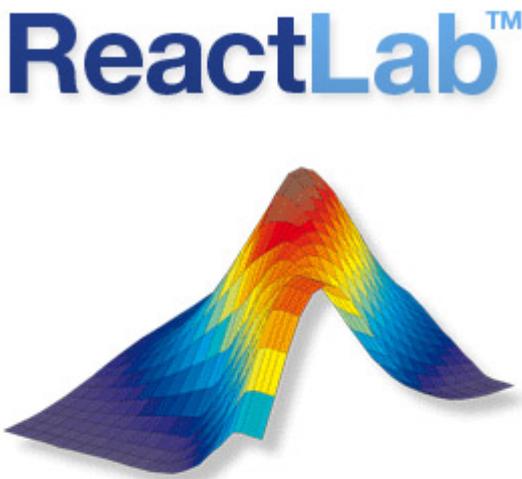


# Kinetic Studio Overview



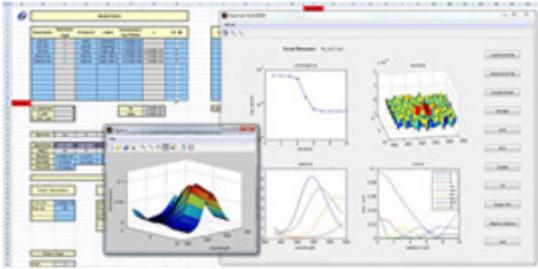
For additional help with SPECFIT/32 please consult the SPECFIT manual.

## ReactLab



**ReactLab™**

*adding new dimensions  
to chemical process  
analysis*



Multivariate Analysis and Reaction Modeling of Chemical Processes from Jplus Consulting.

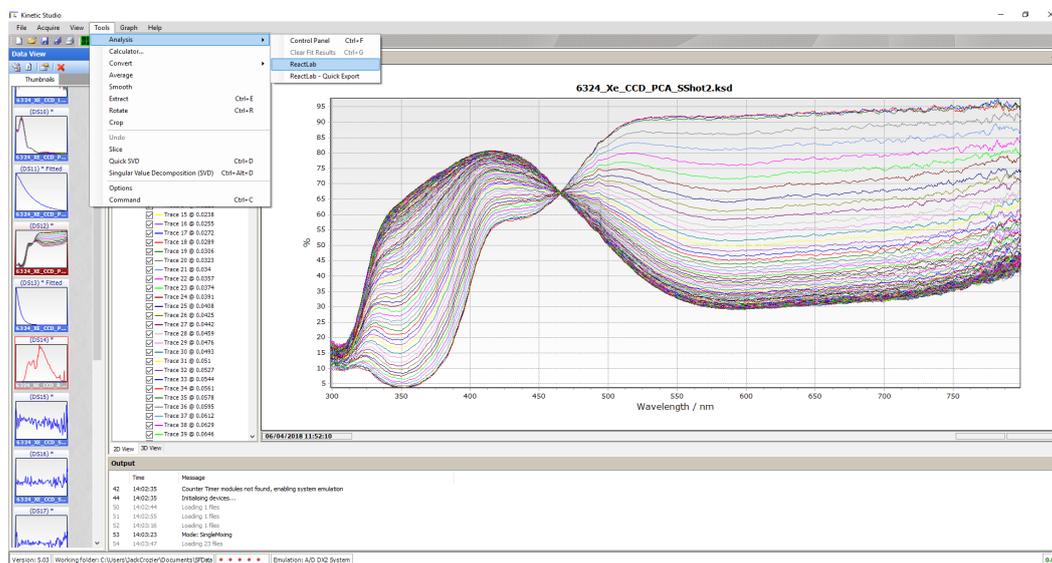
Web site: [www.jplusconsulting.com/](http://www.jplusconsulting.com/)

The ReactLab family of products currently comprise two software applications for the modeling and analysis of multivariate spectrophotometric chemical process data. These Windows applications enable the global fitting of such data to chemical reaction schemes in order to establish and quantitatively analyse the underlying reaction mechanism yielding all the reaction rate constants and equilibrium constants as well as the concentration profiles and spectra of all intermediate species.

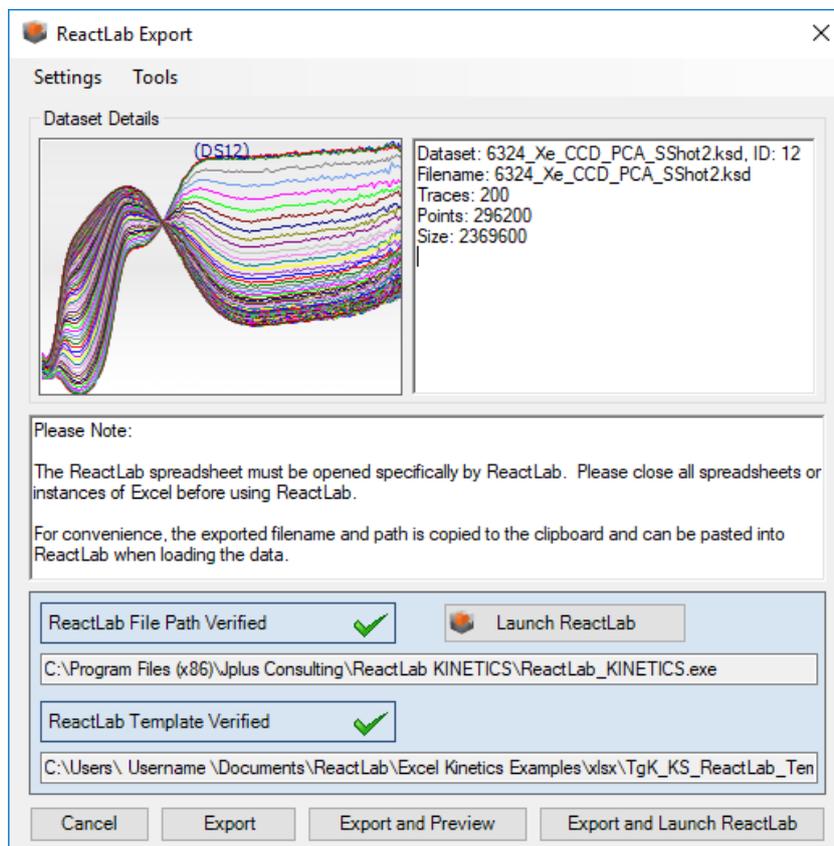
Convenient support for export to ReactLab™ KINETICS software has been included within the software.

## Exporting from Kinetic Studio into ReactLab

With the dataset to be exported open, select the **'Tools'** menu from the top of the screen. Next choose **'Analysis'->'ReactLab'**.



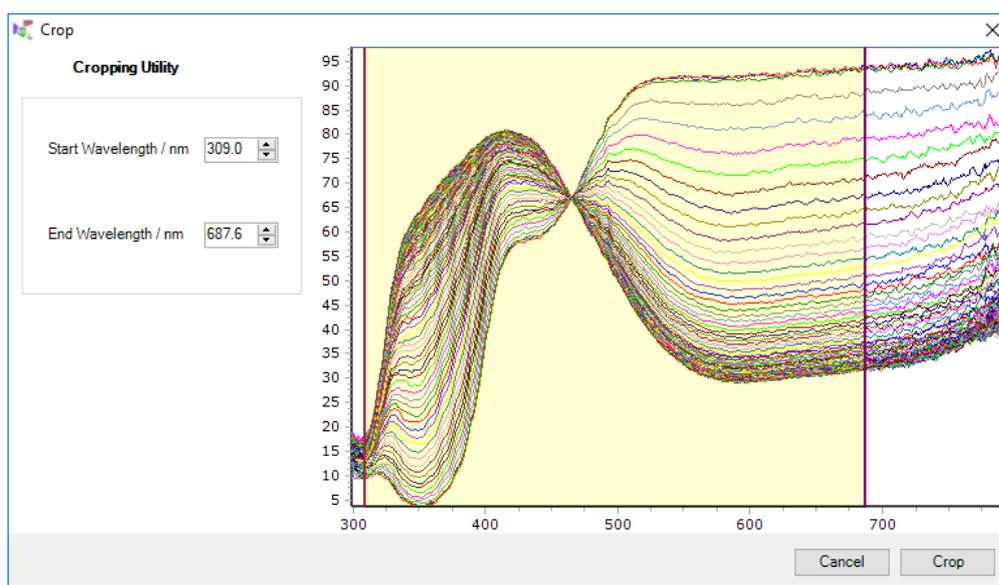
This will open the 'ReactLab Export' window. If ReactLab has previously been configured, the **'ReactLab - Quick Export'** facility can be used to speed up the export process of multiple datasets.



The first time the export to ReactLab facility is used, it must be configured with the appropriate paths. This tells Kinetic Studio where the ReactLab program resides and which ReactLab template should be used for data export.

To change the paths, either select the appropriate menu item from the '**Settings**' menu or press the file path and template verification buttons. This will present a file browser dialog to locate either the ReactLab executable and / or the ReactLab Excel template that should be used for data export.

Prior to exporting the data, the dataset can be cropped if necessary using the '**Crop**' option under '**Tools**'.



Once the dataset is ready for export, simply press one of the [Export] buttons.

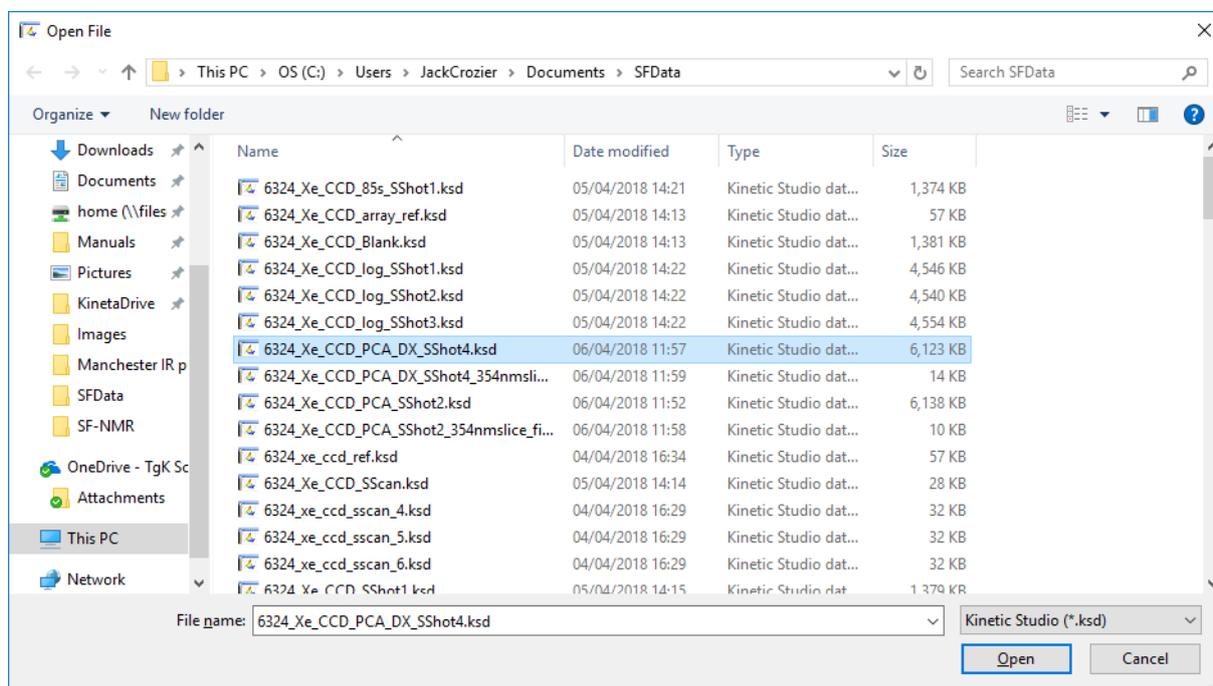
The data will be exported into the Excel ReactLab template and saved to a new location chosen via a standard Save File dialog.

For convenience the filepath is copied to the Windows Clipboard for quick pasting into the ReactLab open file facility.

Tip: Pressing the key combination CTRL-V simultaneously will paste items from the clipboard.

## How to Load a Dataset

Opening data items can be achieved by either using the **'Open...'** option from the **'File'** menu, or by using the toolbar button. This will produce the following dialog:

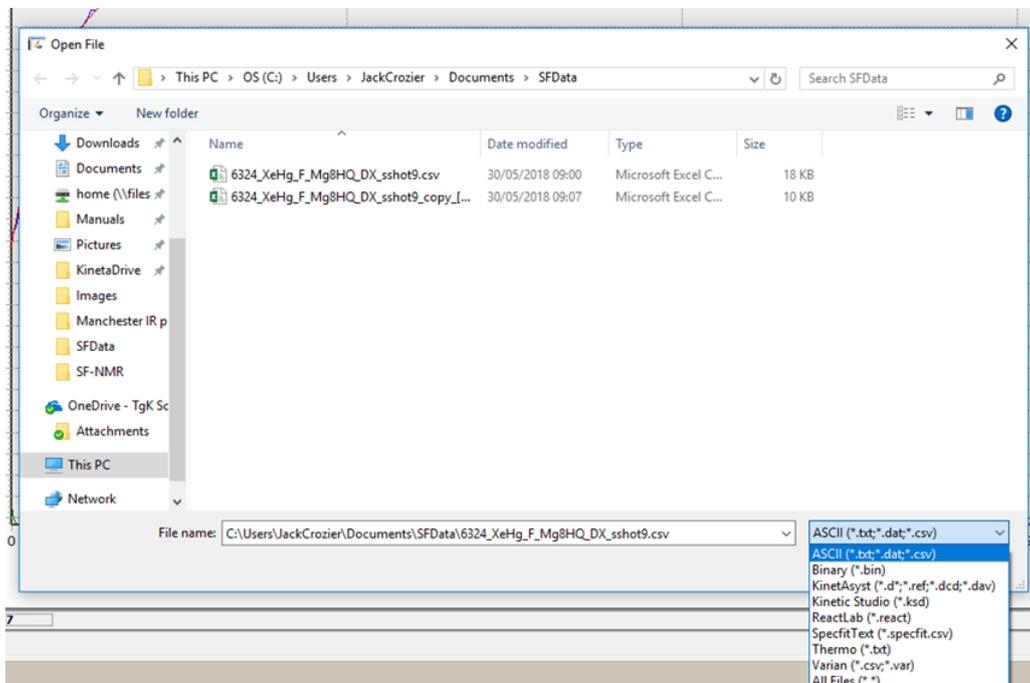


This will initially display the current working folder. Navigate to the drive and folder where the data is located, and select one or more files from the list.

## How to Import Data

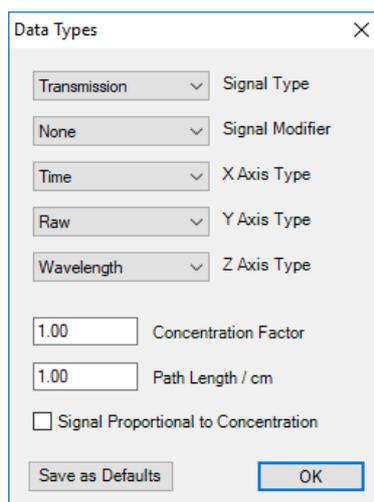
Importing data is very similar to loading a standard dataset. ,

Kinetic Studio can import text files, comma delimited files and older KinetAsyst files.



Choose **'Open'**, as above. Change the **'Files of type:'** drop down list to the file type of choice and select a file or files from the files list.

Once data has been selected, the Data Types dialog appears. Choose the relevant Signal and Axis Types and press **'OK'**.



If Kinetic Studio successfully imports the data, it will appear within the thumbnail panel and automatically display the data in the main graph view.

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# Stopped-Flow; Initial Setup and Getting Started



Before beginning to collect data, it is necessary (as with any spectrophotometer), to set up the optical signals so that the analogue signals fit within the operating range of the A/D converter and so that measured signals can be interpreted (where appropriate), in absorbance units.

These guidelines to setting up for optical measurements can generally be applied across the range of techniques supported by Kinetic Studio (i.e. Single and Double Mixing Stopped-flow, Temperature-Jump).

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# Photomultiplier



## How to perform a Scan Blank (Absorbance)

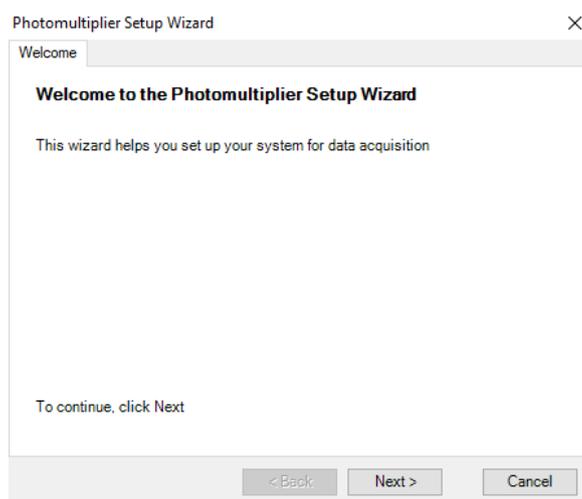
*This mode allows users to set up for absorbance measurements to be taken over a range of wavelengths and are thus able to acquire kinetic data at different wavelengths and/or acquire single scans to characterise reagent spectral information.*

When the system is configured for absorbance measurements, before acquiring new data it is first necessary to align and focus the optics, set the correct photomultiplier voltage and scan a blank. Scanning a blank involves the monochromator scanning a wavelength span acquiring 100% (maximum incident light) and 0% (dark condition) transmission reference levels.

Before performing this operation, ensure that the optical cell contains pure water or a buffer solution.

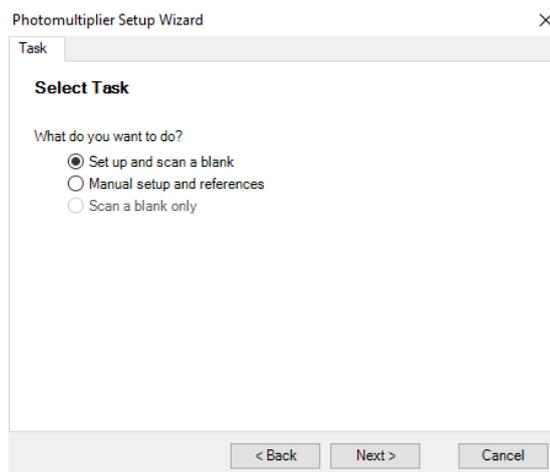
To perform an auto-setup scan blank, perform the following steps:

Press the '**Spectrometer**' button to enter the manual setup mode.  
This will display the '**Photomultiplier Setup Wizard**'.

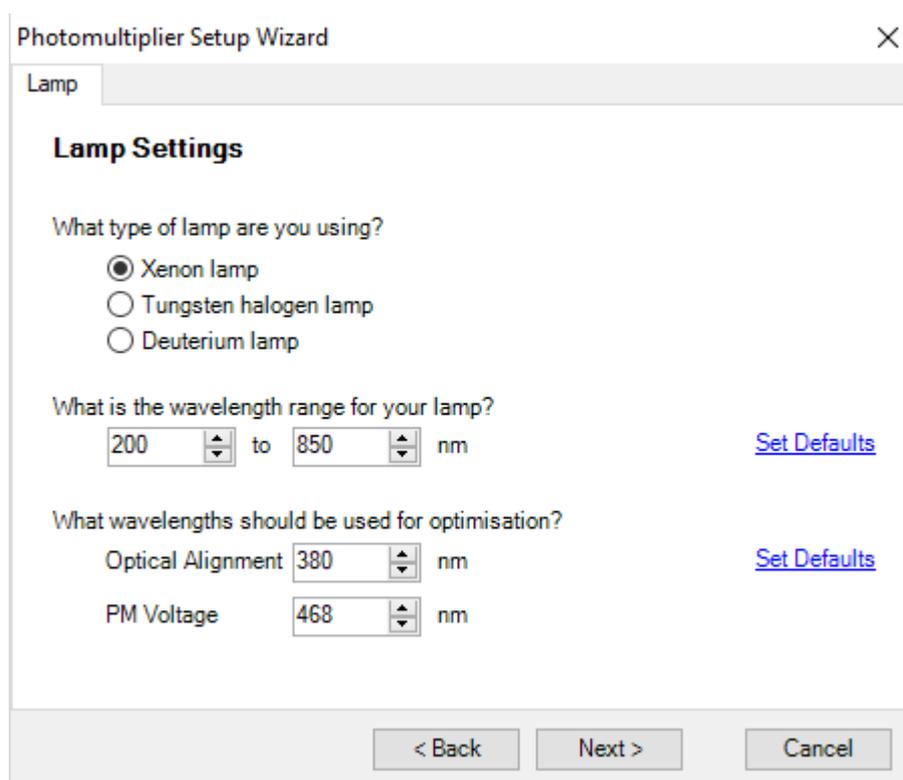


Next the dialogue will offer choice of a scanned set up and a **manual set up**. The former invokes the use of a scanned blank over a user selected wavelength range, the latter provides for users content to set up at just a single wavelength.

A scanned blank is appropriate for absorption studies only; the manual set up can be used for absorption and fluorescence.



Here, we will begin with selecting the **'Set up and scan a blank'** to begin the wizard driven setup. The **'Lamp Settings'** panel enables the user to select the lamp currently within the system. This in turn automatically fills the wizard with typical wavelength ranges, optimisation and alignment wavelengths for the blank.



The first step is to select the lamp currently fitted to the system.

If the typical values for the wavelength range are not suitable or require adjustment, please edit the desired start and end wavelength spans for the lamp.

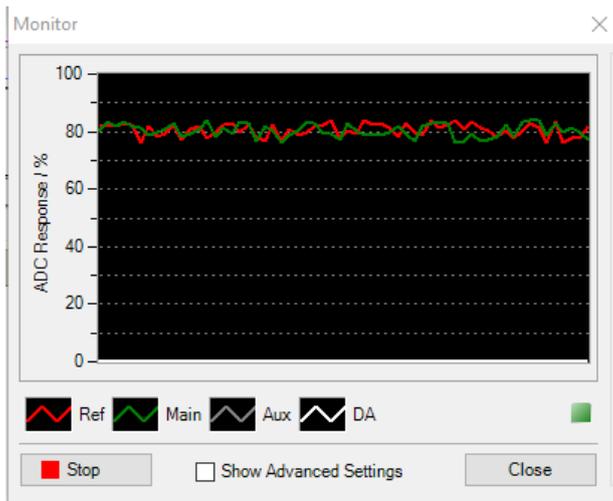
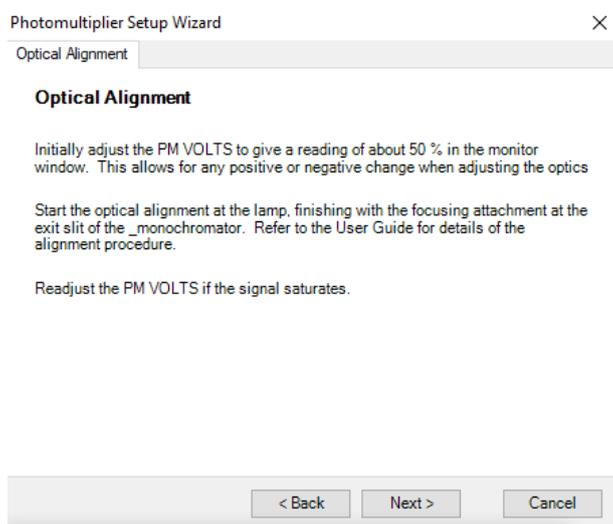
To ensure optimal distribution of the signal over the scan blank wavelength span, the user should optimise the optics at one wavelength, and then maximise the PM volts at another wavelength. This is especially critical when using a wide wavelength span.

To restore the default values for a particular lamp use the '**Set Defaults**' link.

It is often convenient to set the lamp optimisation wavelength to suit the wavelength range where absorbance changes are to be studied.

The edit field labelled '**PM Volts**' is the wavelength where the PM volts are maximised; this ensures that the maximum signal span is achieved under normal circumstances. With the Xenon and QTH lamps, this is set at 480 nm as this is where the system exhibits a maximum signal.

Once the lamp and wavelength ranges have been set, please press the '**Next**' button to proceed with the first stage of optimisation.

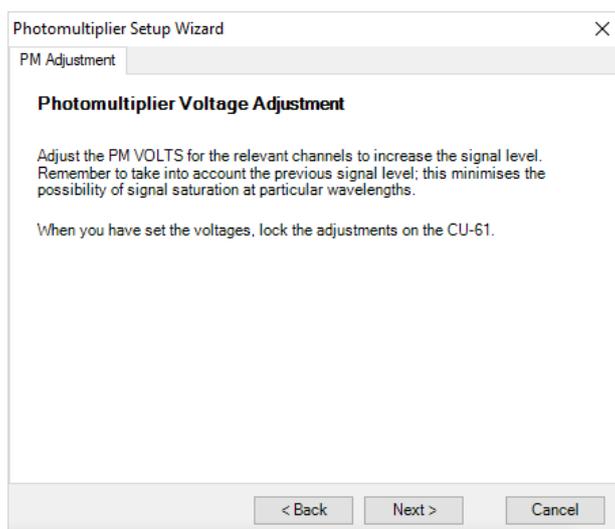


As the '**Optical Alignment**' page appears the monochromator will move to the specified 'Optical Alignment' wavelength as specified on the previous page of the wizard.

To perform the optimisation, open the PM-61s photomultiplier shutters and adjust the PM Volts for the main channel (green trace), the reference channel (red trace) and, if applicable, the auxiliary channel (grey trace), increasing the signal at the Live Display to about 50. This is simply used as a mid-point to allow for positive and negative shifts when the optics are adjusted.

Follow the procedure for optimising the optical components as described in the Section 4 of the SF-61DX2 User Manual. In brief, adjust the lamp position and its alignment. It may be necessary to adjust the PM volts for the particular channel should the signal saturate.

After finishing the optimisations, return the signal level back to 50 on the Live Display, this is simply used as a relative indication of signal level when moving to the next wavelength.



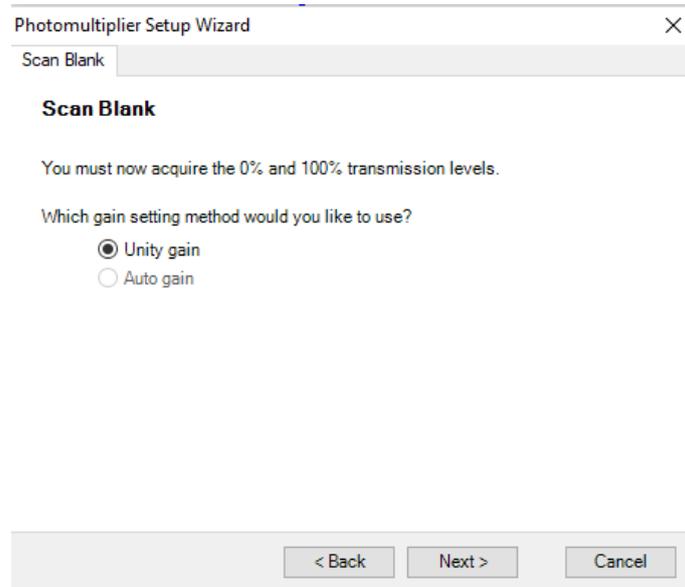
As the '**Photomultiplier Voltage Adjustment**' page appears, the monochromator will move to the adjustment wavelength as specified previously.

Increase the PM Volts for the relevant channels, so setting the signal level(s) to about 80% full scale. You must ensure that the signal does not saturate, i.e. go above 100% full scale.

As a quick check it is worthwhile clicking the '< Back' button to ensure the signals are not saturated at the previous wavelength where the optical optimisation was performed. If they are saturated, go to the next page and lower the PM Volts for the relevant channels.

This concludes the optimisation process. Clicking the 'Next>' button will present the '**Scan Blank**' page.

## Scanning the Blank

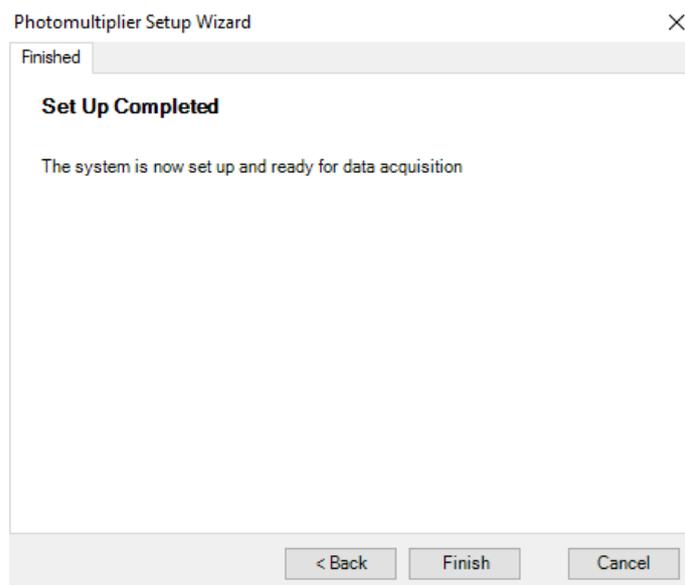


The scan blank process is fully automated.

The process will begin with a full monochromator re-calibration.

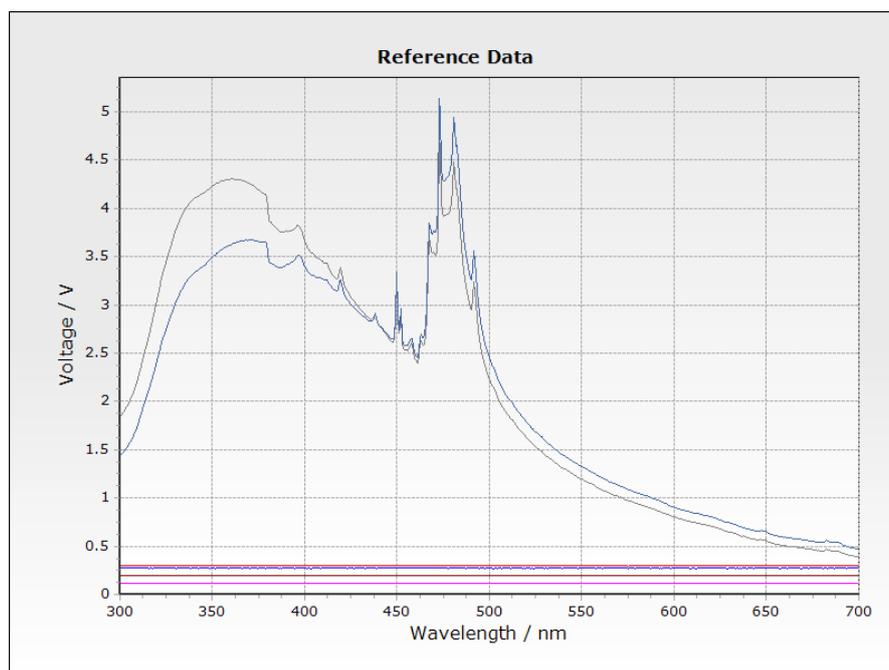
The scan blank process consists of scanning a baseline with no light (0%) and with the filter wheel open (100%). Whilst traversing through the wavelength range, the system will automatically insert appropriate filters.

After both the 0% and 100% scans have completed, the system will acquire static baseline references and present the '**Set Up Completed**' panel.

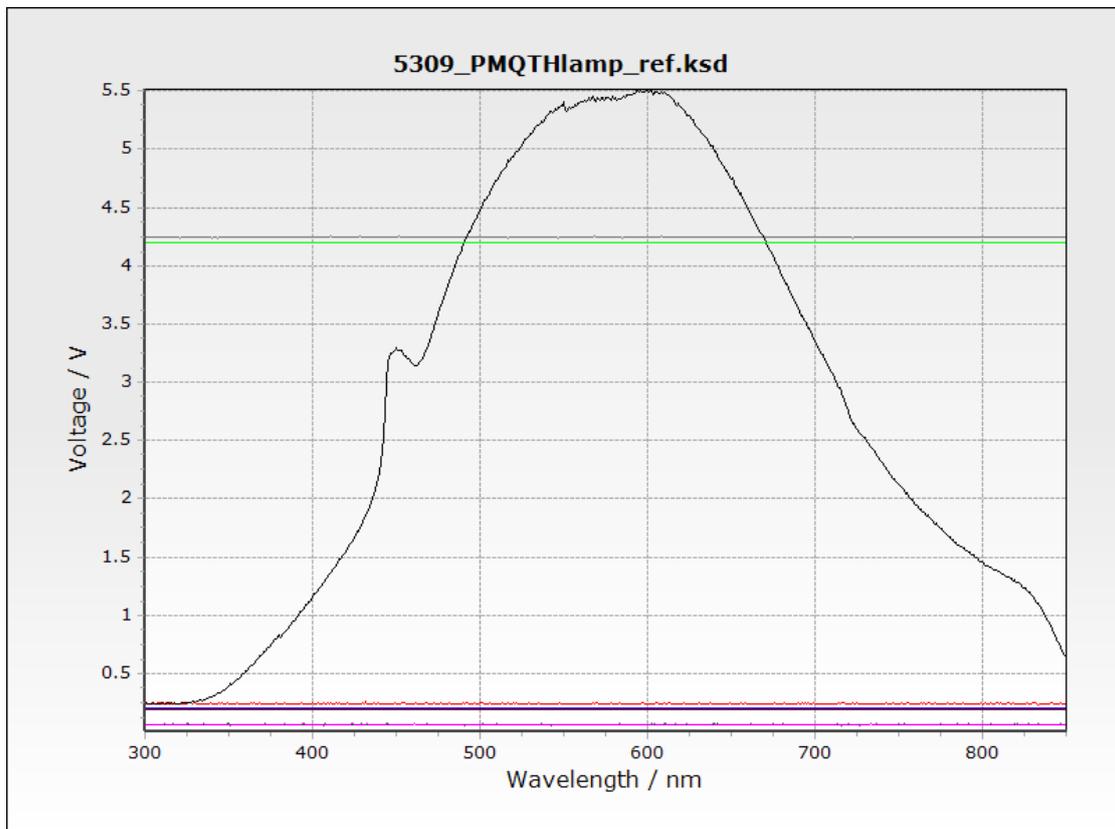
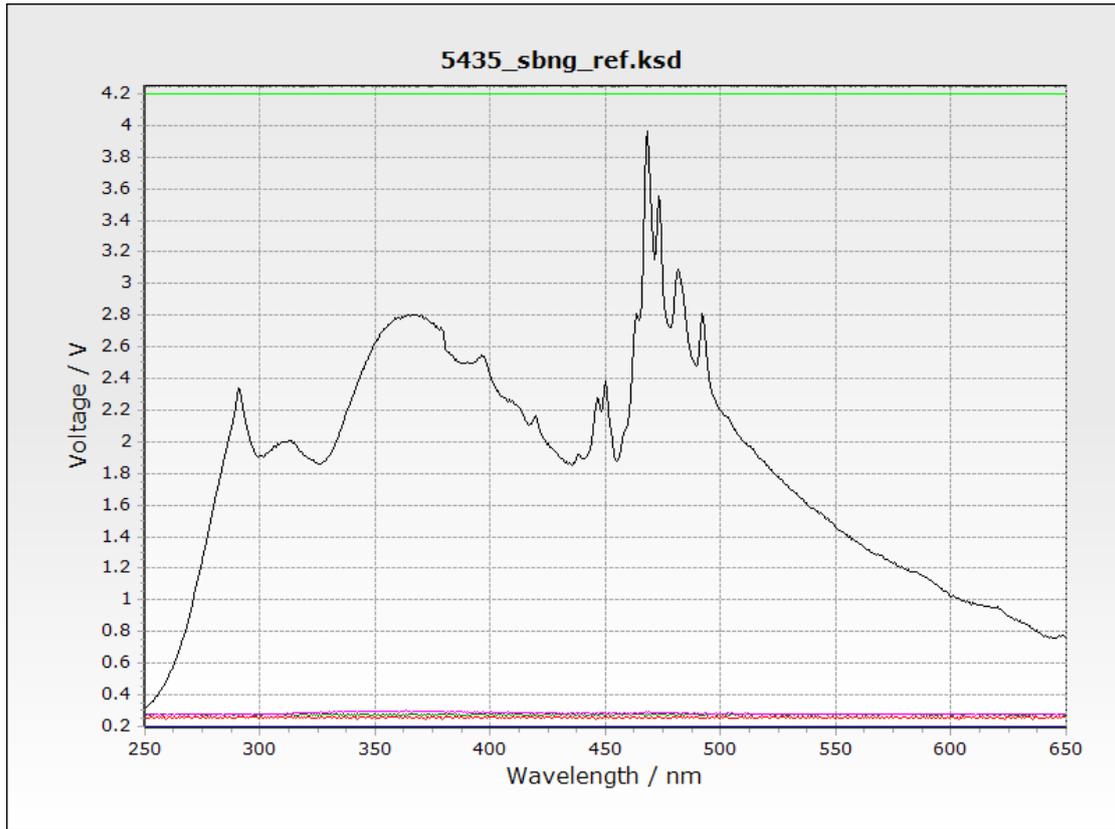


The system is now ready for use.

An example of a scan blank collected with the xenon lamp:



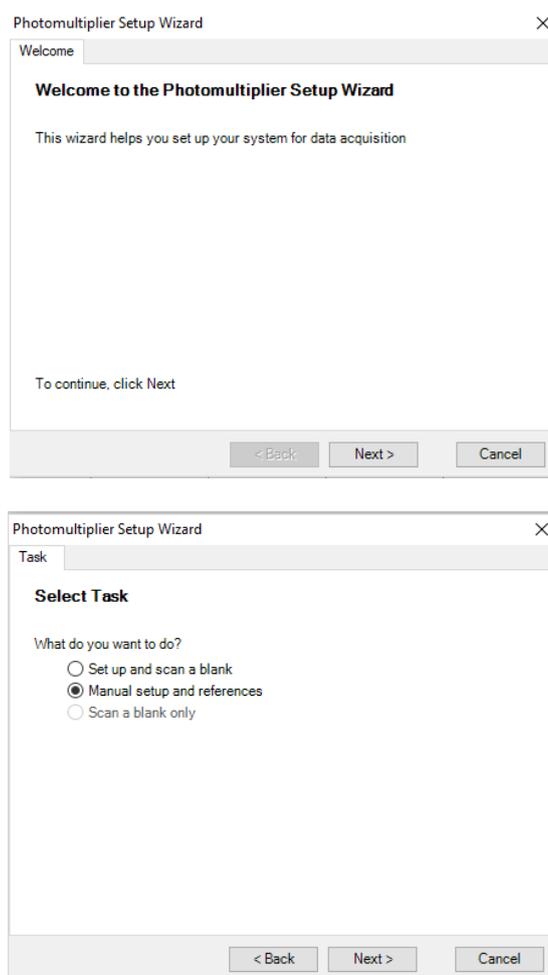
Here are a couple of additional example reference data sets:



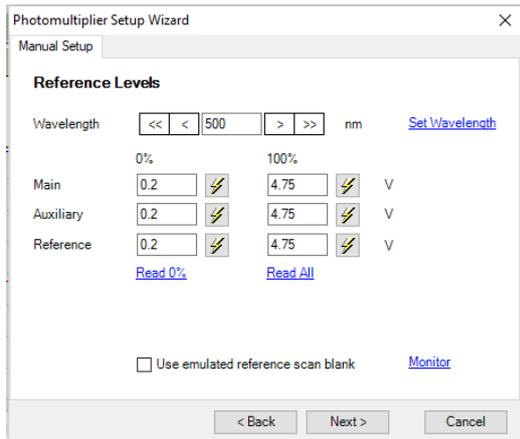
## How to set the Reference / Signal Levels at a Single Wavelength

This mode allows users to **set up for absorbance or fluorescence measurements at a single wavelength**. Typically this would represent **setting the excitation wavelength for fluorescence mode and a simple single wavelength mode for absorbance studies**.

Press the '**Spectrometer**' button to enter the manual setup mode. This will display the '**Photomultiplier Setup Wizard**'.



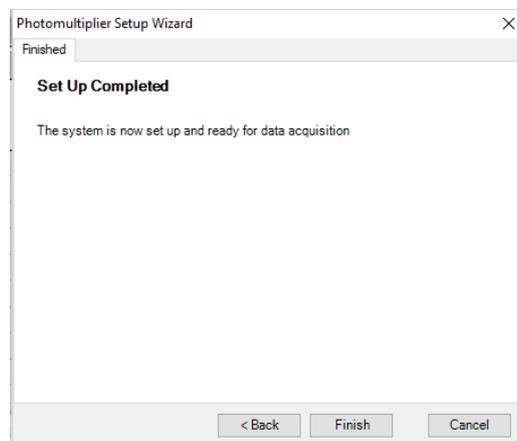
Select the '**Manual setup and references**' to begin a manual setup.



The manual setup panel will be visible along with the live display. This allows the user to change wavelength and read signals for both 0% and 100% for each channel.

When used for absorbance measurements, 100% (maximum incident light) and 0% (dark condition) transmission reference levels are required to be recorded for a single wavelength. When used for fluorescence measurements, this single wavelength is the excitation wavelength and although reference levels can be acquired, normally the Live Display is used simply to maximise the fluorescence signal by optimising the optics and maximising the PM Volts at this wavelength.

Simply exiting the setup dialog by pressing 'Next' and 'Finish' enables data collection at the set wavelengths. Often fluorescence measurements are made without any set references.



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## General usage instructions for manual setup

1. Enter the monochromator wavelength into the '**Wavelength**' box. This is then set and recorded for each subsequent shot.

Note: Ensure the correct optical filters are in place if required (eg fluorescence emission).

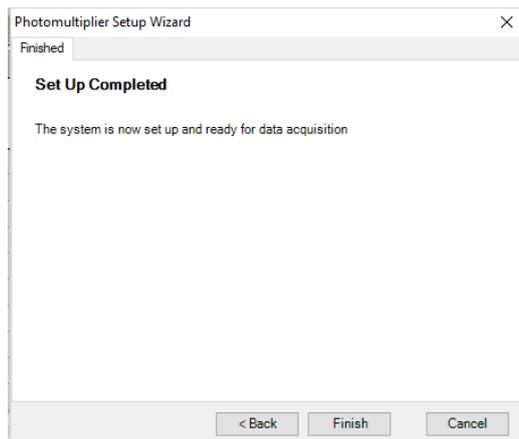
2. The Monitor or live display accompanies the manual setup dialog in order to view the signal levels for each of the enabled detector channels.

## Manual Setup for Absorbance

1. Ensure the flow circuit and in particular the observation cell is flushed and as such contains water or buffer solution.
2. Optimise the optics for maximum signal.
3. Increase the PM Voltage for the relevant channels until the signal trace approaches 80% span. Remember for older instruments, an increase in light intensity manifests itself as a decrease in signal level. Note that the Main and Reference Channels must be set.
4. Collect the 0% reference data by either pressing the button(s) next to each channel in the 0% column or enter them numerically, or press the '**Read 0%**' link to collect for both channels. Remember to shutter the light manually for 0% reference levels.- only older stuff.
5. Collect the 100% reference data by either pressing the button(s) next to each channel in the 100% column or enter them numerically, or press the '**Read 100%**' link to collect for both channels. Remember to open the shutter to light manually for 100% reference levels.

Note: As mentioned older instruments may have inverted voltage levels displaying a high reading for 0% and a low reading for 100%.

6. Once the values have been read, press the '**Next**' button to complete the manual reference wizard. Then click '**Finish**' to return to the main control panel.



### Manual Setup for Fluorescence

The user will need to have prior knowledge of the expected fluorescence change, whether it is increasing or decreasing with time.

#### If the fluorescence is increasing

1. Load both reagents into the Sample Handling Unit and mix by doing a few shots. After an appropriate time, the contents of the optical cell will now give a fluorescent signal comparable with the maximum fluorescence at the end of the reaction.
2. With the appropriate filter(s) fitted to the photomultiplier(s), open the shutter(s) and apply PM Volts until the signal level responds.
3. Optimise the optics for maximum signal. The user should note a few points here: the fluorescence may decrease due to photo bleaching – replenishing the cell contents periodically will offset this problem, however, the timescale of the bleaching process is probably an unknown factor here. Secondly, the excitation wavelength can be adjusted to find maximum response, especially when using line sources such as mercury lamps.
4. Increase the PM Volts to set the signal level close to an 80% span. Remember for older instruments, an increase in light intensity manifests itself as a decrease in signal level. Note: Reference channel must be set – usually in Single Beam mode for fluorescence and will appear at just above 80%.
5. If desired, the user can record the fluorescence reaction 0% and 100% levels, by first selecting the '**Read 0%**' button and recording the 0% level, remembering to shutter the light and then selecting the '**Read 100%**' button and recording the 100% level ensuring any light shuttering used is open.

If required, the dialog can be closed to enable data acquisition by selecting the **'Next'** and then **'Finish'** buttons to exit.

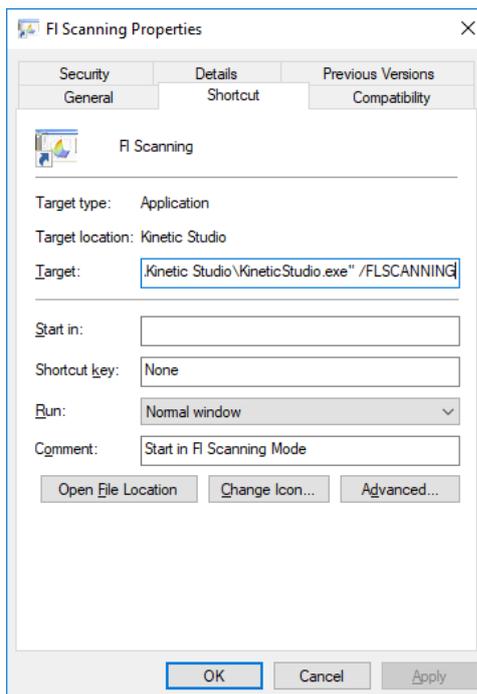
### **If the fluorescence is decreasing**

1. Load the fluorescent reagent into the Sample Handling Unit and mix it with buffer by doing a few shots. The contents of the optical cell will now give a fluorescent signal comparable with the initial fluorescence of the reaction.
2. With the appropriate filter(s) fitted to the photomultiplier(s), open the shutter(s) and apply PM Volts until the signal level responds.
3. Optimise the optics for maximum signal. The user should note a few points here: the fluorescence might well decrease due to photo bleaching – replenishing the cell contents periodically will offset this problem, however, the timescale of the bleaching process is probably an unknown factor here. Secondly, the excitation wavelength can be adjusted to find maximum response, especially when using line sources such as mercury lamps.
4. Increase the PM Volts to set the signal level close to an 80% span. Remember for older instruments, an increase in light intensity manifests itself as a decrease in signal level.
5. If desired, the user can record the fluorescence reaction 0% and 100% levels, by first selecting the **'Read 0%'** button and recording the 0% level, remembering to shutter the light and then selecting the **'Read 100%'** button and recording the 100% level ensuring any light shuttering used is open.

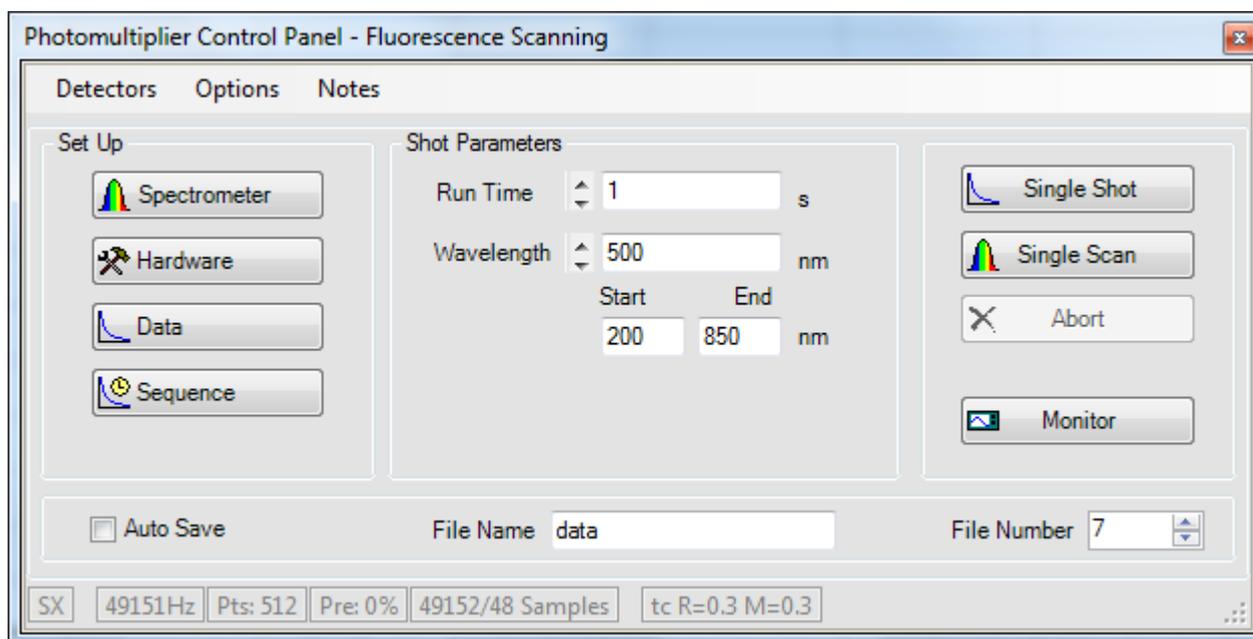
If required, the dialog can be closed to enable data acquisition by selecting the **'Next'** and then **'Finish'** buttons to exit.

## Fluorescence Excitation Scanning

Kinetic Studio should be started in fluorescence scanning mode. Normally this is done by clicking on the 'Fluorescence Scanning' icon installed along side the standard Kinetic Studio icons. If this is not visible, please either re-install Kinetic Studio placing a tick in the 'Fluorescence Scanning' option or manually create an additional icon and add a command line entry of /flscanning.



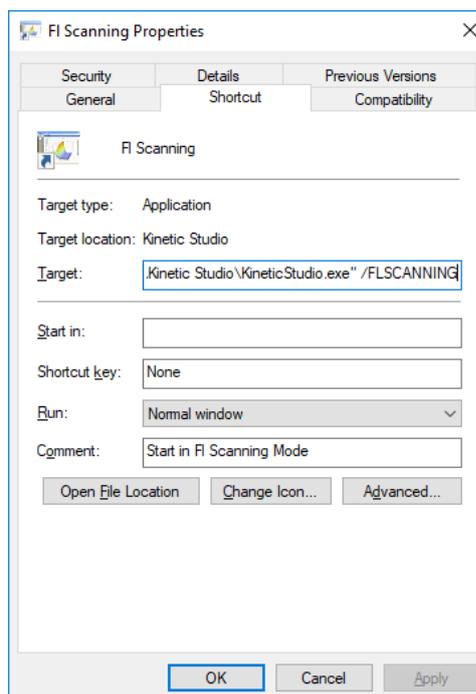
When enabled, the control panel will indicate it's running in 'Fluorescence Scanning' mode.



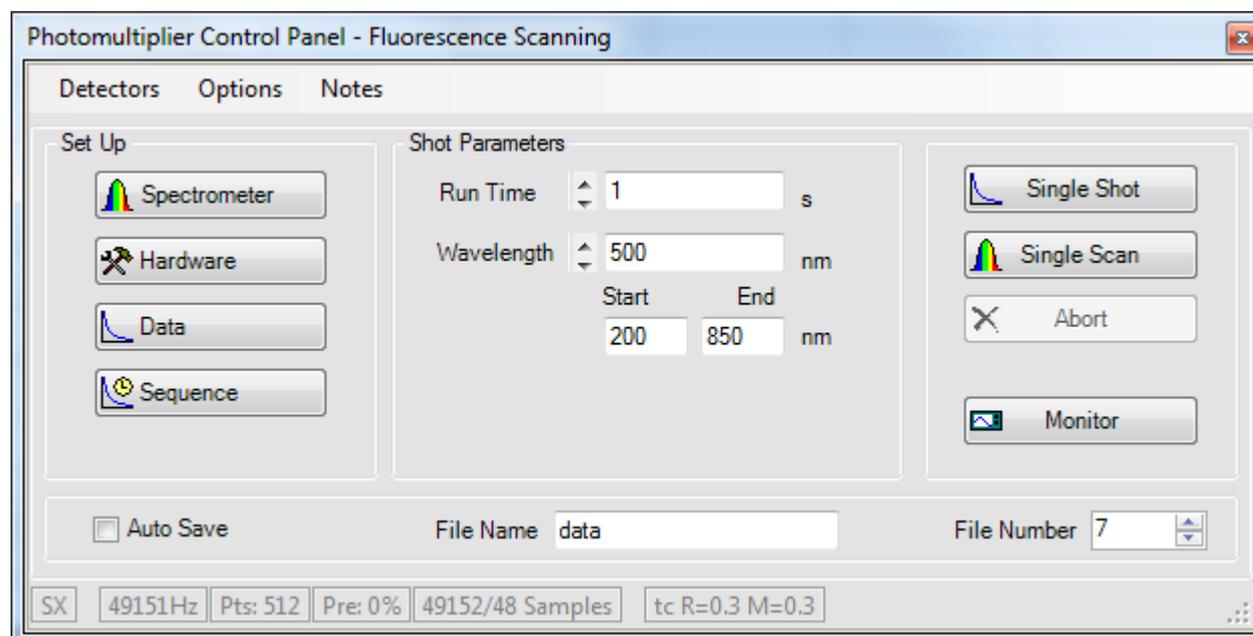
The control panel is ready to be used. A wavelength can be entered into the 'Wavelength' field. Alternatively if zero order is required, please enter 0.

## Fluorescence Emission Scanning

Kinetic Studio should be started in fluorescence scanning mode. Normally this is done by clicking on the 'Fluorescence Scanning' icon installed along side the standard Kinetic Studio icons. If this is not visible, please either re-install Kinetic Studio placing a tick in the 'Fluorescence Scanning' option or manually create an additional icon and add a command line entry of /flscanning.



When enabled, the control panel will indicate it's running in 'Fluorescence Scanning' mode.



The control panel is ready to be used. A wavelength can be entered into the 'Wavelength' field. Alternatively if zero order is required, please enter 0.

## Setting the Data Type and Dataset Parameters

Note the default data type is Transmission on Channel 1. To change this or set up for multi-channel acquisition, the user should configure which channels should be enabled and set the data type.

Configuring the correct data type is essential as this dictates some of the data processing immediately after a shot and indeed how the data can be subsequently manipulated.

The '**Data Type**' options are:

- Unknown
- Transmission
- **Fluorescence**
- Light Scatter
- Conductivity

The '**Modifiers**' available are:

- Unknown
- **None**
- Polarisation
- Anisotropic Polarisation

The screenshot shows a 'Data Settings' dialog box with the following configuration:

- Channel 1 (Main):** Enabled . Data Type: Transmission. Data Type Modifier: None.
- Channel 2 (Aux):** Enabled . Data Type: Fluorescence. Data Type Modifier: None.
- Dataset & Shot Settings:** Data Points: 512. Oversamples: 48. % Pretrigger: 0.
- Save 'Data Type' settings on exit.
- Buttons: OK, Cancel.

Each channel can be enabled by checking or unchecking the '**Enabled**' checkbox. (Note that this function mirrors the channel select boxes on the main control panel).

The number of data points can be increased or decreased resulting in more or less detailed datasets. Typically, the default setting of 512 is recommended and widely used – users are encouraged to keep this default setting until they are confident that they understand the consequences of the more advanced settings.

Oversampling helps to increase the quality of data and resolution. It has the ability to increase signal to noise by collecting number of data points and averaging them together. Typically this is

set at 48 but it can be increased or decreased depending on the experiment. For T-Jump acquisitions speed tends to be priority so often the oversamples may be set to 1.

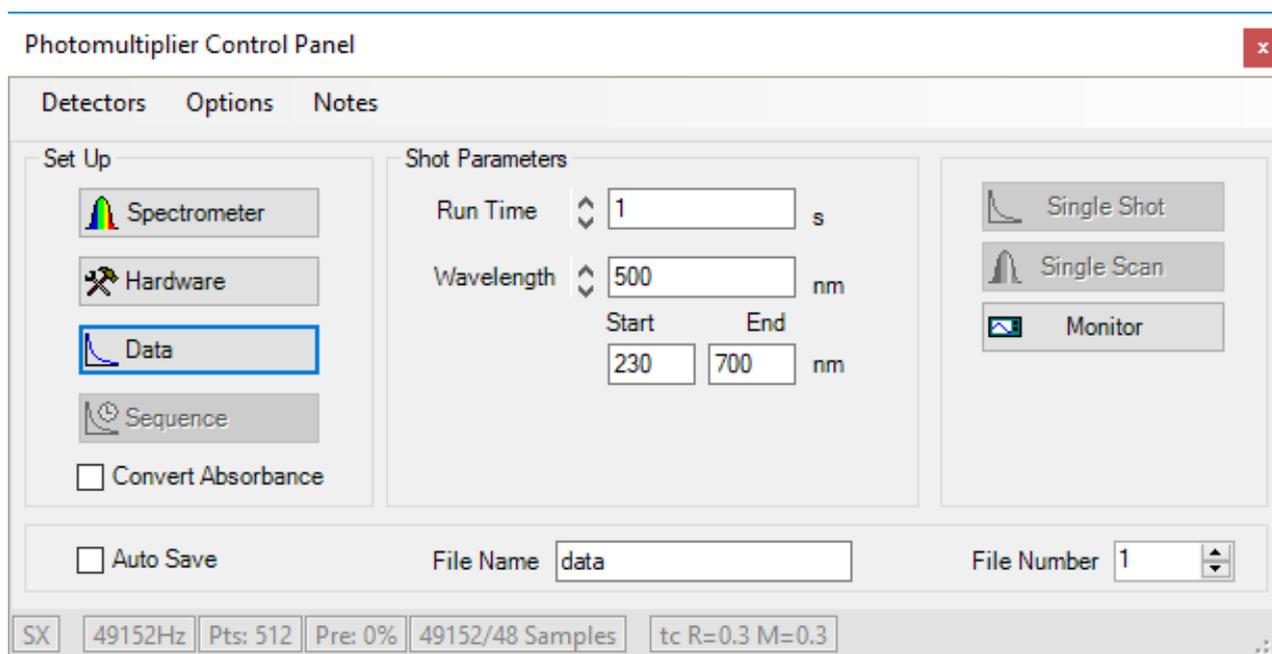
Pretrigger can be configured as a percentage of the run time prescribed. (This advanced feature allows observation of the flow time prior to stopping in stopped-flow acquisitions).

## Executing a Shot

With the instrument set up with appropriate optical signal levels, the user can start to collect data.

**Important: At this stage it is important that the instrument is filled with reagent or left with buffer or water in the flow circuit, the drive syringes should be in contact with the push plate(s) and generally ready for operation. All instructions and cautions detailed in the hardware manual should be observed.**

The user's attention is drawn to a further examination of the Acquisition Control Panel:



The following features should be noted and considered for the shot:

**'File Name':** The user should provide a filename which will enable them to identify their data. An File Number will be added to each file name and will increment with acquisitions. Data will be saved to the working folder set under the File menu item. Data collected will default to "data..." until set by the user.

**'Auto-save':** Defaults to not enabled – check the box to set. This is not necessarily recommended until the user is quite happy that collected data are as required – it can be too easy to store lots of priming and false shots!

**'Notes':** While many users are quite happy to identify their data by file name and folder this feature allows a text window to open which can be used to further enhance identification of experimental conditions etc and any user input comments. Leave blank by default if not required.

**'Convert to absorbance':** This option should be checked when the data type is transmission/absorption and the user would like their data converted to absorbance units directly upon acquisition. Note other data types such as fluorescence will inhibit this selection.

**'Logarithmic timebase':** should be enabled if the user wants the acquisition time base to be logarithmic (ie not linear). Ideal for multi-phasic observations; not good for fast time bases, lag phases, inexperienced users. Leave unchecked if unsure about this. The setting is available under the **'Options'** menu.

More explanation is given later in this section for these features – at the moment, it is best to simply concentrate on getting going ...

### **To do a shot ...**

Select the Wavelength and the Run Time to be used for the measurement then use the Single Shot button and the user should observe the emptying of the Stop/Waste syringe followed by the actuation of the air drive to effect a stopped-flow mixing shot.

### **To scan the contents of the cell ...**

Hit the Single Scan button and the monochromator will slew between the Start and End wavelengths shown below this control revealing the spectrum of the reagent or product.

In the following, more detail is given for those users wishing to take advantage of the more advanced features relating to data set parameters...

## **Optimising the Dataset Parameters**

For many users, the setting up Kinetic Studio for an experiment is to configure the number of data points required, oversampling and pretrigger.

The number of data points relates to the final dataset and graph. This can be adjusted depending on the experimental requirements and trace detail. The typical number of data points used in an experiment is 512.

Oversampling can be used to improve signal to noise and hence the quality of data. For every data point, oversampling corresponds to the number of additional samples that are averaged together.

For example, the screenshot above shows 512 data points have been specified with 48 oversamples. This means the data acquisition device will acquire 512 x 48 samples (24576 samples in total). Every data point is an average of 48 samples. The software will automatically perform the averaging after the experiment has completed.

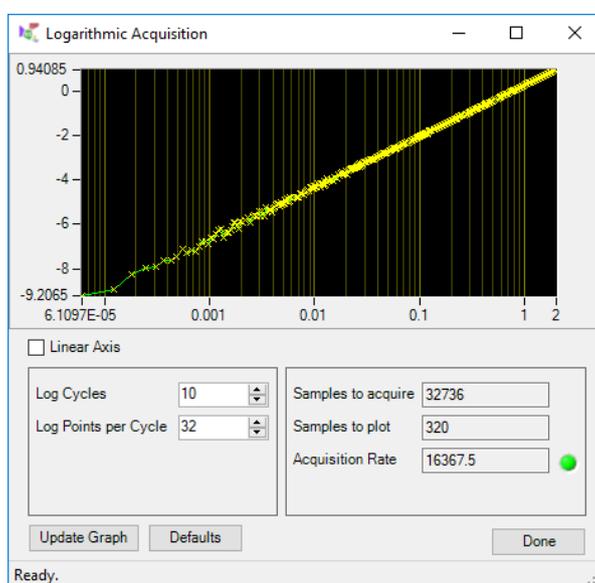
Pretrigger can be used to acquire data just prior to the trigger point. The amount of pretrigger is specified as a percentage of the run time.

## Setting the Run Time

The experimental run time can be manually entered by editing the **'Run Time'** numeric box or alternatively a series of standard run times can be applied by clicking the small up and down arrows next to the **'Run Time'** box.

For shots requiring a log acquisition and hence being displayed with a logarithmic x-axis, the **'Logarithmic timebase'** option can be enabled. The standard log timebase applied is 10 log cycles with 64 points per log cycle. Kinetic Studio intelligently uses the additional samples acquired for logarithmic processing to apply data averaging improving the signal to noise.

Should the experiment require it, a custom log mode facility can be accessed by pressing the link **'Logarithmic timebase'**. This will display a log parameter editing utility as shown.



If the Conductivity device being used is an option for the Stopped-Flow system, the ability to edit the **'Age-Time'** will be made available.

## Specifying a file name

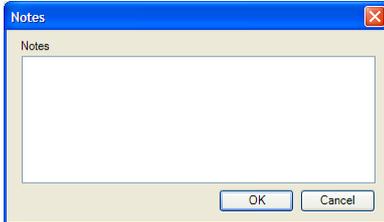
Within the **'Data Set File Name'** area to the bottom left of the control panel, enter a file name that corresponds to the experiment being performed. If required (generally recommended), create a folder for the group of experiments being performed and save all related data into that folder.

Kinetic Studio provides a convenient facility to specify a working folder.

Please consult the **'How to Change the Current Working Folder'** subsection within the **'Dataset File Management'** section.

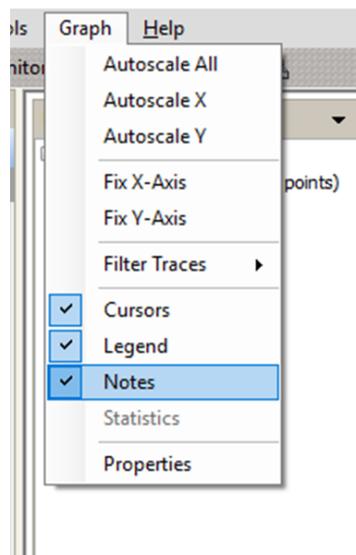
## Adding Notes to a Dataset

Clicking the **'Set Notes for Dataset'** in the **'Notes'** menu of the control panel will display a small notes editor.

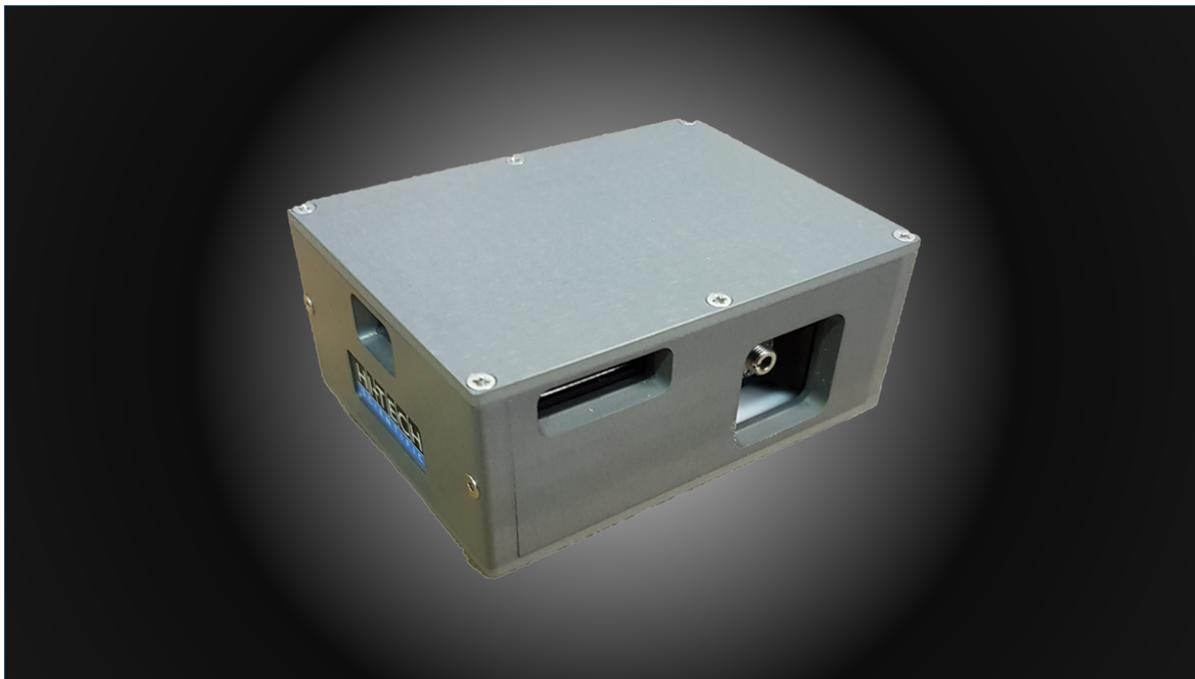


The notes editor provides a facility to save experimental information or comments with a dataset. These notes will be applied to every shot.

Notes for a given dataset can be displayed and edited by selecting the **'Notes'** menu item under the **'Graph'** menu as shown below.



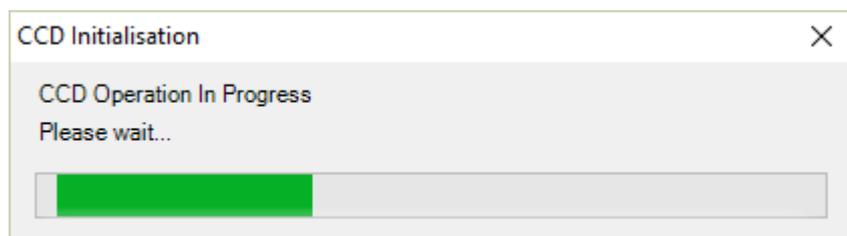
# CCD Array



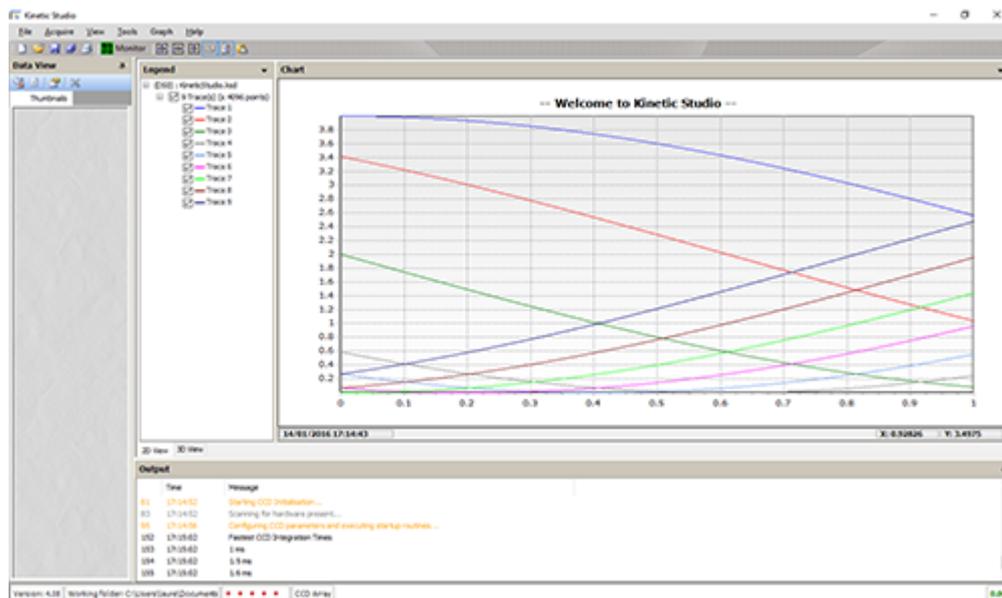
## Launching Kinetic Studio in CCD / Spectrometer Mode

When Kinetic Studio starts in one of the CCD modes it will automatically detect the connected device. If it is unable to find a supported CCD device an error message will be displayed in the log window at the bottom of the screen.

If a CCD device is successfully detected, Kinetic Studio will initialise the device and run some tests to confirm functionality and performance. Please wait for these tests to complete.



Once the tests have finished the software is ready for use.



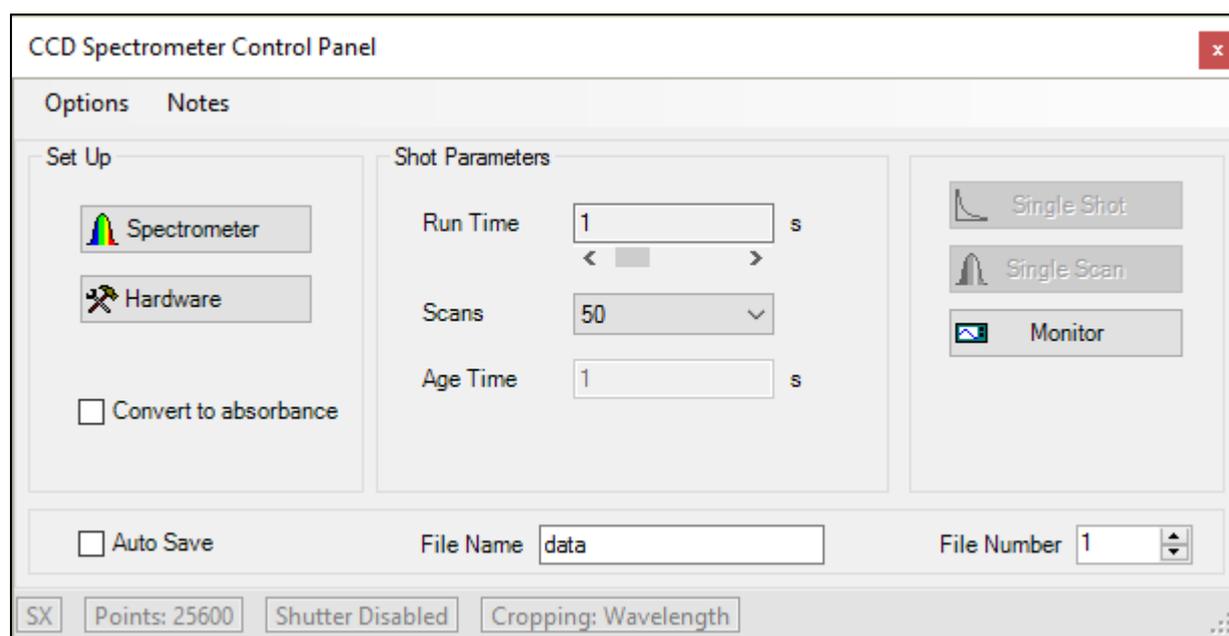
To start using the CCD press the 'New Document' icon in the top left of the toolbar, press **'File' -> 'New'** or select **'Control Panel'** under the **'Acquire'** menu.

## Standalone CCD Mode - Set Up, Calibrate, Scan a Blank

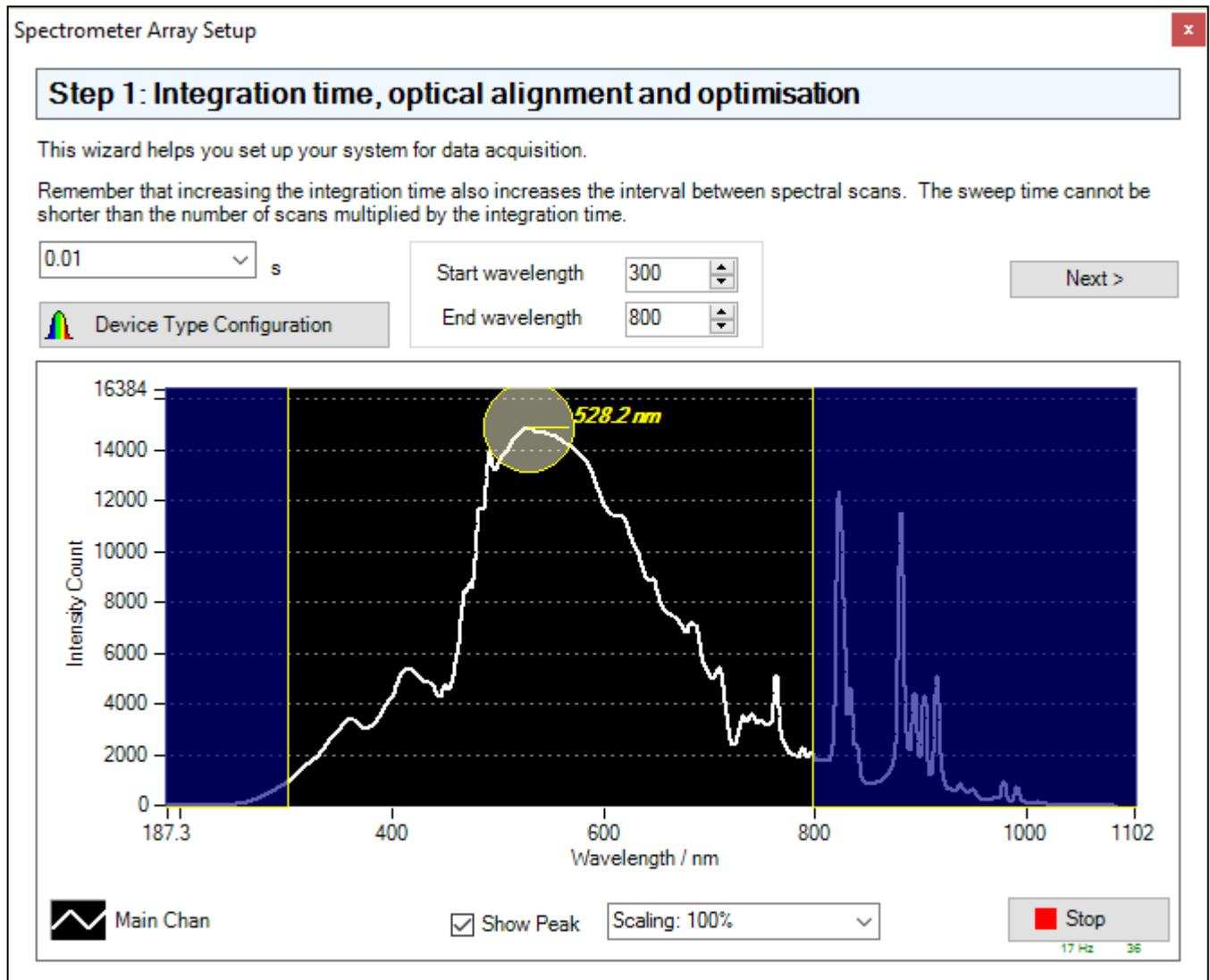
*This mode allows users to set up the CCD Array detector for the acquisition of whole UV/VIS absorbance spectra both as single scans and multi-scans in a kinetic mode.*

The screen shot below shows the CCD Array Control Panel which provides the access to the set up and acquisition routines necessary for the effective use of the CCD array detector with the stopped-flow system.

Press the '**Spectrometer**' button under Set Up to enter the manual setup mode:



This will display the '**Array Setup Wizard**' dialogue:



Now the user can make adjustments to the integration time, optical alignment and depending on the light source, the light intensity and hence signal level. Note that this signal must not saturate the full scale.

In addition to setting the integration time, the user can select the region of interest. The live display shows the full spectrum from the CCD including areas that may not be of interest. The 'Start wavelength' and 'End wavelength' panel will set the yellow vertical lines on the display highlighting the current region of interest. Anything outside of this region will be cropped and can be disregarded. Please ensure the signal IS NOT saturated within the region of interest, otherwise this can affect subsequent scans and shots by giving invalid data in saturated regions.

**ADVANCED:** Areas outside of the region of interest can be saturated (full signal beyond the scale of the device). This allows the optics to be optimised for the region of interest. This can be of particular help in the UV region.



*Technical Help: the integration time is the time between resets on the array - it sets the maximum scan rate such that the minimum integration time allows the maximum scan rate. This also relates to the availability of light and noise performance - with a bright, intense source (eg. the xenon lamp) more photons are available and thus the shortest integration times should be usable.*

The '**Hardware Type and Settings**' button provides access to the following configuration options:

Spectrometer Array Setup x

### Hardware Type and Settings

Spectrograph Array Post-Processing

Enable Smoothing ●

Apply smoothing to blank:  Yes / No

Apply smoothing to data:  Yes / No

Savitzky-Golay ●

Savitzky-Golay Smoothing Enabled:  Yes / No

Savitzky-Golay Window Coefficients:

Savitzky-Golay Window Length:

Binning ●

Pixel binning Enabled:  Yes / No

Pixel binning length:

Moving Average ●

Symmetric moving window smoothing enabled:  Yes / No

Symmetric moving window passes:

Symmetric moving window length:

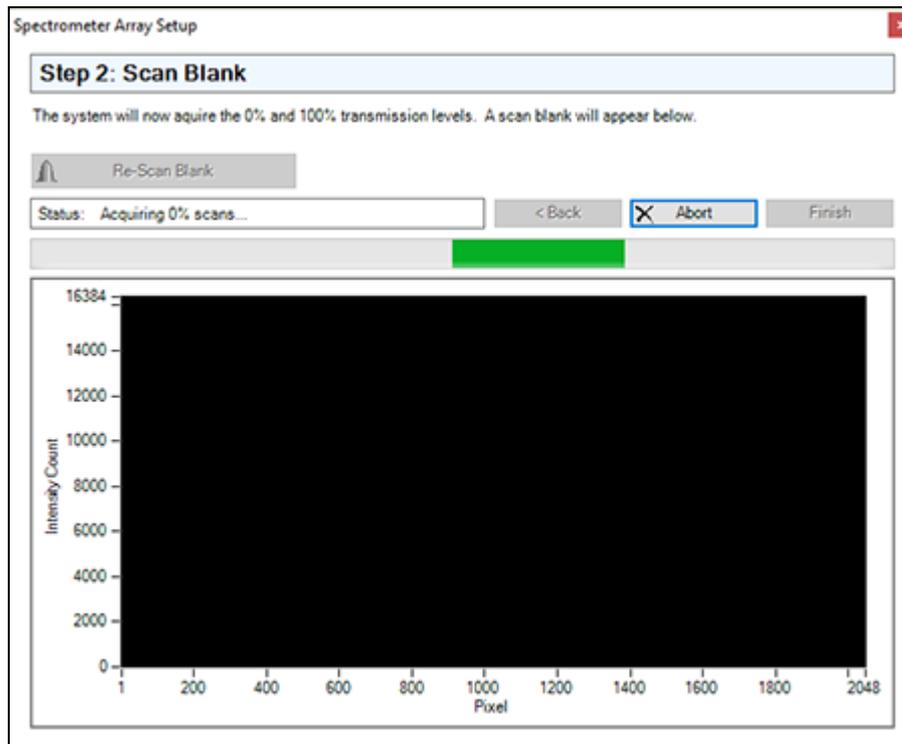
Scan Blank Averaging ●

Multiple blank scan average:

[Done](#)

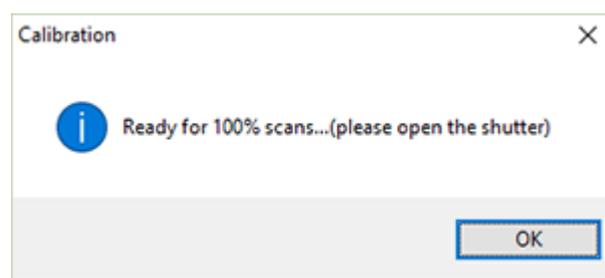
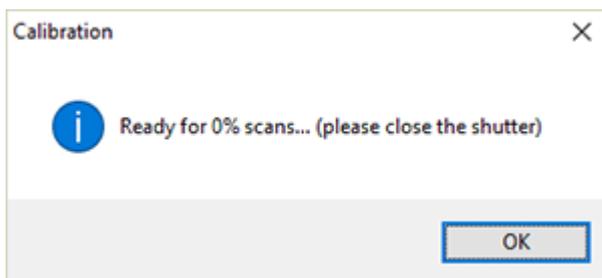
If any of these options are set, the settings will be applied to every scan or shot automatically. Please note that post-acquisition smoothing, binning and averaging is also available via the '**Tools**' menu. When the options have been chosen, press '**Done**' to return to step 1 of the setup wizard.

After the optics have been optimised and an appropriate integration time select, press **'Next'** to proceed to the 0% and 100% reference scans procedure on step 2 of the wizard.



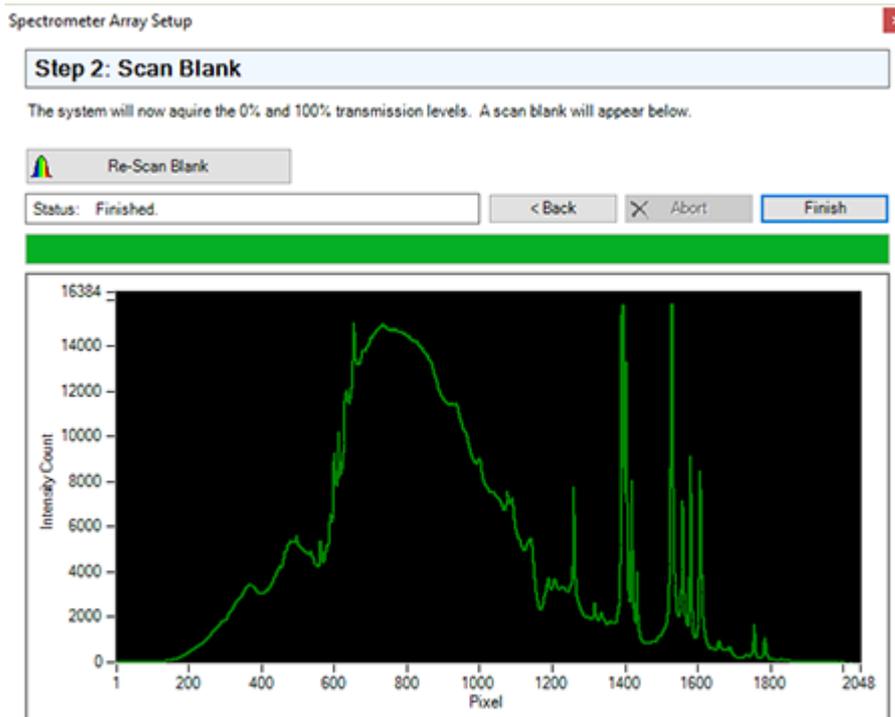
Using **'Next'** the system will acquire the 0% Transmission and 100% Transmission data and thus the scan blank.

If a shutter cable assembly is available with a supported light source, or the optics are part of an integrated system supplied by TgK Scientific Ltd then the 0% and 100% acquisitions will be automated. For all other hardware Kinetic Studio will prompt to close the light off for a 0% scan reference and then re-open the light source for the 100% reference



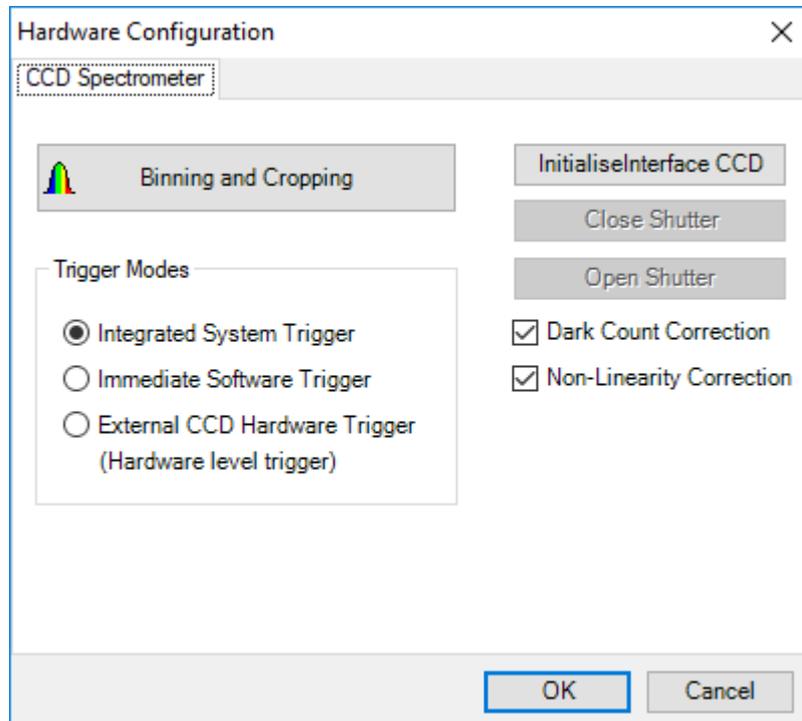
Kinetic Studio will run some initialisation routines on the CCD after it has acquired the 0% and 100% references and will display a please wait notice.

Once the scan blank and initialisation process has completed the software will present a **'Finish'** button and finalise the setup process.



## CCD Hardware Options

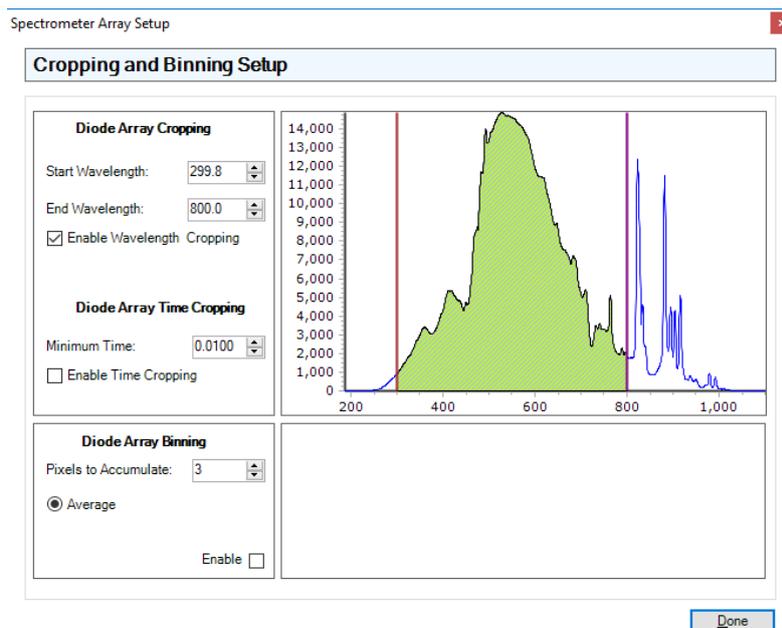
To access hardware options, press the **'Hardware'** button on the control panel. This will display the 'Hardware Configuration' panel as below:



## Binning and Cropping

Kinetic Studio allows the user to automatically crop diode array data or 'bin' pixels.

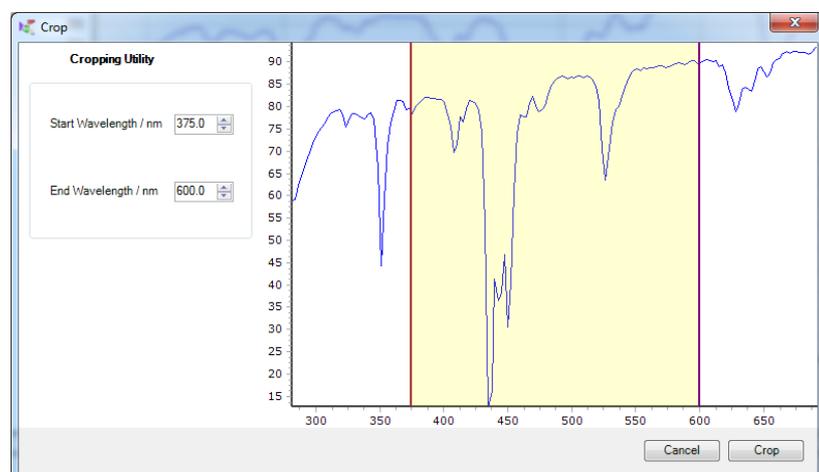
Pressing the **'Binning and Cropping'** button will show the following setup screen:



Binning allows the user to potentially increase signal to noise by combining adjacent pixels on the CCD Array unit. For example, this means a 1024 pixel diode array with binning set to 2 will appear as a 512 pixel diode array. Options here include **'Average'** (bin the pixels and use the average of the binned pixels), **'Cumulative'** (add the value of the binned pixels together) and **'Adaptive'** (using the number of pixels to accumulate as a maximum threshold, the software increases or decreases the amount of averaging depending on the light throughput. This should yield improved signal to noise in regions that have less light or more absorbance).

Enabling the cropping option causes scans to be automatically 'trimmed' to the region of interest. This is the area highlighted in green.

A manual, post-acquisition cropping facility is available via the **'Tools'** -> **'Crop'** menu item.



If the dataset has been rotated, the cropping edit boxes will reflect time rather than wavelength.

## Trigger Modes

The trigger modes panel in the Hardware Configuration Window allows different criteria to be set for data acquisition to start. These will normally be set by Kinetic Studio depending on the shortcut used to launch it.

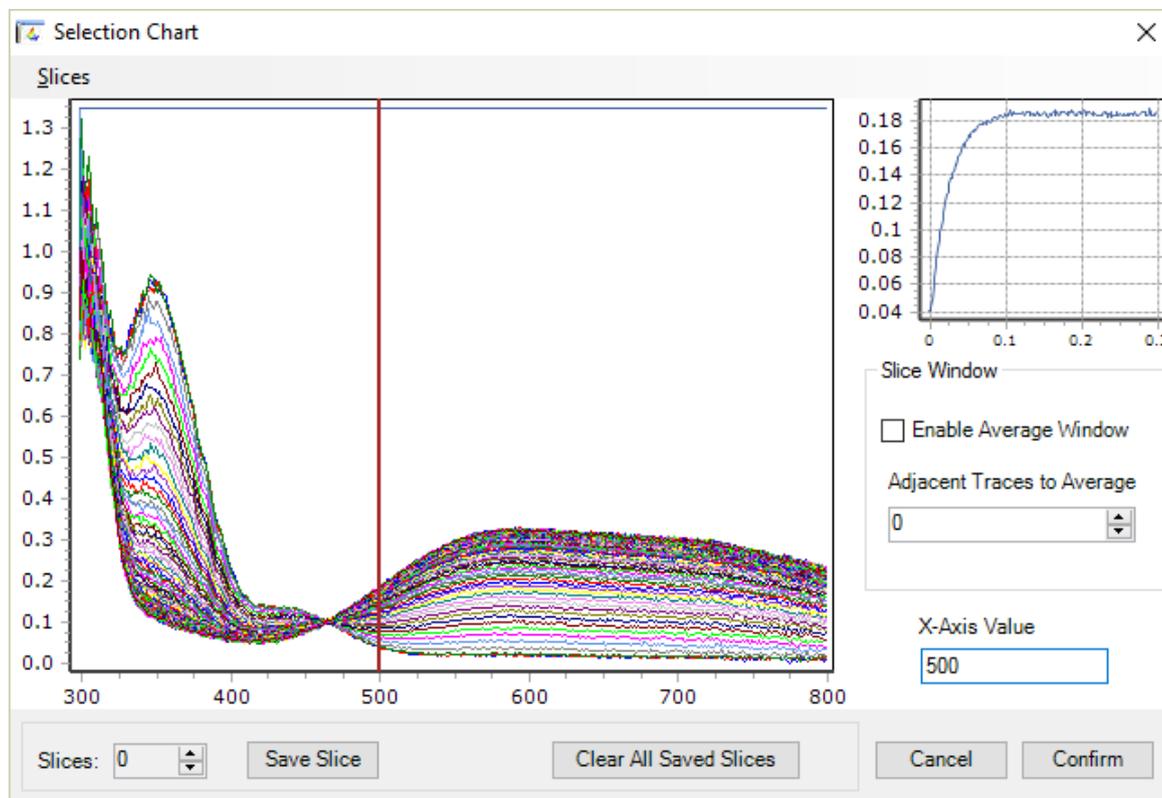
**'Integrated System Trigger'**: Use when the CCD is part of a SF61-SX/DX system.

**'Immediate Software Trigger'**: Use when there is no connected trigger functionality. Data acquisition starts immediately after pressing **'Single Shot'**. It is recommended to extend the run time when using this mode to allow for manually activating the drive.

**'External CCD Hardware Trigger'**: Use when the CCD has a connected external trigger system.

## Slice

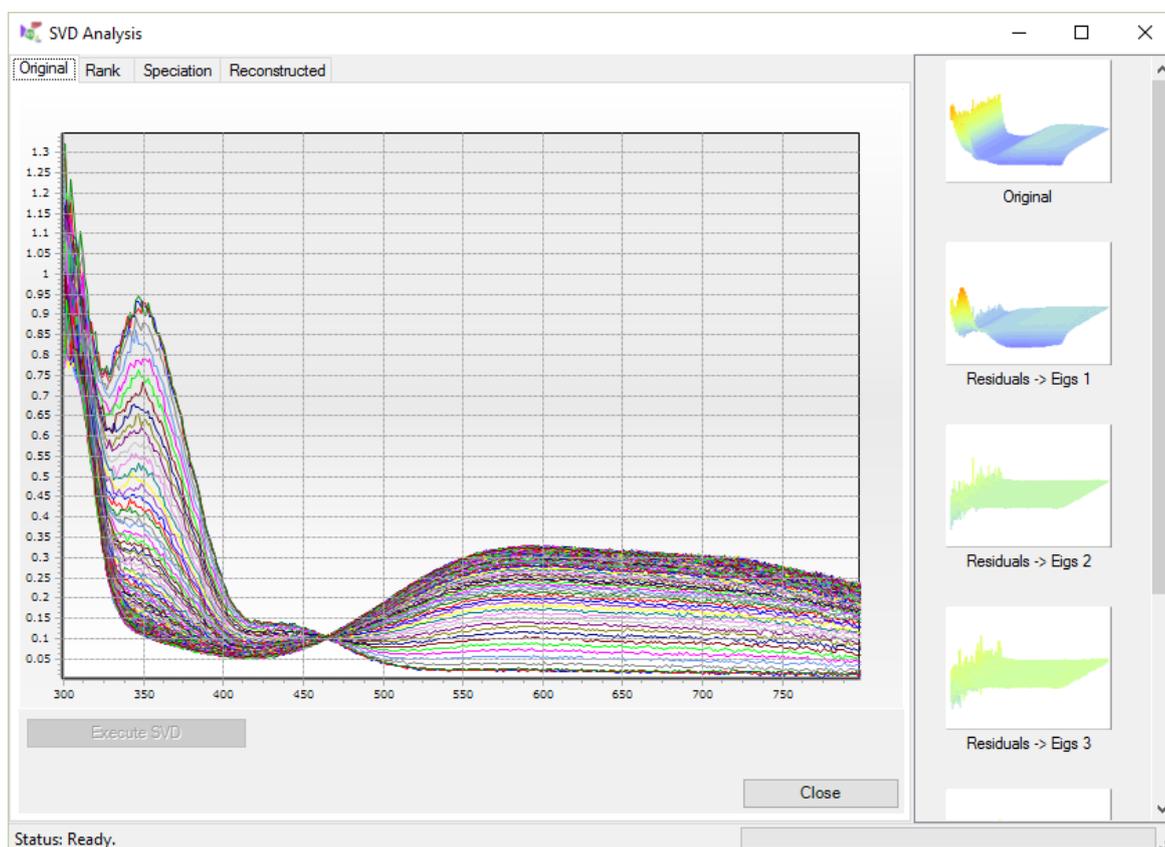
Kinetic data can be sliced at a particular wavelength. This enables the user to quickly extract 2D data in terms of signal versus time at a specific wavelength.



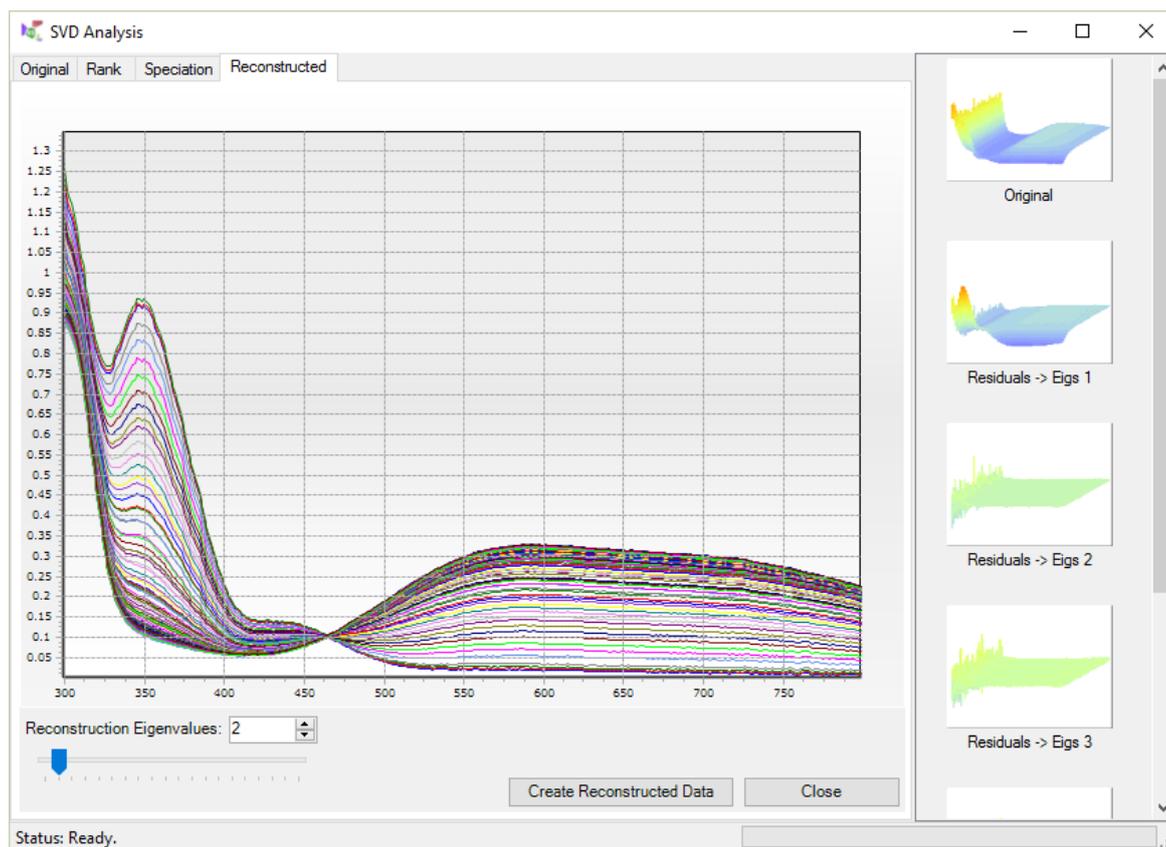
## SVD Analysis

Kinetic Studio provides access to Singular Value Decomposition facilities in addition to various other data processing functions and analysis tools.

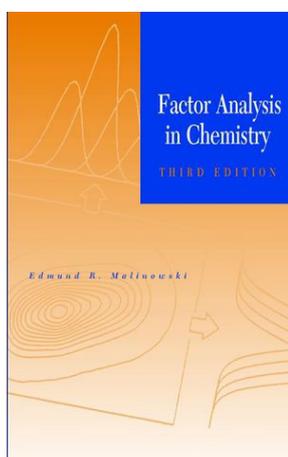
**Please ensure data is saved prior to using the SVD tool.**



SVD is a powerful tool that can help analyse the complex stages and components in a dataset. Additionally Kinetic Studio allows the data to be reconstructed from a reduced number of eigenvectors facilitating the removal of unwanted components / noise.



For more information on SVD, factor analysis and other related analytical techniques that may be useful, please refer to the following book:



Factor Analysis in Chemistry

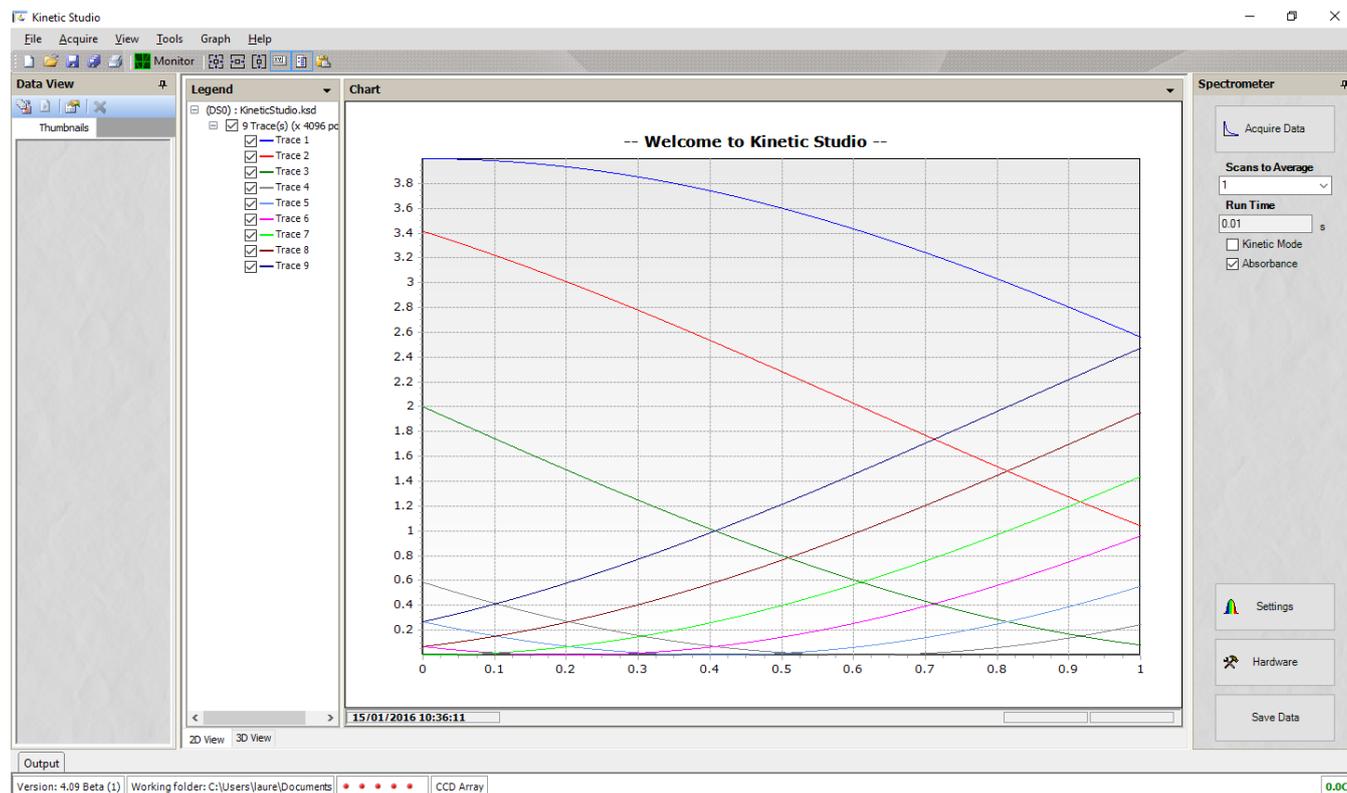
Edmund R. Malinowski

ISBN: 978-0-471-13479-4

## Spectrometer Mode

Kinetic Studio can be started in a dedicated Spectrometer Mode providing quick and easy access to single and multiple scans with averaging.

A dedicated control panel is added to the right-hand side of the screen putting all of the required functions for using the CCD right in front of the user.



Spectrometer mode when used in conjunction with a cuvette holder and suitable light source gives the convenience and flexibility of a modular bench-top spectrometer.

Additional functions and features are being added to spectrometer mode with new versions of Kinetic Studio. If there are specific features or suggestions that you would like to be considered for inclusion, please contact TgK Scientific Ltd.

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# How to use the Sequence Setup

The features found in this dialogue are used to execute shots (for stopped-flow) and jumps (for temperature-jump), and acquire data using user tailored routines enabling experiments to be run in a semi-automated fashion often aiding productivity and enhancing reproducibility. Upon exiting this dialogue, the acquisition control panel "Shot button" will display **'Acquire Series'** ... clicking this will commence the execution of the defined sequence without any further user intervention (except to refill reagents if necessary).

Programmed Acquisition Sequence

Shot Details

	Wavelength	Run Time	Z Value
*			

Acquisition Mode:  
Single Shot  Confirm after each shot [Clear Shots](#)  
 3D data set

Time: 1 Wait time between shots 60  
  Enable delay between shots

Set Sequence

Wavelength | Age Time | Averages

Wavelength

Start 450 nm  
End 550 nm  
Interval 20 nm

[Set Wavelengths](#)

## Average sequence

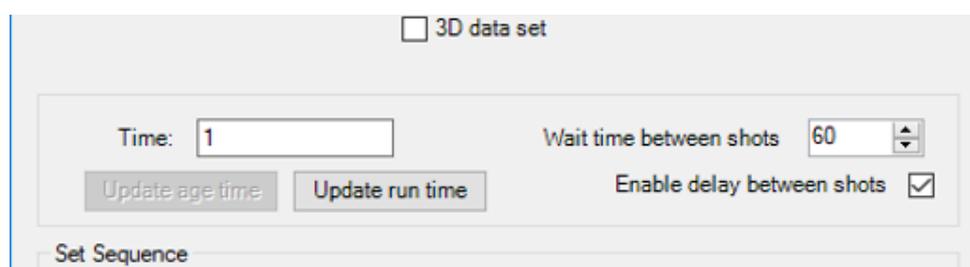
The tab labelled "Averages" accesses a sequence where the user can set up a sequence of shots at a defined wavelength over a defined run time – The user enters the desired number of shots to be executed and these are subsequently presented with their average in a combined data set.

## Time Delay Sequence

A delay period can be added to a sequence to allow for temperature equilibration between shots. This is particularly pertinent to T-Jump operation where multiple jumps are necessarily averaged and to T-pod operation where the mixing cell is kept at a different temperature to the syringes. The difference in this mode is that a 'delay period' can be defined within the sequence dialogue. This allows the sample to cool sufficiently before the next shot in the sequence. If the user should want to execute the shot early (ie. before the delay period has expired), the user can cancel this 'delay period', after which the shot will fire.

In addition to the cooling period, the T-Jump will wait for a number of seconds prior to the shot so that the capacitor bank has time to charge and prepare for the shot (this can be changed under the **'Tools'...'Options'...'User'...'TJump'...'TJump Recharge Time'** option).

The cooling period is set to 60 seconds by default. This can be enabled using the check box in the sequence dialogue and is available for editing:



## Wavelength Sequence

The tab labelled "Wavelength" allows the automated collection of shots over a wavelength range collecting a shot at each discrete wavelength and then making the sequence of shots into a pseudo 3D block where spectra-kinetic data can be assembled from the multiple shots.

## Age Time DX Sequence

The tab labelled "Age Time" is applicable to double mixing instruments where the user can run a sequence of DX shots where the age time increments as defined by the user in the dialogue. The resulting data will be a series of shots where the age time becomes the third dimension.

## In the Event a Sequence Stalls

While it is best to organise a sequence of shots that only consume reagent within the limits of the drive syringe, often it is inevitable that a sequence of shots will run out of reagents. Under such circumstances two options are available:

**When more reagent is available:** The sequence can be halted by clicking '**Abort**' within the Acquisition in Progress dialogue. The user is confronted by options to Continue or Cancel. This allows for refilling and then using the Continue command, the sequence will continue.

**When more reagent is not available:** In this case the Cancel command will abandon any further attempt to complete the sequence but keep the data collected so far.

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# Double Mixing Stopped-Flow



## Enabling Double Mixing Modes

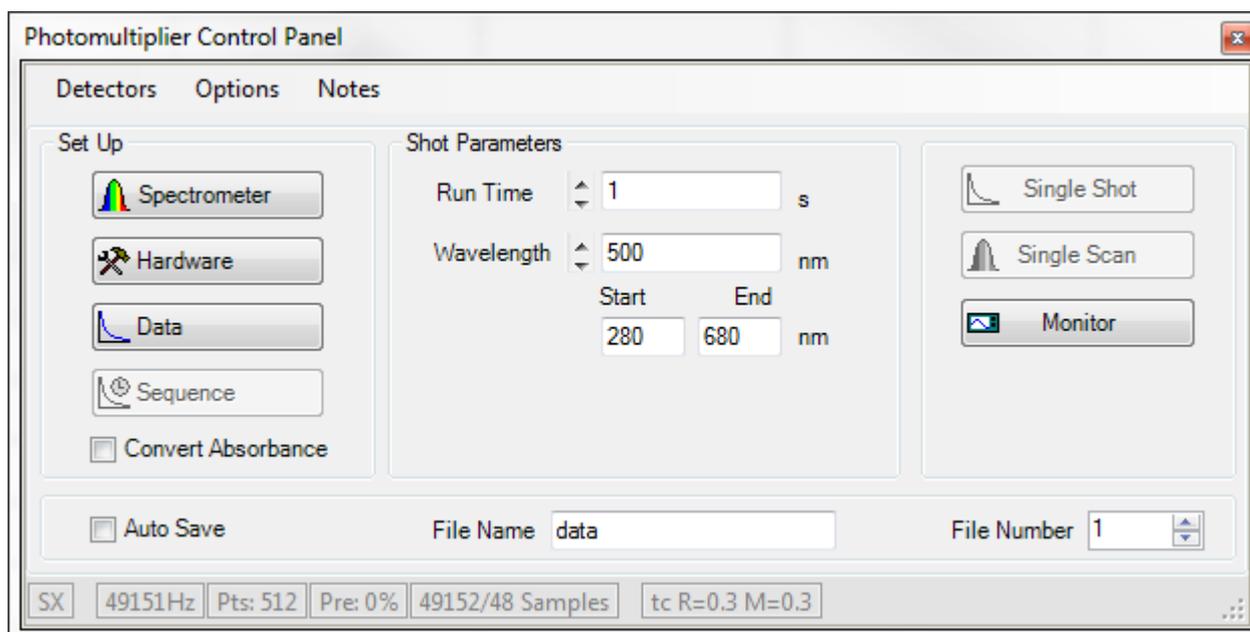
The double mixing (DX) (sometimes called, sequential mixing) capability of the SF-61DX2 instruments is controlled from Kinetic Studio – the following provides a guide to how to run a DX experiment.

It should be understood that SF-61DX2 units can be configured to operate in either single mixing mode or double mixing mode at the click of a (mouse) button. The sample handling unit features a convenient light on the front panel to indicate which mode it is currently set to.

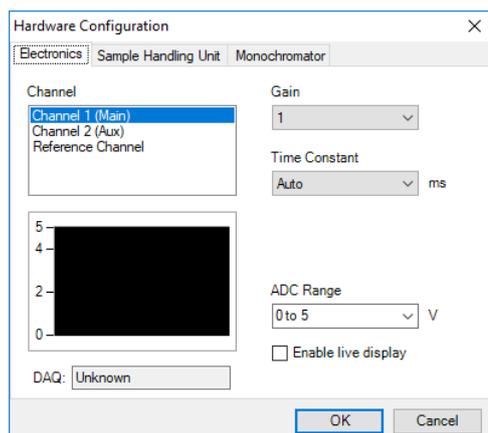
It is important to adjust the waste stopping block to the correct volume depending on whether single or double mixing mode is enabled. This takes into account the syringe volumes within the instrument and the volume required for the first and second stages of the sample delivery.

The user is urged to study the appropriate sections of the hardware manual to familiarise themselves with the system plumbing and the general arrangement of the sample handling unit – failure to do this will likely limit the users full understanding of the DX operation.

Upon start up and initialisation of the system, the default condition of single mixing mode (SX) is selected. The user need only continue to read this section if double mixing mode (DX) is required.



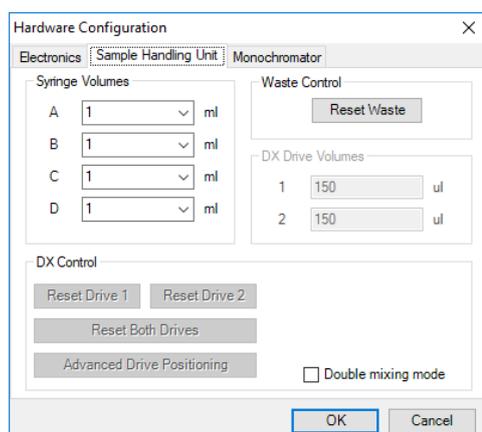
On the control panel, please press the '**Hardware**' button to access the sample handling unit configuration panel.



This will display the panel as pictured here.

The first panel displayed is the '**Electronics**' page.

Click on the '**Sample Handling Unit**' tab.



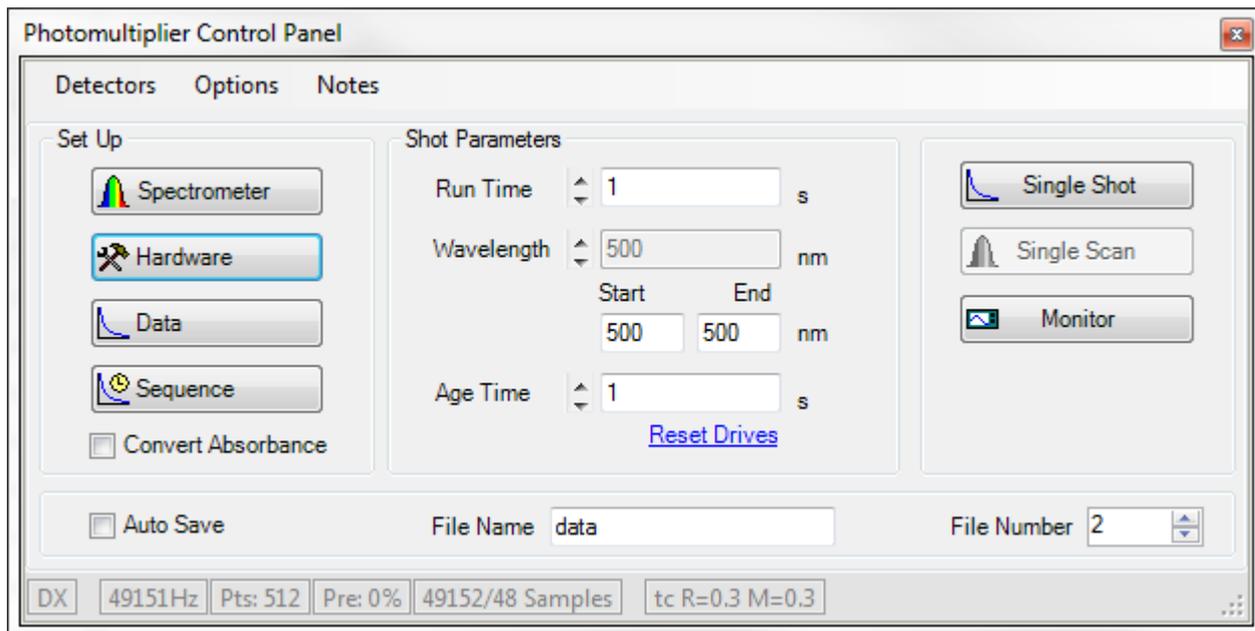
Enabling or disabling double mixing mode is done by adding or removing the check mark in the '**Double mixing mode**' box in the bottom right corner.

As soon as the check mark is added or removed the command is sent to the instrument. The indicator on the front panel of the SHU will immediately change to reflect the current mode of operation.

**Note:** For double-mixing experiments the stop block has to be set to 240  $\mu$ l stopping volume. To move the stop block, loosen the three socket screws (at the rear of the Sample Handling Unit) and with the WASTE/STOP valve set to WASTE, pull down the stop syringe piston until it reaches 240 ml. Move the stop block with the piston and tighten the three screws. (Refer to the hardware manual).

Ensure the stepper motor drives are reset – this can be achieved by clicking the relevant reset button in the '**Sample Handling Unit**' tab in the Hardware Configuration dialog.

Next, the Acquisition Control Panel is used in a similar manner to single mixing mode and the acquisition run time and wavelength is set (it is assumed that optical and data set up protocols have been gone through at this stage).



Note that in DX mode the Age Time control is enabled – this is where the user can set a target age time and thus effect the pre-mixed sample incubation time between the two drives.

It is strongly advised that users become familiar with the operation in this mode and consider starting with some test runs with water or buffer. An age time should be set – and it is recommended that a modest value such as the default 1 second is used initially. Click the Single Shot button and the user should hear (albeit faintly) the whirring sound of the stepper motor driven blocks moving into position as a precursor to doing the shot. With the block sets, the Stop/Waste valve will toggle and the stop syringe empty ... soon after Drive 1 will fire and part fill the stop syringe, then after the time delay, Drive 2 will complete the action of the sample delivery and thus effect the stop flow shot and data acquisition.

## How to Reload the Drive Syringes During a Shot

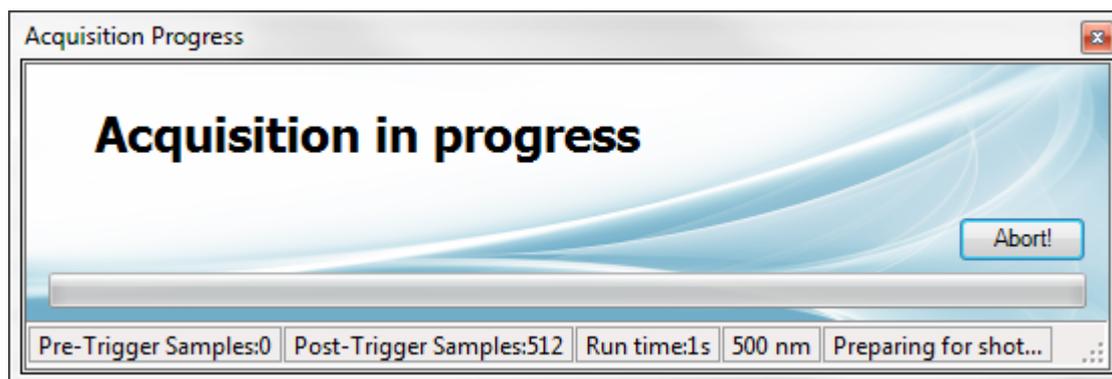
This action is mostly required during a multi-shot sequence where more shots are programmed than can be obtained from the drive syringes. This happens because there is insufficient solution in the drive syringes to fill the stop syringe, the data acquisition does not get triggered and the air drive does not deactivate.

Depending on which mixing mode is used, follow the relevant steps...

### How to Reload the Drive Syringes During a Single Mixing Shot

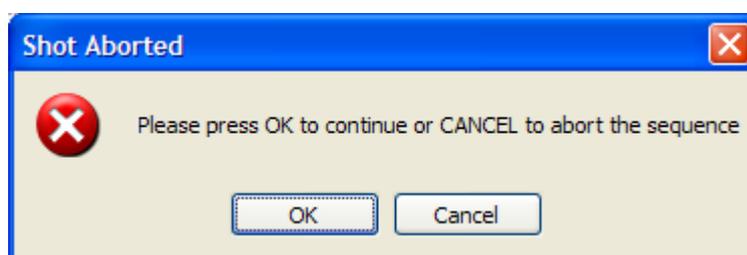
The software will be waiting for a trigger signal but if there was insufficient volume remaining in the syringe, the trigger mark will never be reached.

To reload the syringe and execute the shot again, the first step is to **'Abort'** the current operation.



If the operation was a single shot, Kinetic Studio will return automatically to the control panel.

If the operation was part of a series, Kinetic Studio will ask what should be done next.

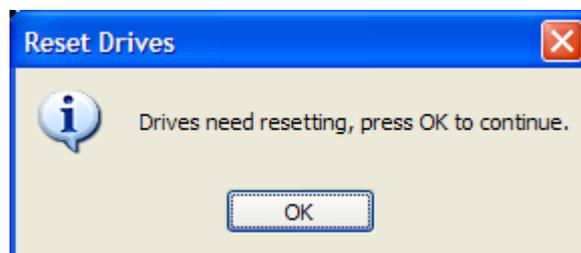


At this point, the syringe can be reloaded. After the syringe has been refilled, a repeat of the last shot can be initiated by pressing the '**OK**' button. Alternatively the '**Cancel**' button can be pressed to abort the current programmed sequence.

## How to Reload the Drive Syringes During a Double Mixing Shot

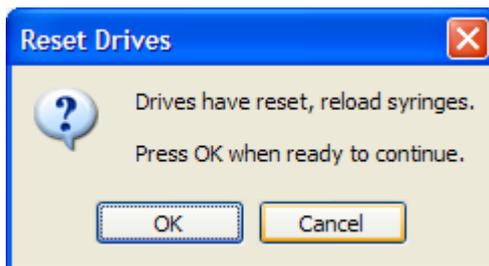
The software intelligently tracks where the stepper motors are during a shot sequence. If it detects there is insufficient volume, it will inform the user and prompt for what should be done next.

Do **NOT** act on the message below until the drive syringes have been reloaded.



Once the drive syringes are ready, press '**OK**'.

Kinetic Studio will then prompt to click '**OK**' to continue and repeat the last shot, or '**Cancel**' to abort the shot sequence.

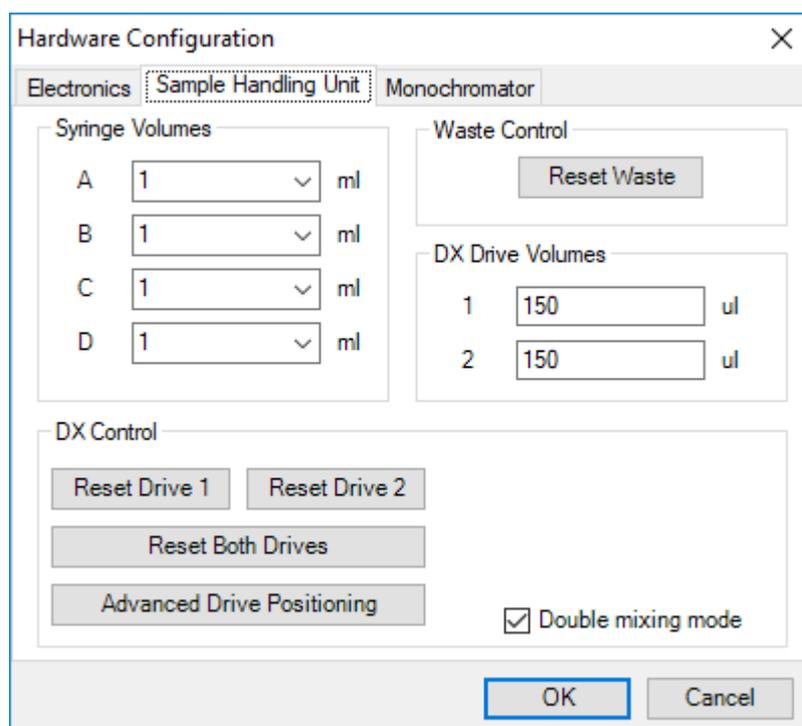


## Advanced Drive Positioning

Kinetic Studio allows accurate and customisable drive positioning for both drives 1 and 2.

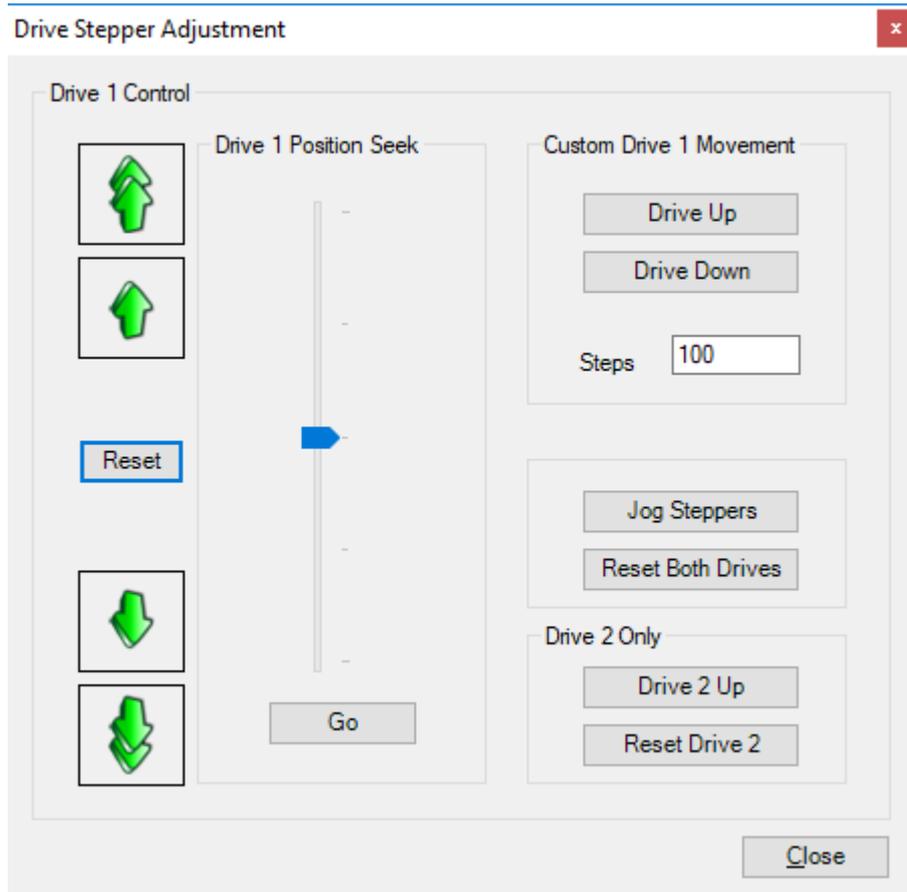
To access this facility, please go to the 'Hardware Configuration' panel by pressing the **'Hardware'** button within the control panel.

Within the 'Hardware Configuration' panel, select the Sample Handling Unit tab.



Press the **'Advanced Drive Positioning'** button.

The software will prompt you to make valves A and B are in the fill position before making any changes.



The green arrow buttons on the left provide fine and larger more granular adjustment of Drive 1.

**'Reset'** will reset Drive 1 only.

The 'Drive 1 Position Seek' section allows the drive to be quickly moved to some general positions. The scale can be approximated to the full range of the drive. For example, if the middle position is selected and the **'Go'** button pressed, the drive will locate to the midpoint.

The 'Custom Drive 1 Movement' enables the user to enter custom stepper movements for drive 1.

# qPod Mode

In this mode of operation the qPod device is fitted as described in the hardware manual providing a configuration that enables the chemical quenching of reaction volumes using the double mixing mode of operation. However, without the need to acquire data from an optical signal, the process is merely one of running a series of time delayed, quenched samples, and collecting them. So in this mode, Kinetic Studio does not activate any control of optical devices, it merely enables control of sample delivery.

The qPod mode of Kinetic Studio provides a simplified control dialogue for the user to set up and command time delay shots and record each shot with its associated instrument parameters in a convenient spreadsheet format. Thus a data log of acquired shots is recorded. When the qPod mode initialised, the following screen greets the user:

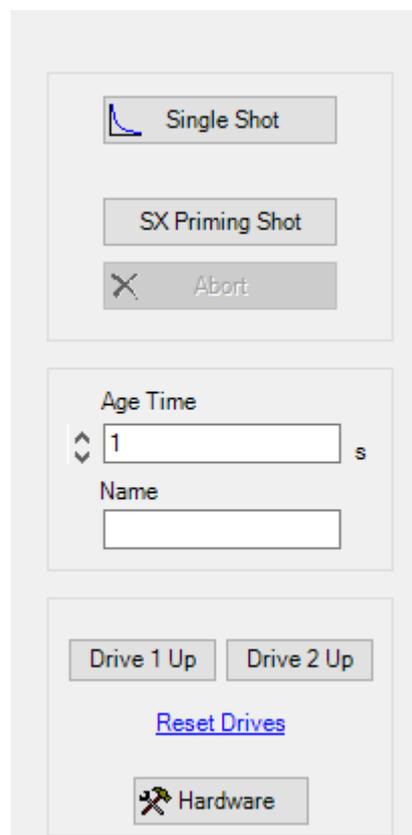
Date	Shot Name	Age Time (s)	Flow Rate 1 (ml/s)	Flow Rate 2 (ml/s)	Temperature (C)	Comment
**						

Output

Time	Message
55 11:49:43	Reset Waste.
61 11:49:43	New Data: Points collected per trace: 100/100
62 11:49:43	Fixed drive: (Simulation)
68 11:49:43	Read actual age time: 1 s
69 11:49:43	Read drive 1 flow rate: 1 ml/s
70 11:49:43	Read drive 2 flow rate: 1 ml/s
74 11:49:43	Mode: SingleShot

Version: 5.03 Working folder: C:\Users\JackCrozier\Documents\SPData qPod Mode

The major area of this is the spreadsheet which records line by line the sequence of shots, the right hand portion is the sample handling unit control panel:



This dialogue provides the features necessary for setting a time delay, commanding a shot and assigning shot names:

**'Single Shot'** is the control to activate a double mixing shot (the normal mode of operation for quenching experiments).

**'SX Priming Shot'** is the control to shoot just C and D syringes (and is used to provide a priming shot to fill or flush the flow circuit).

**'Abort'** enables the user to abort a single shot.

The **'Age Time'** entry box is where the user sets the desired age time. this can be entered numerically from the keyboard or using the up/down arrows.

The **'Name'** entry box allows the user to set the name for each shot - this will be retained during a session until edited by the user and will appear under "Name" in the report spreadsheet.

**'Drive 1 Up'** and **'Drive 2 Up'** are controls that allow the user to move the drive up (via the stepper motors) in an incremental fashion and thus facilitate partial filling of the drive syringes.

**'Reset Drives'** is the control that allows the user to reset the two drives resetting them to the fully retracted, (start) position.

**'Hardware'** opens up another dialogue that provides the user with options to reset drive syringe sizes, shot volumes and other instrument parameters. Generally the default values should be left set unless a special syringe configuration is used.

## Executing a qPod Shot

The operation of the qPod should be studied in the appropriate section of the hardware manual. The operation of Kinetic Studio in this mode is very straightforward and the following screen shot shows the result of running a shot:

The screenshot displays the Kinetic Studio software interface in qPod Mode. The main window features a data table with the following columns: Date, Shot Name, Age Time (s), Flow Rate 1 (ml/s), Flow Rate 2 (ml/s), Temperature (C), and Comment. A single data entry is visible for the date 30/05/2018 12:05:29, with a shot name of 'emulated test', an age time of 1, flow rates of 1 ml/s for both channels, and a temperature of -949.0C. A large, empty comment field is provided for each entry. To the right of the table is a control panel with buttons for 'Single Shot', 'SX Priming Shot', and 'Abort'. Below these are input fields for 'Age Time' (set to 1 s) and 'Name' (set to 'emulated test'). Further down are buttons for 'Drive 1 Up', 'Drive 2 Up', 'Reset Drives', and 'Hardware'. At the bottom of the window is an 'Output' log showing a sequence of messages with timestamps, including 'Reset Waste', 'New Data: Points collected per trace: 100/100', 'Fired drive! (Emulation)', 'Read actual age time: 1 s', 'Read drive 1 flow rate: 1 ml/s', 'Read drive 2 flow rate: 1 ml/s', and 'Mode: Single/king'. The status bar at the bottom indicates 'Version: 5.03', the working folder 'C:\Users\JackCrozier\Documents\SPData', and the current mode 'qPod Mode'.

Date	Shot Name	Age Time (s)	Flow Rate 1 (ml/s)	Flow Rate 2 (ml/s)	Temperature (C)	Comment
30/05/2018 12:05:29	emulated test	1	1	1	-949.0C	

```

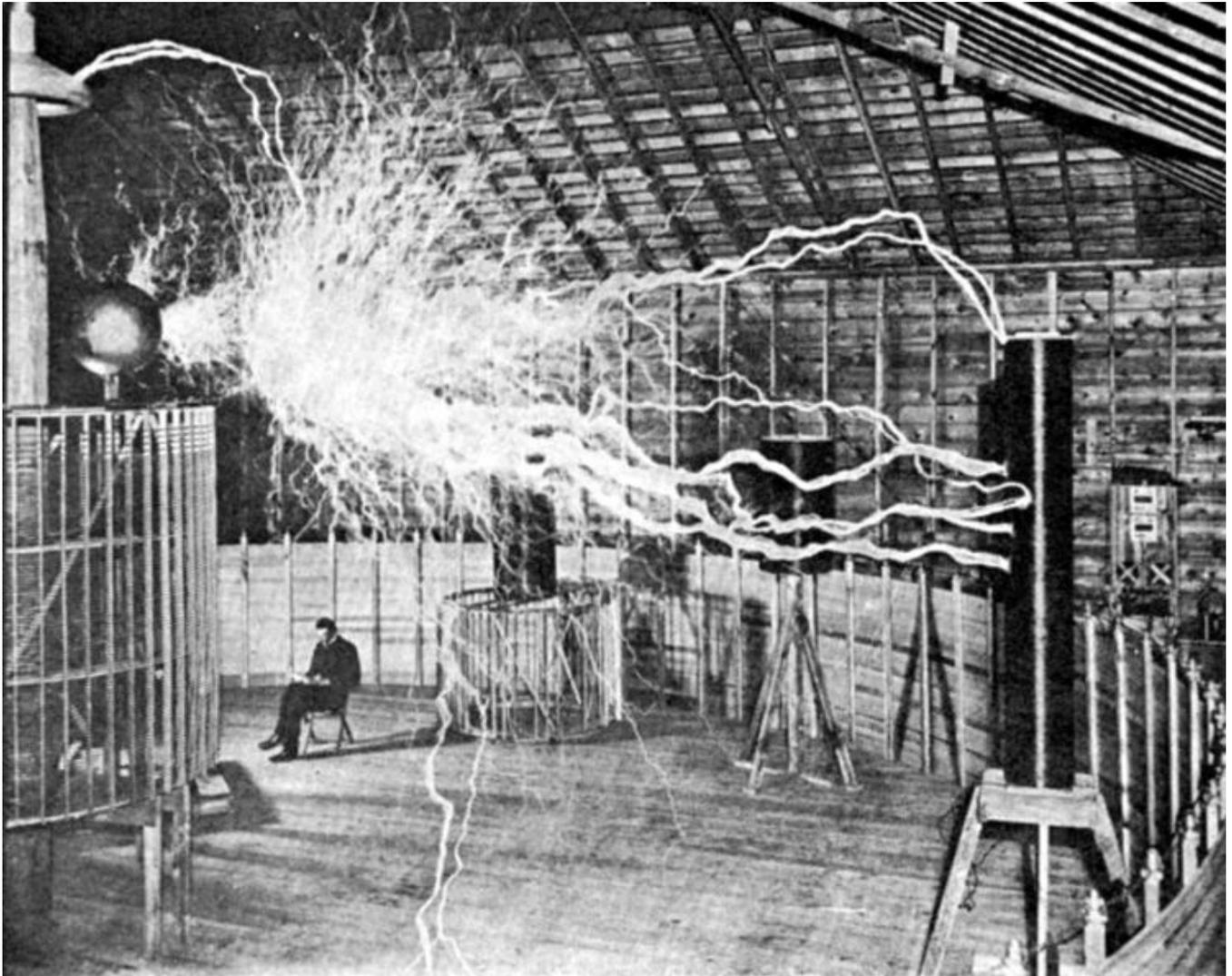
Output
Time      Message
101 12:05:29 Reset Waste.
107 12:05:29 New Data: Points collected per trace: 100/100
108 12:05:29 Fired drive! (Emulation)
114 12:05:29 Read actual age time: 1 s
115 12:05:29 Read drive 1 flow rate: 1 ml/s
116 12:05:29 Read drive 2 flow rate: 1 ml/s
120 12:05:29 Mode: Single/king
  
```

Here the shot has been recorded by the first line entry with age time and other parameters acquired during acquisition. Note that a substantial section has been allocated for the user to enter comments - it is anticipated that this will provide the basis for identifying samples taken from the quench-flow experiment that will be subject to other analytical techniques.

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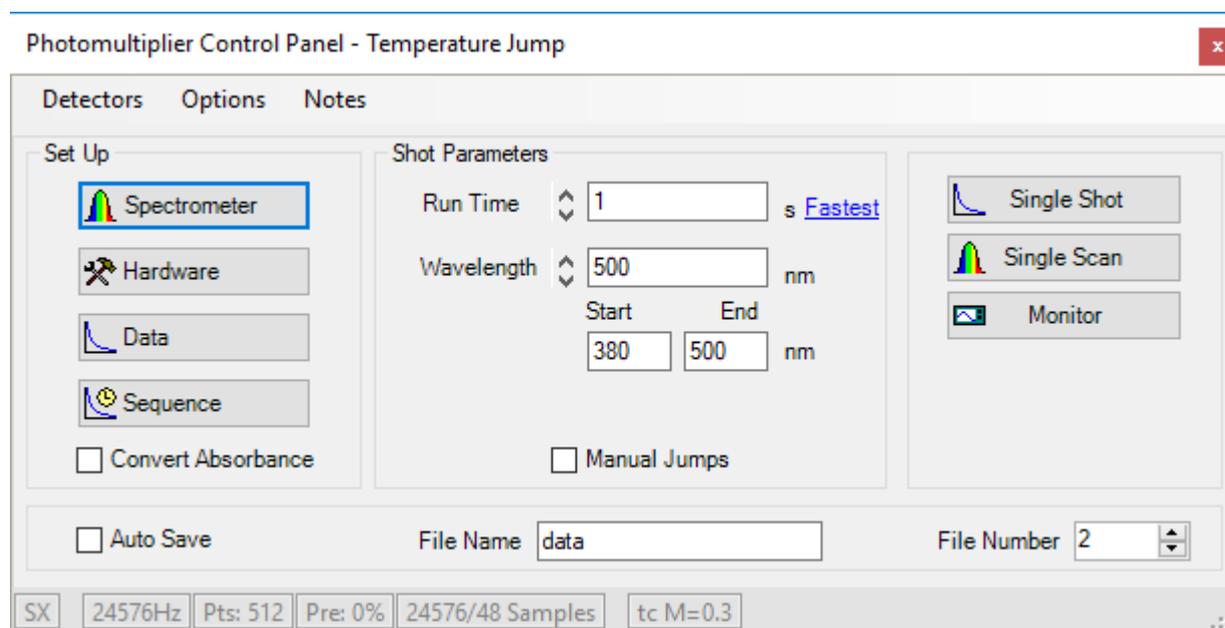
# T-Jump Mode



## How to Setup for an Experiment in T-Jump Mode

### Overview

The Control panel can be displayed by selecting the new document icon on the shortcut bar or by selecting '**Control Panel**' under the '**Acquire**' menu.



The configuration of the software will depend on the type of experiment.

### Initial Setup

#### Setting the Data Type and Dataset Parameters

The first step in setting up Kinetic Studio for an experiment is to configure the main channel and setting the data type.

For T-Jump, the data type options are limited to, and set under '**Data**':

The relevant '**Data Type**' options are:

- Transmission
- Fluorescence
- Light Scatter

And are only applicable to Channel 1(Main). The '**Modifiers**' available are:

- Unknown
- None

The screenshot shows the 'Data Settings' dialog box with the following configuration:

- Channel 1 (Main):**
  - Enabled:
  - Data Type: Transmission
  - Data Type Modifier: None
- Channel 2 (Aux):**
  - Enabled:
  - Data Type: Fluorescence
  - Data Type Modifier: None
- Dataset & Shot Settings:**
  - Data Points: 256
  - Oversamples: 1
  - % Pretrigger: 0
- Save 'Data Type' settings on exit.
- Buttons: OK, Cancel

Each channel can be enabled by checking or unchecking the '**Enabled**' checkbox.

The number of data points can be increased or decreased resulting in more or less detailed datasets. The default is 512 but for T-Jump, it is often necessary to use fewer (eg 256).

Oversampling helps to increase the quality of data and resolution. It has the ability to increase signal to noise by collecting number of data points and averaging them together. By default this is set at 48 but may be reduced to access shorter, faster time scales. Indeed, for T-Jump acquisitions speed tends to be priority so often the oversamples may be set to 1.

Pretrigger can be configured as a percentage of the run time prescribed. It is particularly useful with T-jump to ensure the whole jump is being monitored.

## Data Trigger Offset

The data trigger offset, found in the main '**Control Panel**' > '**Options**', should be set to  $5 \times 10^{-6}$  seconds. This is to allow for the time for the electronics to initiate.

## Time Constants

The time constant,  $t_c$ , is set in '**Control Panel**' > '**Hardware**' is an electronic filter applied to the signal. For a detailed description see the hardware manual.

The time constant must be set so as to reduce the noise in the system while not affecting the signal amplitude. For the fastest reactions the time constant should be set to  $1 \times 10^{-6}$  seconds.

Optimising and maximising the signal to noise ratio is achieved by the appropriate use of the time constant, over-sampling data points and averaging a sequence of (multiple) shots.

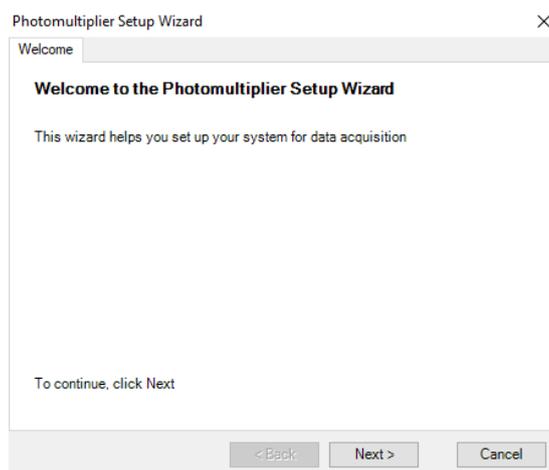
## Recharge Time

The recharge time is found in '**Tools'...'Options'...'TJump Recharge Time'**. This sets the time taken to charge up the capacitor banks and is set to 10 seconds by default.

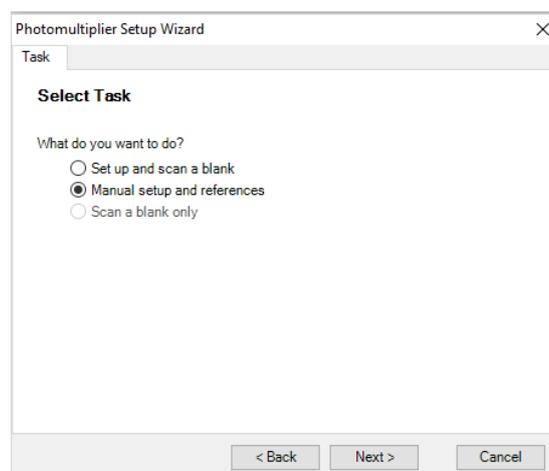
## Setting the References / Spectrometer Manual Setup

This section guides the user through setting up the system optically and enabling data collection.

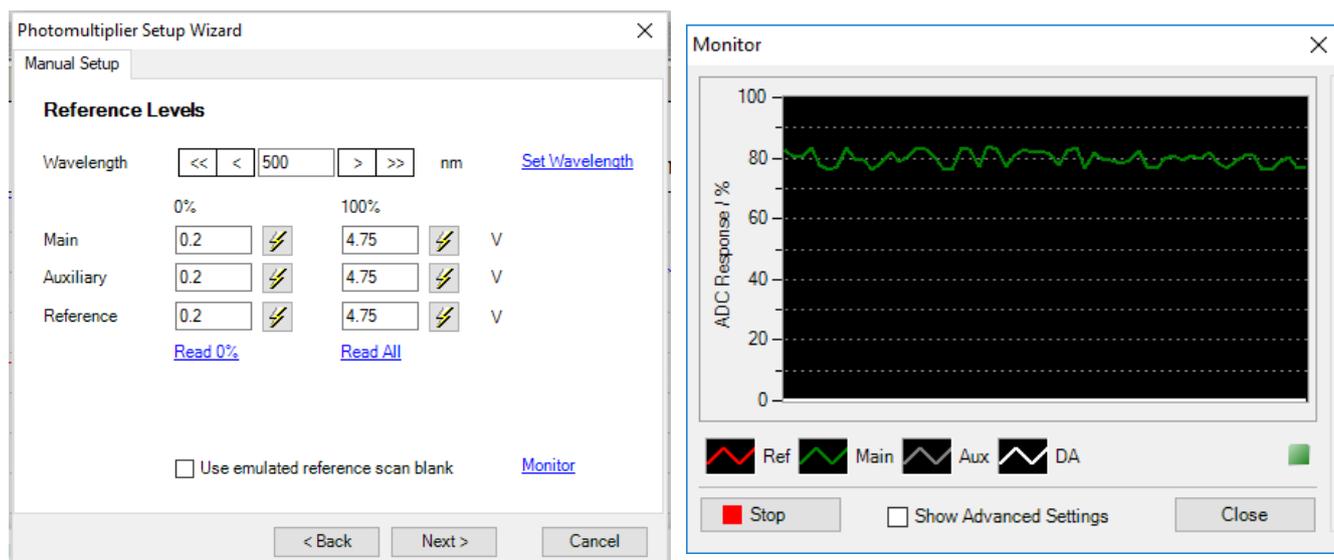
Press the '**Spectrometer**' button to enter the photomultiplier setup routine. This will display the '**Photomultiplier Setup Wizard**':



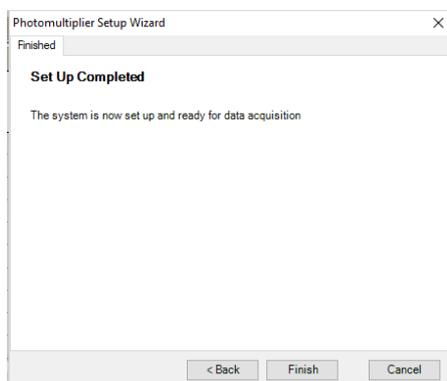
Select the '**Manual setup and references**' to begin a manual setup:



The manual setup panel will be visible along with the live display. This allows the user to set the wavelength and read signals for both 0% and 100% for the main channel. Note: Aux and Ref Channels are mute in T-Jump Mode.



When used for absorbance measurements, 100% (maximum incident light) and 0% (dark condition) transmission reference levels are required to be recorded for a single wavelength. When used for fluorescence measurements, this set wavelength is the **excitation wavelength** and although reference levels can be acquired, normally the Live Display is used simply to maximise the fluorescence signal by optimising the optics and adjusting the PM Volts at this wavelength.



Simply exiting the setup dialogue by pressing '**Next**' and '**Finish**' enables data collection at the set wavelength. Often fluorescence measurements are made without any set references.

### Manual Setup for Absorbance

Specific details for a manual setup for absorbance:

1. Ensure the flow circuit and in particular the observation cell is flushed and as such contains water or buffer solution.
2. Optimise the optics for maximum signal.

3. Increase the PM voltage until the signal trace approaches 80% span.
4. Collect the 0% reference data by either pressing the button(s) next to each channel in the 0% column or enter them numerically, or press the '**Read 0%**' link to collect for both channels. Remember to shutter the light manually for 0% reference levels.
5. Click **Read all** to read the 0% and 100% levels and simply exit the setup dialogue by pressing '**Next**' and then '**Finish**' to enable data collection.

## Manual Setup for Fluorescence

The user will need to have prior knowledge of the excitation & emission wavelengths for the fluorophore to set the monochromator at the correct excitation wavelength and to put the correct emission filter in the optical coupling.

1. Ensure that the fluorescence lens and the appropriate emission filter are fitted in the path of the photomultiplier.
2. Introduce the sample solution to the cell via one of the luer fittings found in the cell block assembly.
3. Open the shutter and increase the (Main Channel) PM volts until the signal level responds.
4. Optimise the optics for maximum signal. The user should note a few points here: the fluorescence may decrease due to photo bleaching – replenishing the cell contents periodically will offset this problem, however, the timescale of the bleaching process is likely to be a relatively slow process. Secondly, the excitation wavelength can be adjusted to find maximum response, especially when using line sources such as mercury/xenon lamps.
5. Increase the PM Volts to set the signal level close to an 80% span. This should accommodate for decreasing or increasing signal levels.
6. Click **Read all** to read the 0% and 100% levels and simply exit the setup dialogue by pressing '**Next**' and then '**Finish**' to enable data collection.

## How to perform a Scan Blank (Absorbance)

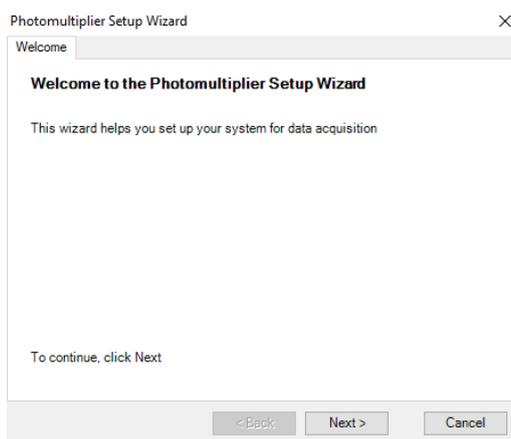
*This mode allows users to set up for absorbance measurements to be taken over a range of wavelengths and are thus able to acquire kinetic data at different wavelengths and/or acquire single scans to characterise reagent spectral information.*

When the system is configured for absorbance measurements, before acquiring new data it is first necessary to align and focus the optics, set the correct photomultiplier voltage and scan a blank. Scanning a blank involves the monochromator scanning a wavelength span acquiring 100% (maximum incident light) and 0% (dark condition) transmission reference levels.

Before performing this operation, ensure that the optical cell contains pure water or a buffer solution.

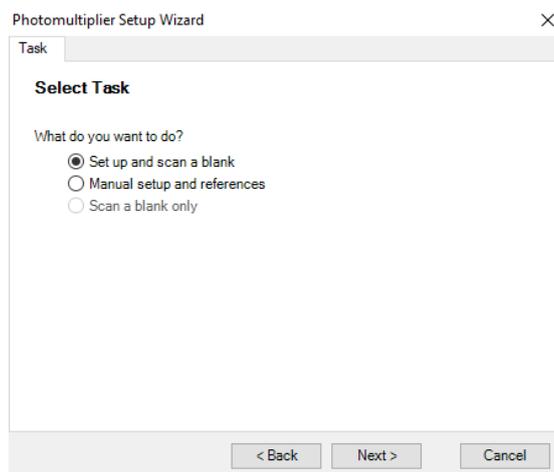
To perform an auto-setup scan blank, perform the following steps:

Press the **'Spectrometer'** button to enter the manual setup mode.  
This will display the **'Photomultiplier Setup Wizard'**.

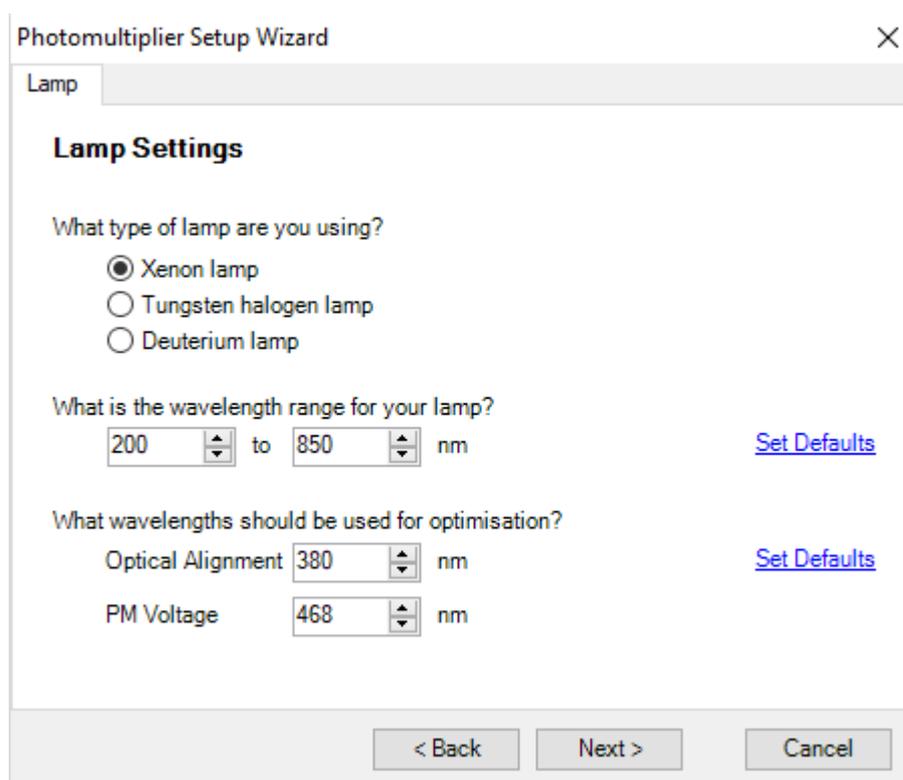


Next the dialogue will offer choice of a scanned set up and a manual set up. The former invokes the use of a scanned blank over a user selected wavelength range, the latter provides for users content to set up at just a single wavelength.

A scanned blank is appropriate for absorption studies only; the manual set up can be used for absorption and fluorescence.



Here, we will begin with selecting the '**Set up and scan a blank**' to begin the wizard driven setup. The '**Lamp Settings**' panel enables the user to select the lamp currently within the system. This in turn automatically fills the wizard with typical wavelength ranges, optimisation and alignment wavelengths for the blank.



The first step is to select the lamp currently fitted to the system.

If the typical values for the wavelength range are not suitable or require adjustment, please edit the desired start and end wavelength spans for the lamp.

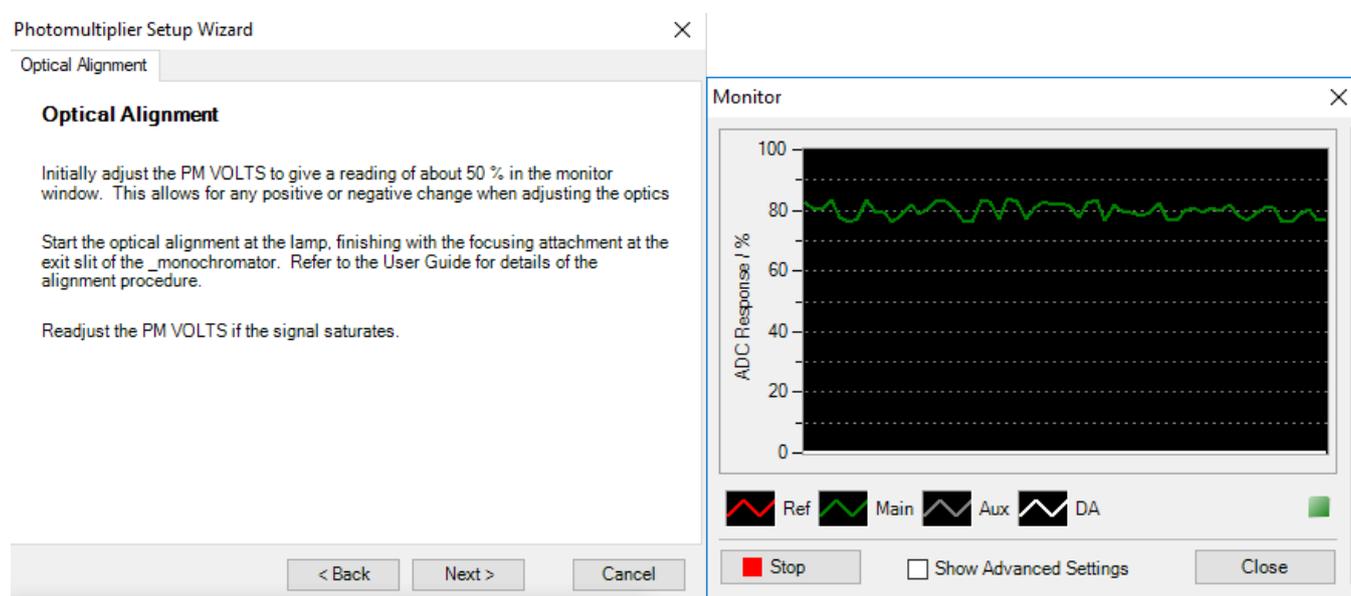
To ensure optimal distribution of the signal over the scan blank wavelength span, the user should optimise the optics at one wavelength, and then maximise the PM volts at another wavelength. This is especially critical when using a wide wavelength span.

To restore the default values for a particular lamp use the '**Set Defaults**' link.

It is often convenient to set the lamp optimisation wavelength to suit the wavelength range where absorbance changes are to be studied.

The edit field labelled '**PM Volts**' is the wavelength where the PM volts are maximised; this ensures that the maximum signal span is achieved under normal circumstances. With the Xenon and QTH lamps, this is set at 480 nm as this is where the system exhibits a maximum signal.

Once the lamp and wavelength ranges have been set, please press the '**Next**' button to proceed with the first stage of optimisation.

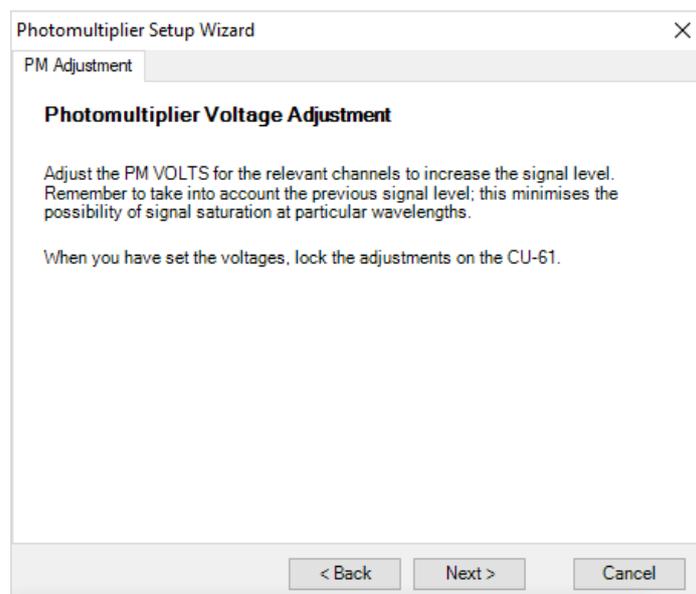


As the '**Optical Alignment**' page appears the monochromator will move to the specified 'Optical Alignment' wavelength as specified on the previous page of the wizard.

To perform the optimisation, open the PM-61s photomultiplier shutters and adjust the PM Volts for the **main channel (green trace)**, increasing the signal at the Live Display to about 50%. This is simply used as a mid-point to allow for positive and negative shifts when the optics are adjusted.

Follow the procedure for optimising the optical components as described in the Section 3 of the TJ-64 User Manual. In brief, adjust the lamp position and its alignment. It may be necessary to adjust the PM volts for the particular channel should the signal saturate.

After finishing the optimisations, return the signal level back to 50% on the Live Display, this is simply used as a relative indication of signal level when moving to the next wavelength.



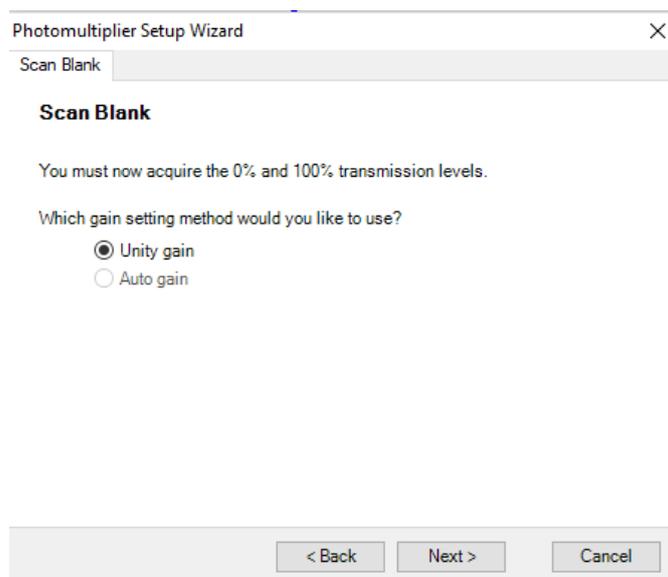
As the '**Photomultiplier Voltage Adjustment**' page appears, the monochromator will move to the adjustment wavelength as specified previously.

Increase the PM Volts for the relevant channels, so setting the signal level(s) to about 80% full scale. You must ensure that the signal does not saturate, ie. go above 100% full scale.

As a quick check it is worthwhile clicking the '**Back**' button to ensure the signals are not saturated at the previous wavelength where the optical optimisation was performed. If they are saturated, go to the next page and lower the PM Volts for the relevant channels.

This concludes the optimisation process. Clicking the '**Next**' button will present the '**Scan Blank**' page.

## Scanning the Blank

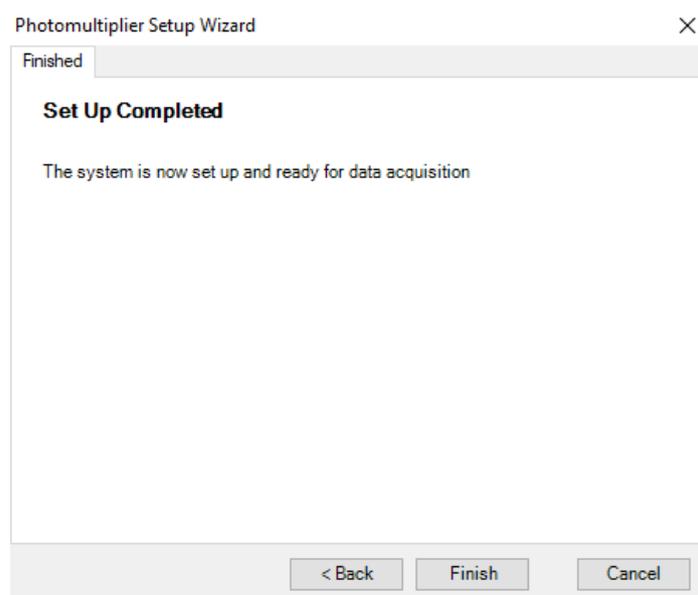


The scan blank process is fully automated.

The process will begin with a full monochromator re-calibration.

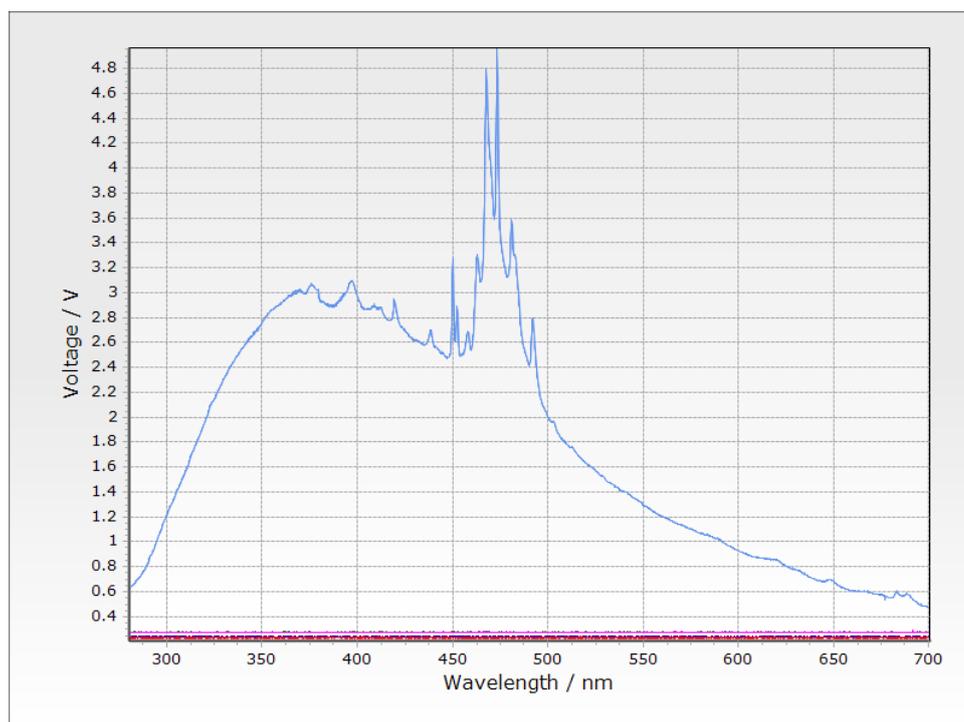
The scan blank process consists of scanning a baseline with no light (0%) and with the filter wheel open (100%). Whilst traversing through the wavelength range, the system will automatically insert appropriate filters.

After both the 0% and 100% scans have completed, the system will acquire static baseline references and present the '**Set Up Completed**' panel.



The system is now ready for use.

An example of a scan blank collected with the xenon lamp:



## T-Jump Shot Sequence

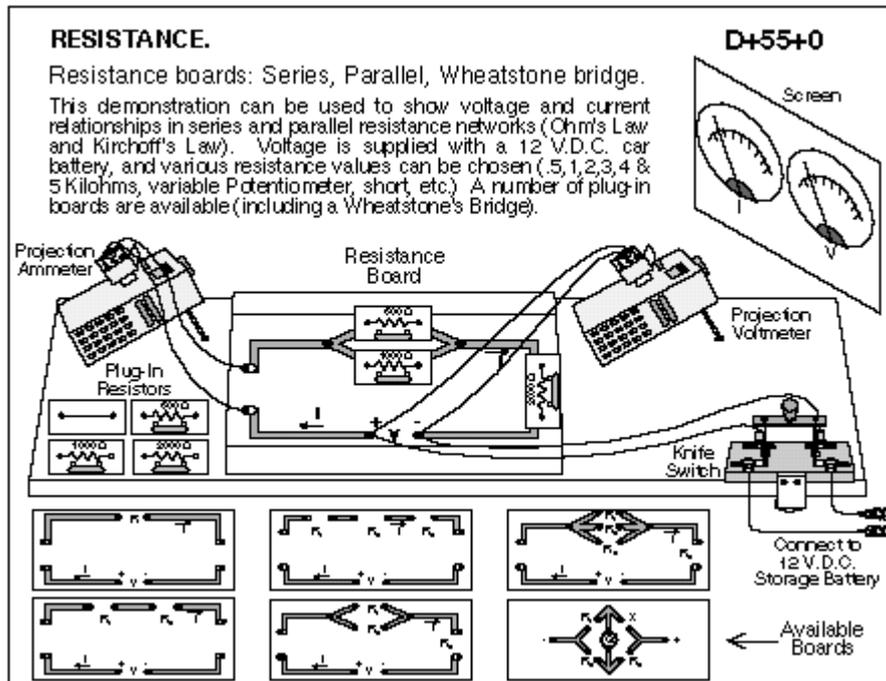
A full T-Jump system is able to perform automated shot sequences.

To learn how to use this facility, please refer to **Time Delay Sequence** on page 86

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# Conductivity Mode



## Conductivity Description

The conductivity cell is a cylindrical cavity in a teflon block, 3mm long and 3mm diameter.

The ends are closed by platinum electrodes. The nominal cell volume is 21 $\mu$ l, and the nominal cell constant 4.24cm<sup>-1</sup>. Reagents are mixed in a T mixer just upstream of the observed volume.

The conductivity meter has the following ranges:

Unit: Siemens (S)

Range 1 0 to 0.5

Range 2 0 to 0.05

Range 3 0 to 0.005

Range 4 0 to 0.0005

Range 5 0 to 0.00005

Range 6 0 to 0.000005

The full scale voltage for each of these is 10V although Range 1 may be limited to 250000 $\mu$ S at 5V - linearity above this level should not be assumed without calibration. Furthermore, operation at cell conductivities above 250000 $\mu$ S should not be attempted at ambient temperatures above 30 degrees C in order to avoid excessive dissipation from the amplifier.

The bias control enables the user to "backoff" up to 10V of signal; this means that signals up to the full scale output of 10V can be offset so that any signal change can be digitised within a 0 - 5V analogue channel input range.

## How to Setup for an Experiment in Conductivity Mode

### Overview

The Control panel can be displayed by selecting the new document icon on the shortcut bar or by selecting '**Control Panel**' under the '**Acquire**' menu.

The configuration of Kinetic Studio will depend on the type of experiment and whether the software is controlling a Conductivity option for the KinetAsyst Scientific Stopped-Flow instrument or whether it's acting as a data acquisition suite for the dedicated Conductivity apparatus.

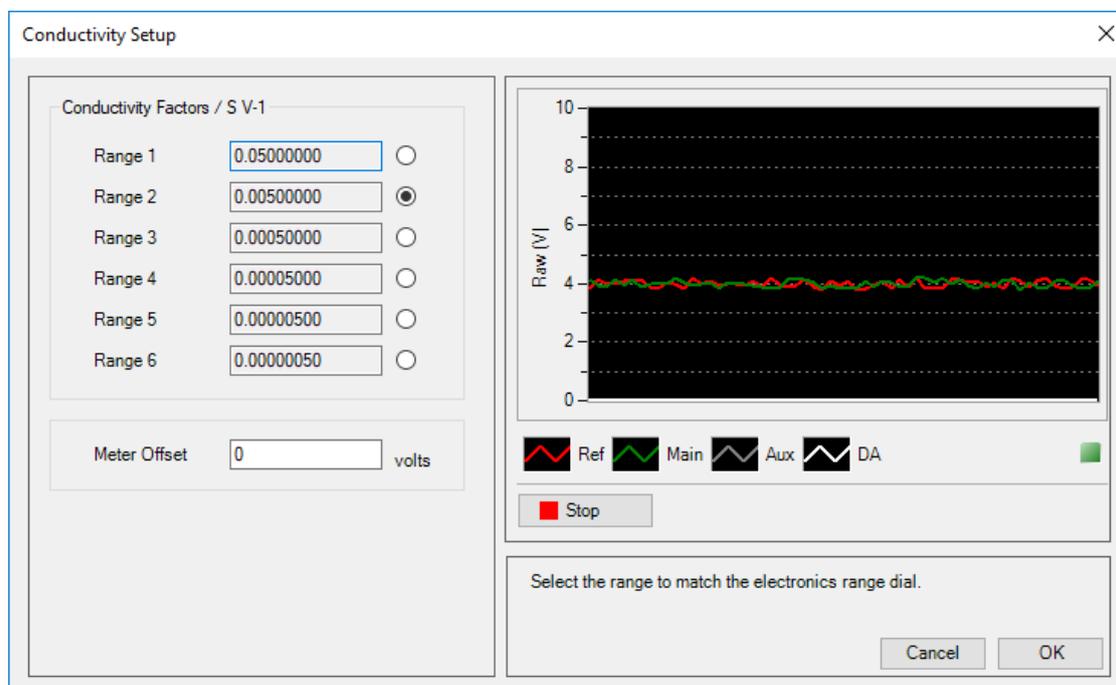
For installation and configuration with the KinetAsyst Stopped-flow system refer to the SF-61DX2/SX2 User Manual, OPT-642 - Conductivity Detection.

For the dedicated Conductivity apparatus, refer to the CSF-21 User Manual.

## Initial Setup

Ensure that the Conductivity meter is switched on for at least 30 minutes to ensure it has stabilised.

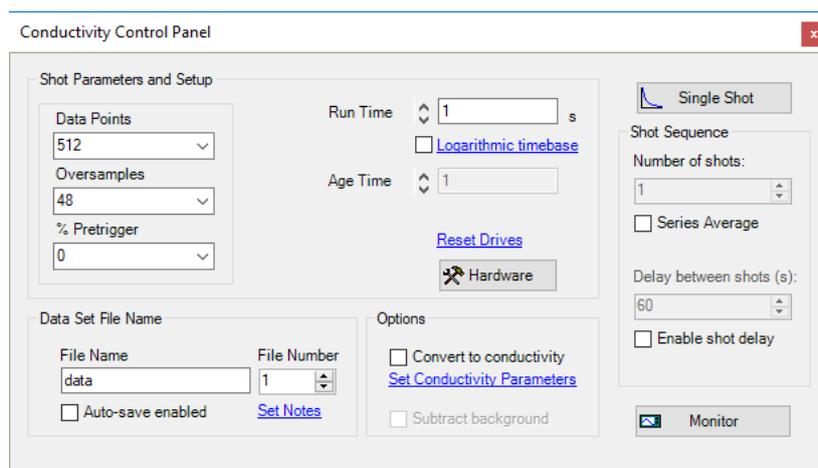
At the Conductivity Control Panel, select '**Set Conductivity Parameters**', a dialog incorporating a live display will appear.



Use the live display to set the RANGE and METER OFFSET at the Conductivity meter. Make sure that solution is pushed through to the mixing cell and start with the RANGE set at 1 and the OFFSET set at 0. Increment the RANGE and observe the signal, do so until the signal maximises. Note that incrementing to the next range causes the signal to fall again and become very noisy - this is because the amplifier has been driven into saturation.

Having adjusted the signal with the selection of the optimal range and meter offset, record these settings by selecting the corresponding '**Range**' check and enter the applied '**Meter Offset**', this is necessary to ensure subsequently acquired data is converted correctly.

## Setting the Dataset Parameters



The first step in setting up Kinetic Studio for an experiment is to configure the number of data points required, oversampling and pretrigger.

The number of data points relates to the final dataset and graph. This can be adjusted depending on the experimental requirements and trace detail. The typical number of data points used in an experiment is 512.

Oversampling can be used to improve signal to noise and hence the quality of data. For every data point, oversampling corresponds to the number of additional samples that are averaged together.

For example, the screenshot above shows 512 data points have been specified with 48 oversamples. This means the data acquisition device will acquire 512 x 48 samples (24576 samples in total). Every data point is an average of 48 samples. The software will automatically perform the averaging after the experiment has completed.

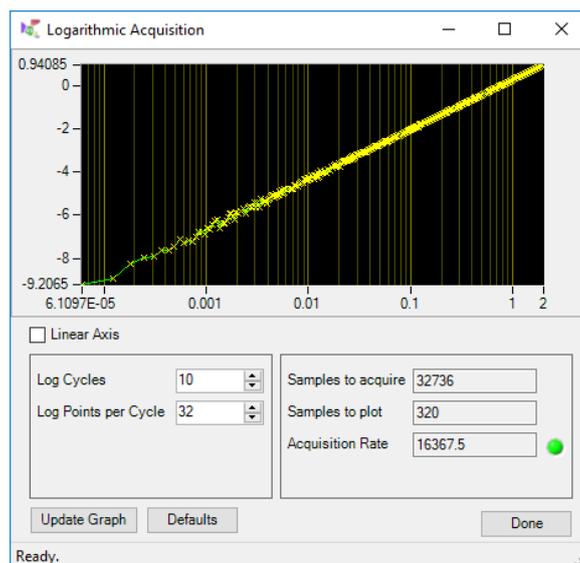
Pretrigger can be used to acquire data just prior to the trigger point. The amount of pretrigger is specified as a percentage of the run time.

### Setting the Run Time

The experimental run time can be manually entered by editing the '**Run Time**' numeric box or alternatively a series of standard run times can be applied by clicking the small up and down arrows next to the '**Run Time**' box.

For shots requiring a log acquisition and hence being displayed with a logarithmic x-axis, the '**Logarithmic timebase**' option can be enabled. The standard log timebase applied is 10 log cycles with 64 points per log cycle. Kinetic Studio intelligently uses the additional samples acquired for logarithmic processing to apply data averaging improving the signal to noise.

Should the experiment require it, a custom log mode facility can be accessed by pressing the link '**Logarithmic timebase**'. This will display a log parameter editing utility as shown.



If the Conductivity device being used is an option for the Stopped-Flow system, the ability to edit the **'Age-Time'** will be made available.

### Specifying a file name

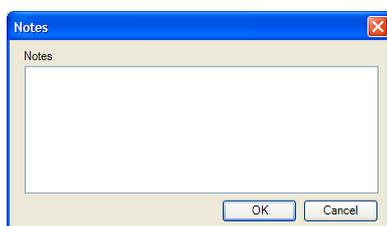
Within the **'Data Set File Name'** area to the bottom left of the control panel, enter a file name that corresponds to the experiment being performed. If required (generally recommended), create a folder for the group of experiments being performed and save all related data into that folder.

Kinetic Studio provides a convenient facility to specify a working folder.

Please consult the **'How to Change the Current Working Folder'** subsection within the **'Dataset File Management'** section.

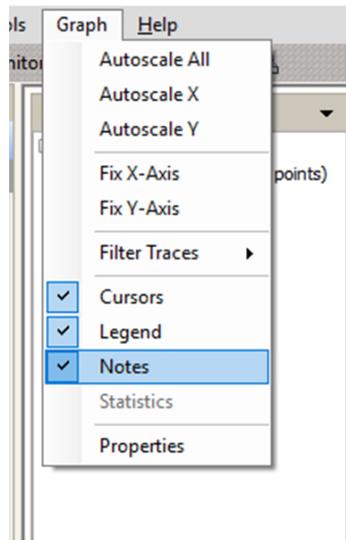
### Adding Notes to a Dataset

Clicking the **'Set Notes'** link in the file name group will display a small notes editor.

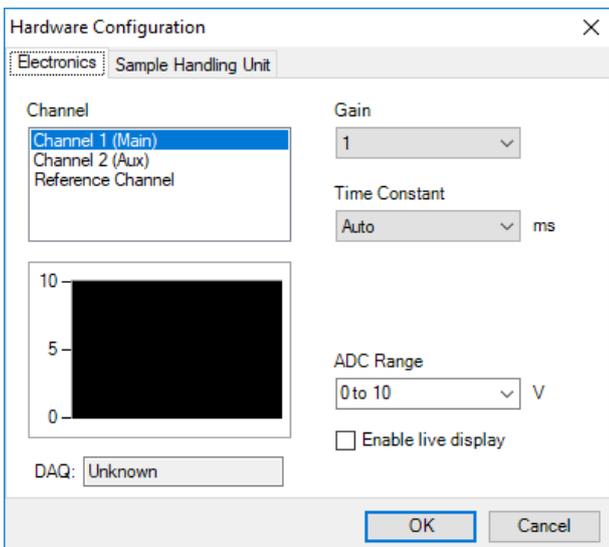


The notes editor provides a facility to save experimental information or comments with a dataset. These notes will be applied to every shot.

Notes for a given dataset can be displayed and edited by selecting the **'Notes'** menu item under the **'Graph'** menu as shown below.

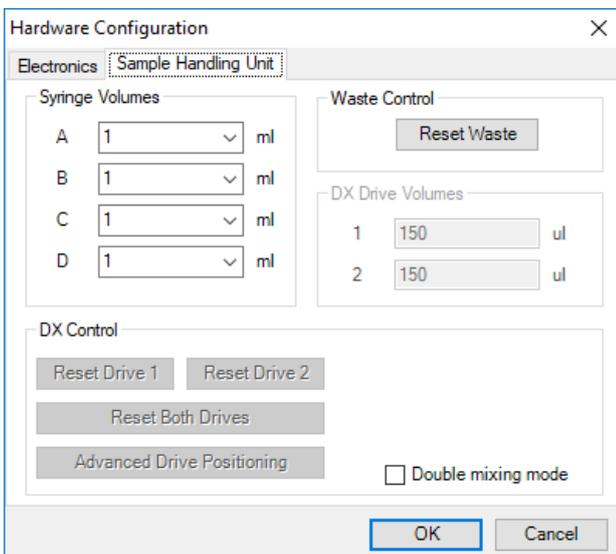


## Adjusting Hardware Options and Signal Conditioning



Some hardware options are only available for the Stopped-Flow Conductivity option.

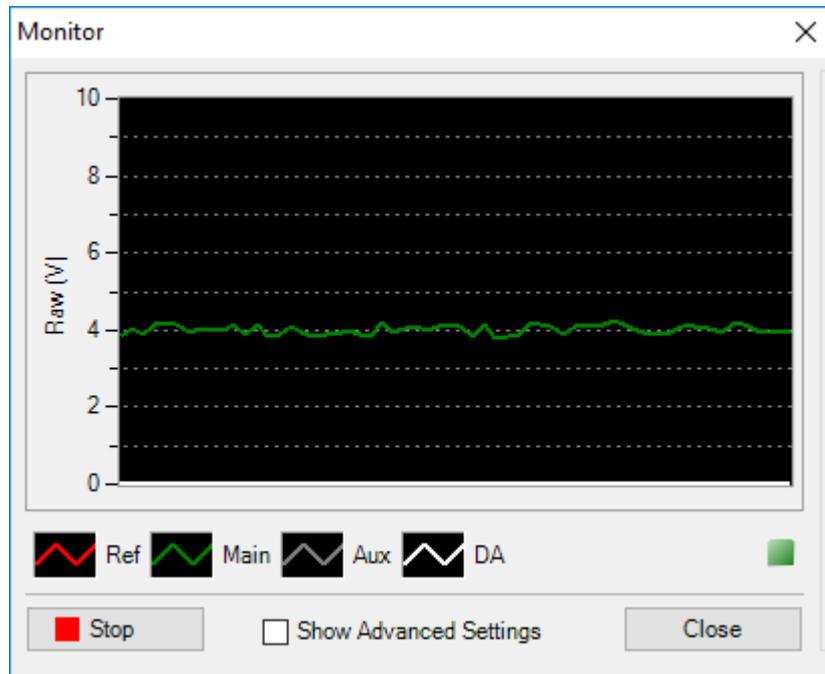
When the full KinetAsyst Stopped-Flow instrument is present, **'Hardware Configuration dialogs'** will be available.



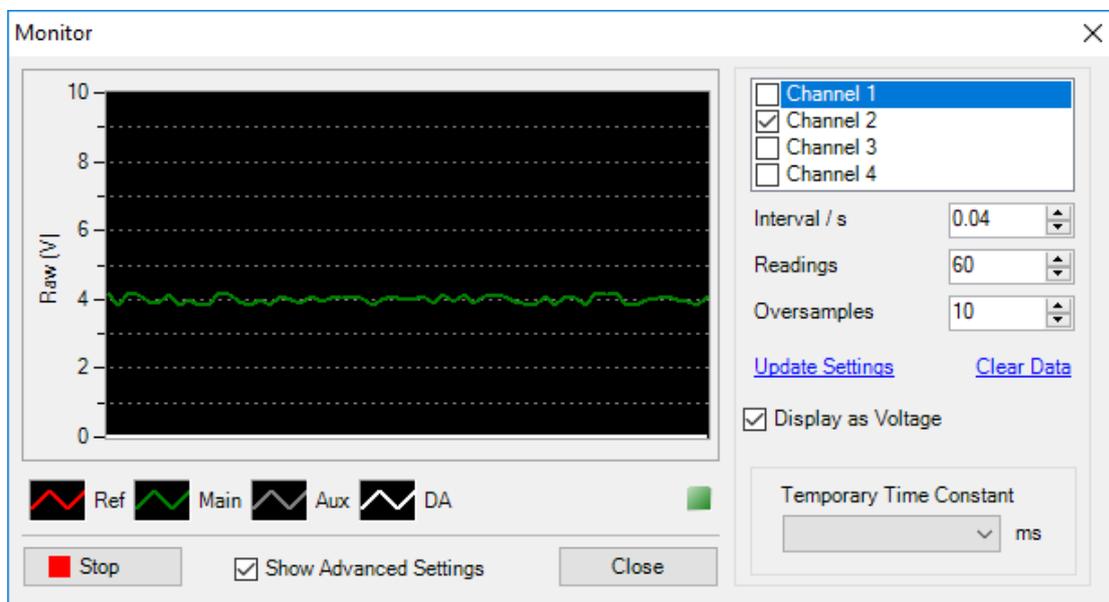
Syringe and drive volumes can be configured.

Single or double mixing modes can be enabled depending on the hardware available.

The control panel provides convenient access to a live display / monitor. This facility enables a live data acquisition window displaying the current voltage levels per channel.



The advanced settings panel provides the following options:



Generally it should not be necessary to use the Advanced monitor settings.

## How to Acquire a Single Conductivity Shot

Having configured the conductivity meter and set the appropriate acquisition parameters at the control panel, next

**Important:** Ensure the drive syringe plungers are in contact with the push plate. Failure to do so may result in damage to the flow circuit should the drive be actuated.

1. If the syringes are air driven, ensure the drive air pressure is set correctly.
2. Set the appropriate stopping volume.
3. Ensure the reactants are loaded into the drive syringes.
4. Set the Drive/Fill valves are set to the Drive position.
5. Where required and if it hasn't been done already, set the appropriate number of **'Data points'**, **'Oversamples'**, etc. for the experiment.
6. Set the data file name and number under **'File Name'** and **'File Number'** on the Control Panel.
7. Prescribe the desired run time for the acquisition, checking the option for Logarithmic time base if required.
8. When you are satisfied with your settings, click the **[Single Shot]** button on the Acquire Control Panel to arm the data acquisition.

The acquisition progress box will appear.

Where the Conductivity option is installed and being used with the full KinetAsyst Stopped-flow system, the instrument control features will automate the entire shot sequence.

For dedicated Conductivity apparatus, the acquisition progress box will include a message **'Awaiting Trigger'**. Where upon the user should initiate a shot by activating the air drive.

9. Where the stop syringe hits the stop block data acquisition starts.

## How to Acquire a Conductivity Shot Series

Kinetic Studio provides a convenient option for collecting a series of data shots and averaging them.

To enable the option ensure there is a check mark in the '**Series Average**' option in the '**Shot Sequence**' group box. When activated the shot button will change to **[Series]**.

The number of shots in the series can be changed by either manually entering a numeric value, or by using the small up and down arrows associated with the '**Number of shots**' edit field.

**Note:** The dedicated Conductivity apparatus only holds a limited amount of sample within the thermostated block. Check the '**Enable shot delay**' box and use the '**Delay between shots**' field to set an equilibration time.

**Important:** Ensure the drive syringe plungers are in contact with the push plate. Failure to do so may result in damage to the flow circuit should the drive be actuated.

1. If the syringes are air driven, ensure the drive air pressure is set correctly.
2. Set the appropriate stopping volume.
3. Ensure the reactants are loaded into the drive syringes.
4. Set the Drive/Fill valves are set to the Drive position.

5. When you are satisfied with your settings, click the **[Series]** button on the Acquire Control Panel to arm the data acquisition.

The acquisition progress box will appear

Where the Conductivity option is installed and being used with the full KinetAsyst Stopped-flow system, the instrument control features will automate the entire shot sequence.

For dedicated Conductivity apparatus, the acquisition progress box will include a message '**Awaiting Trigger**'. Where upon the user should initiate a shot by activating the air drive.

6. Data acquisition is initiated when the stop syringe hits the stop block.
7. On completion of the acquisition process, data appears in the graph window and the sequence is repeated. The acquisition progress bar will appear for each shot. This sequence will continue to be repeated for the programmed number of shots.

If during the shot series the drive syringes reach their full span, the drive syringes will have run out of solution. This results in the current shot not completing and hence the acquisition will not have been initiated. Cancel the current shot and reload the drive syringes.

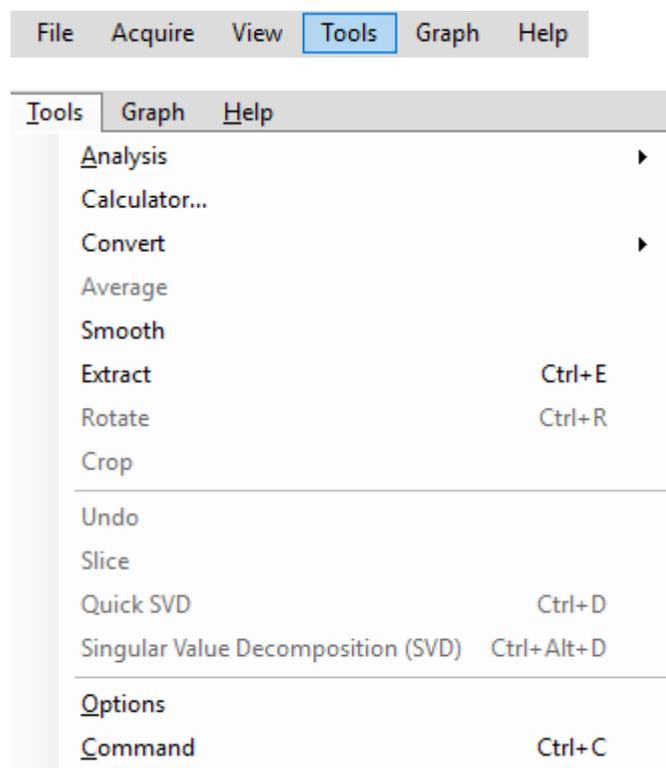
For the Conductivity option being used with the full KinetAsyst Stopped-flow system, the software will either wait for the shot to be cancelled or a prompt will appear depending on the mixing mode, where upon the drive syringes can be re-loaded and the shot series continued when ready. During the process, there is also the option to abort the shot series altogether.

8. On completion of the shot series, the newly displayed dataset will show the overlaid shot data and include the determined average

# Data Manipulation

## Introduction

Data manipulation functions can be found in the Tools menu.



The functions available will depend on the type of dataset.

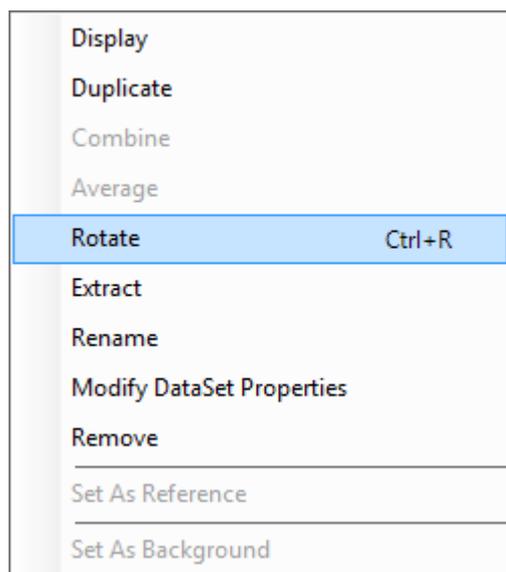
For convenience conversion operations can be quickly accessed via the '**Convert**' sub menu.

Several functions can also be accessed via the thumbnail right-click menu.

## Rotating a Dataset

Three dimensional datasets can be conveniently rotated using the '**Rotate**' option shown in the main '**Tools**' menu above or by using the keyboard shortcut '**Ctrl**+'**R**'.

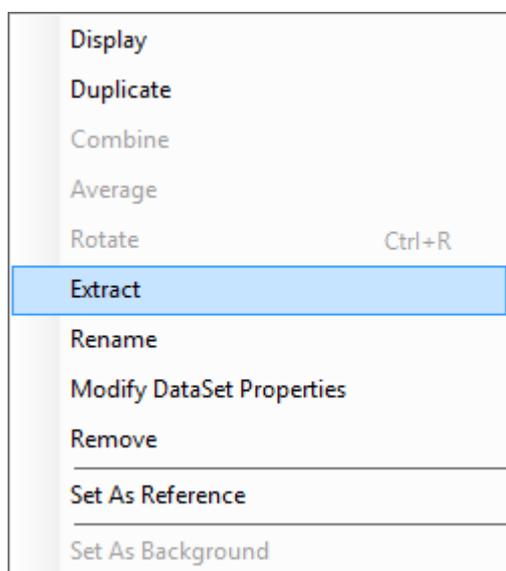
Additionally a new context menu item has been added to the thumbnail navigator allowing single or multiple datasets to be rotated.



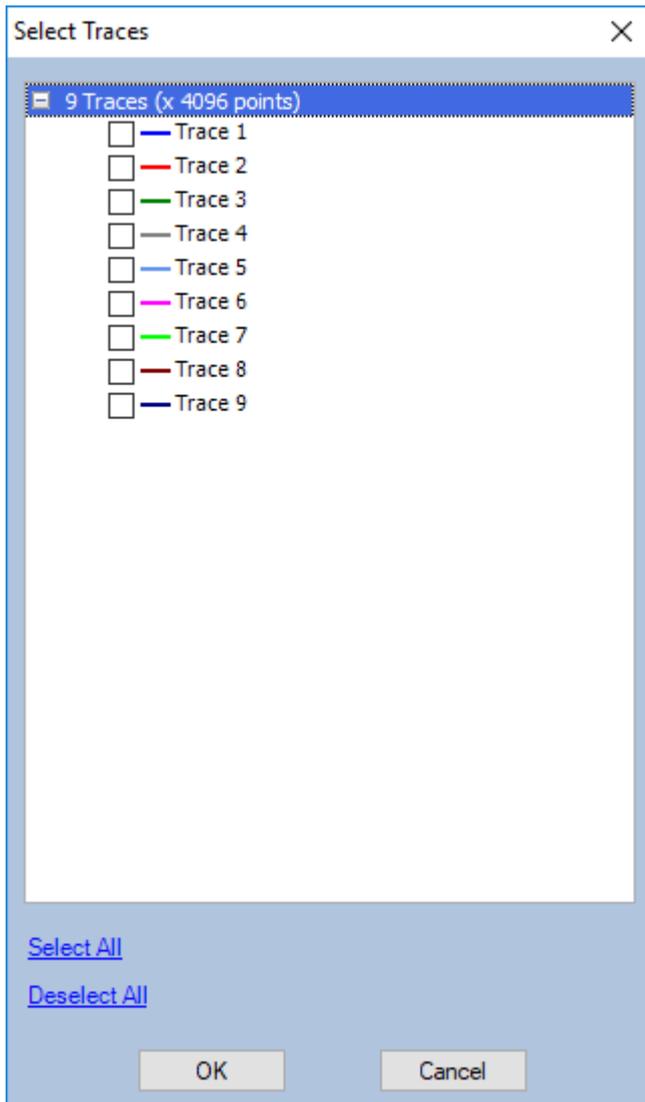
## Extracting Traces from Datasets

There are three ways to extract traces from a dataset.

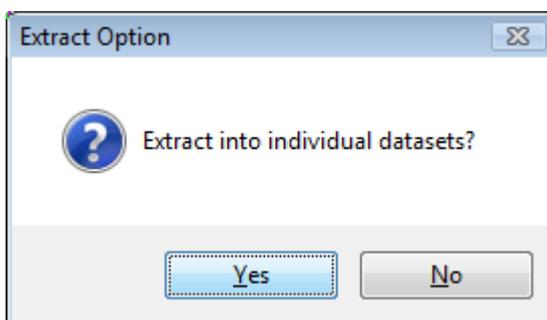
The first method accessible via the **'Tools' ... 'Extract'** menu item (or by pressing the **'Ctrl' + 'E'** shortcut key) provides a list of all traces within the currently displayed dataset. Each trace can be selected or deselected which in turn determines what is copied into a new dataset.



The trace selection dialog will be displayed. This allows the selection of which traces should be extracted.



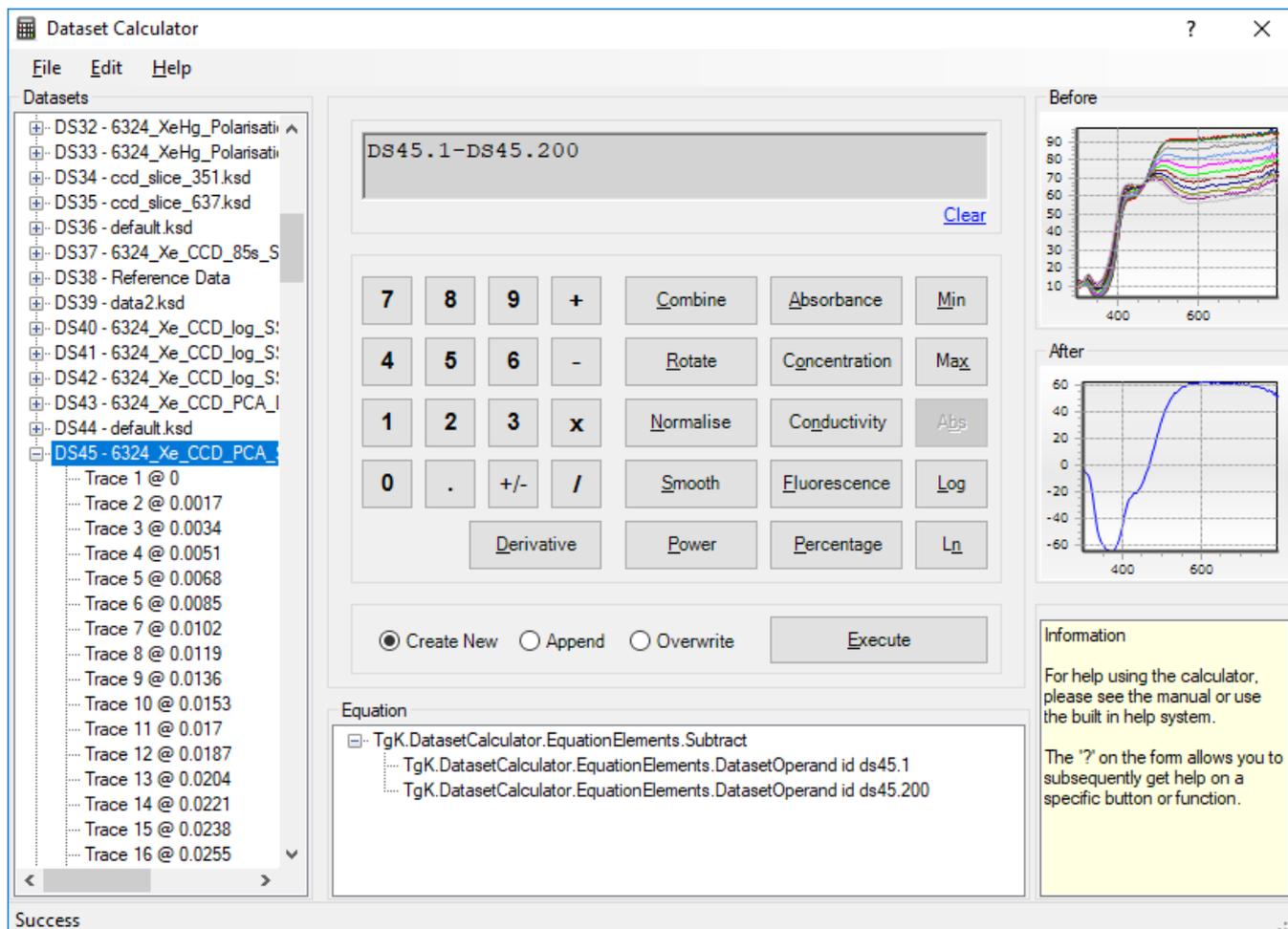
The traces can be extracted into a single new dataset, or each trace can be placed into a new dataset.



## Dataset Calculator

Kinetic Studio provides a convenient dataset calculator for data manipulations. This allows mathematical expressions to be entered and applied to data on all applicable axes.

The maths engine uses a script processor allowing complex calculations to be entered if required. These scripts may then be saved and recalled at a later date.

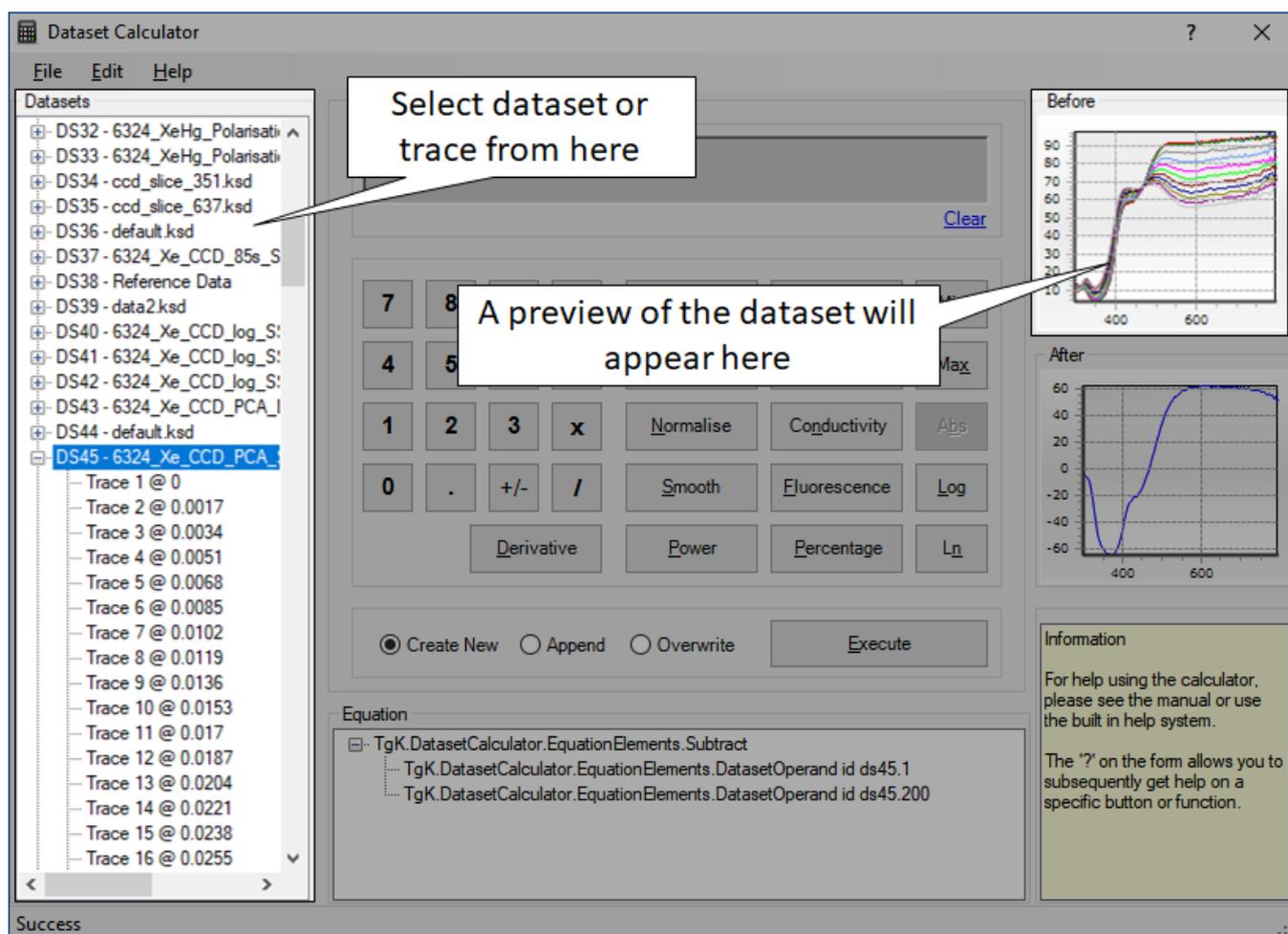


The left hand panel within the Dataset Calculator provides a list of available datasets. Each dataset can be expanded to show the contained traces.

Calculations can be applied per dataset or per trace.

## Previewing a Dataset

To preview a graph of the dataset, select either a dataset or trace within the left-hand dataset list. As soon as something is selected, a 'Before' preview of the dataset or trace will appear on the right hand side.



## Entering Datasets or Traces into the Calculator

Traces or datasets can be entered into the calculator in two different ways.

The first method is to double mouse-click a dataset or trace. The calculator will enter the details into the edit window. A dataset entry consists of the dataset identifier DS# so in the above example, DS1 would represent the whole dataset.

If a specific trace within the dataset is to be manipulated, this will be displayed as DS1.# where DS1 represents dataset with ID 1 and # is the trace number. For example, DS1.1 means dataset 1, trace 1. DS2.5 would represent dataset 2, trace 5.

If a dataset is entered with a trace number of 0, this also represents the whole dataset.

Further examples,

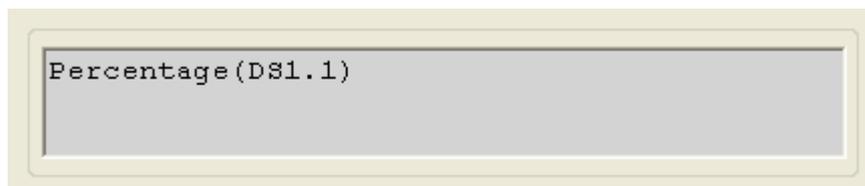
DS1 represents dataset 1, all traces.

DS1.0 represents dataset 1, all traces.

DS1.4 represents dataset 1, trace 4.

The second method of entry is to type the dataset details into the editor using the keyboard.

After entering the dataset or trace, the trace can be manipulated by pressing one of the function buttons or by entering an equation. Example,



After entering a calculation, a preview of the resulting dataset will be generated within the 'After' preview panel.

When ready, select the option to either Append the result into an existing dataset or 'Create New' dataset. Then press [Execute] to perform the calculations.

## Calculator Functions

Below is a summary of the various calculator functions and how to use them.

### Copying a Dataset

The calculator offers a fast and convenient way of copying a dataset and then working on the copy. To do this simply enter the dataset into the calculator and press [Execute]. A copy of the dataset will then be generated.

### Extracting a Trace

The dataset provides a quick facility to extract a trace into a new dataset. To do this simply enter the dataset and trace details into the calculator. Example, DS1.2 will take a copy of trace 2 from Dataset 1 and place it into a new dataset.

### Convert a Dataset Type

Kinetic Studio provides convenient options to convert the data type.

For more information about absorption of light, concentration, extinction coefficients and their relationship in Beer Lambert's law please see the reference manual.

The following conversions are available depending on the original data type and compatibility:

#### **Convert to Absorbance**

This allows the dataset to be converted to absorbance units.

Syntax: Absorbance(DS#)

- The signal type must be Transmission

Supported Ordinates:

- Raw
- Concentration
- Percentage

## Convert to Concentration

Convert to a concentration dataset using Beer's Law relationships.

Syntax: Concentration(DS#,ConcentrationFactor)

ConcentrationFactor is Absorbance / Molar Absorptivity

Supported Ordinates:

- Raw
- Absorbance
- Percentage

## Convert to Conductivity

Convert raw conductivity data into processed conductivity datasets.

Syntax: Conductivity(DS#,Range,Factor)

Syntax: Conductivity(DS#,Range,Factor,Span,ZeroLevel)

The second syntax should only be used with 'Percentage' data, typical of KinetAsyst conductivity data.

- Raw
- Percentage

## Convert to Fluorescence

This function allows fluorescence polarisation datasets to

Syntax: Fluorescence(DS#)

The data must be fluorescence polarisation type.

## Convert to Percentage

Convert a dataset from absorbance units to percentage.

Syntax: Percentage(DS#)

Supported Ordinates:

- Absorbance
- Concentration

## Combine

Combine multiple datasets or traces together into a new dataset.

Syntax: `Combine(DS#.#,COMMA_DELIMITED_DATASETS)`

Examples,

`Combine(DS1,DS2,DS3)`

`Combine(DS1.2,DS2.7, DS4.2)`

## Rotate

Rotate a 3D dataset from wavelength to time or time to wavelength.

Syntax: `Rotate(DS#.#)`

Examples,

`Rotate(DS3)`

`Rotate(DS3.0)`

## Normalise

This processes the y axes data so that the new range is factored by the new upper and lower limits. The resulting data is re-scaled.

Syntax: `Normalise(DS#.#,LOWER,UPPER)`

Example,

`Rotate(DS2.3,0,2)`

## Smooth

The smooth function uses the Savitzky-Golay smoothing filter. This method performs polynomial regression of degree  $k$  on a distribution of at least  $k+1$  equally spaced data points. This approach helps to preserve the features of a distribution.

Reference: The Savitzky-Golay smoothing filter was created by Abraham Savitzky and Marcel Golay in 1964. "Smoothing and Differentiation of Data by Simplified Least Squares Procedures". Analytical Chemistry 36 (8): 1627–1639.

Syntax: Smooth(DS#. #,SMOOTHINGPOINTS)

Example,

Smooth(DS3,13)

## Power

This is otherwise known as exponentiation. It is written as  $a^n$ . Where  $a$  is the base and  $n$  the exponent.

Syntax: Power(DS#. #,EXPONENT)

Example,

Power(DS8,2)

## Derivative

This executes differentiation to a given 'ORDER' on a dataset or trace. Differentiation calculates the rate of change of  $y$  with respect to  $x$  hence the derivative is a measure of how a function  $y$  changes as its input  $x$  changes.

Syntax: Derivative(DS#. #,ORDER)

Example,

Derivative(DS12,3)

**Min**

Find the minimum value in a given dataset or trace.

Syntax: Min(DS#. #)

Example,

Min(DS10)

**Max**

Find the maximum value in a given dataset or trace.

Syntax: Max(DS#. #)

Example,

Max(DS8)

**Abs**

Calculate the absolute value of floating point number.

Syntax: Abs(VALUE)

Example,

Abs(-2.65)

**Log**

Calculate the log of a number, dataset or trace to a given base.

Syntax: Log(DS#. #)

Syntax: Log(DS#. #, Base)

Syntax: Log(Number)

Syntax: Log(Number, Base)

Examples,

Log(DS20);

Log(4, 10)

## **Ln**

Calculate the natural log of a number, dataset or trace.

Syntax: Ln(DS#.#)

Example,

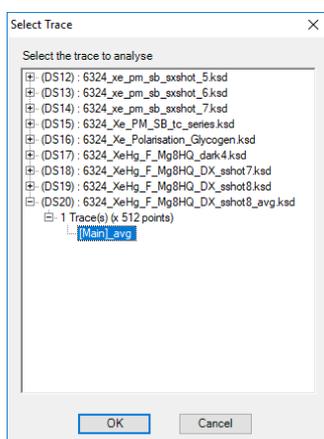
Ln(18)

# Data Fitting

## Introduction

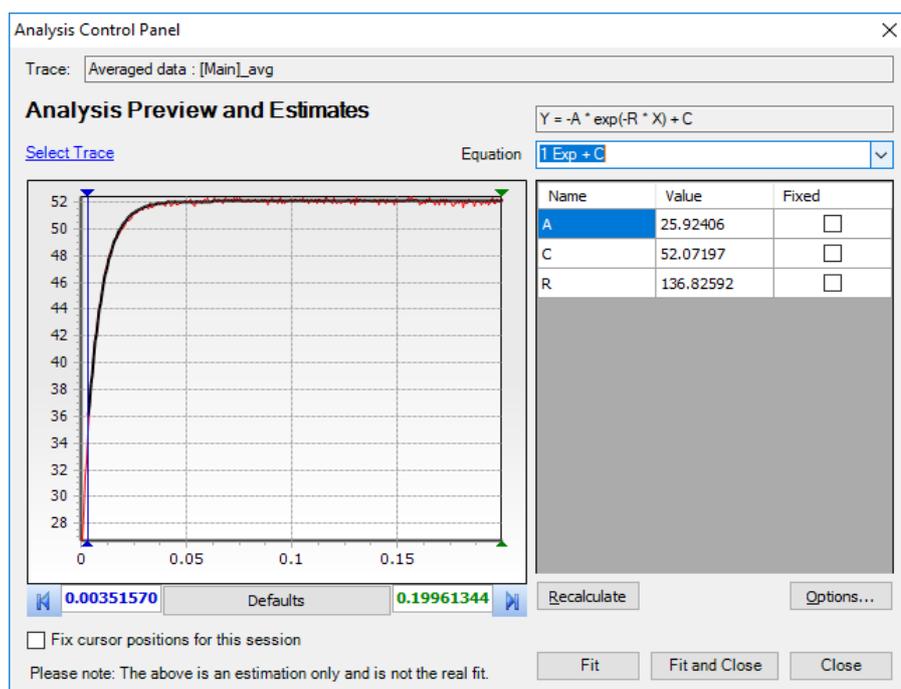
The Analysis Control Panel provides easy access to switch to different traces (Select Trace) and to fine tune the fitting parameters. A number of pre-defined models are included as standard and are available via the 'Equation' drop-down box. In addition to the standard models, a user definable equation editor is available.

## Fitting a Dataset with a Standard Model



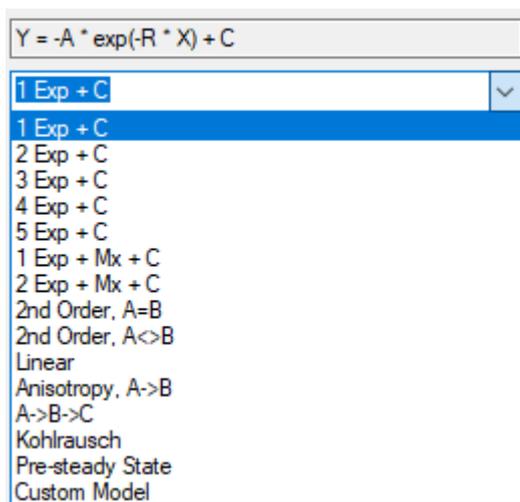
Kinetic Studio will ask which trace to fit. This is done by displaying a summary screen of all datasets in memory.

After selecting the trace, the Analysis Control Panel will be displayed.



The Analysis Control Panel will attempt to calculate parameter estimates automatically for the user based on the selected equation – these values can be adjusted if required, and they can be **'Fixed'**, i.e. the values are constrained by checking the **'Fixed'** options.

The fitting equation can be changed by the **'Equation'** drop down menu. All common models are included as well as the Custom Model function to allow more exotic systems to be examined. See **Custom User Definable Equations** on page 141 for more information.

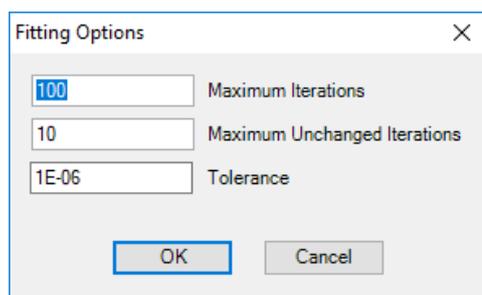


The graph displays the chosen trace and an estimated fit.

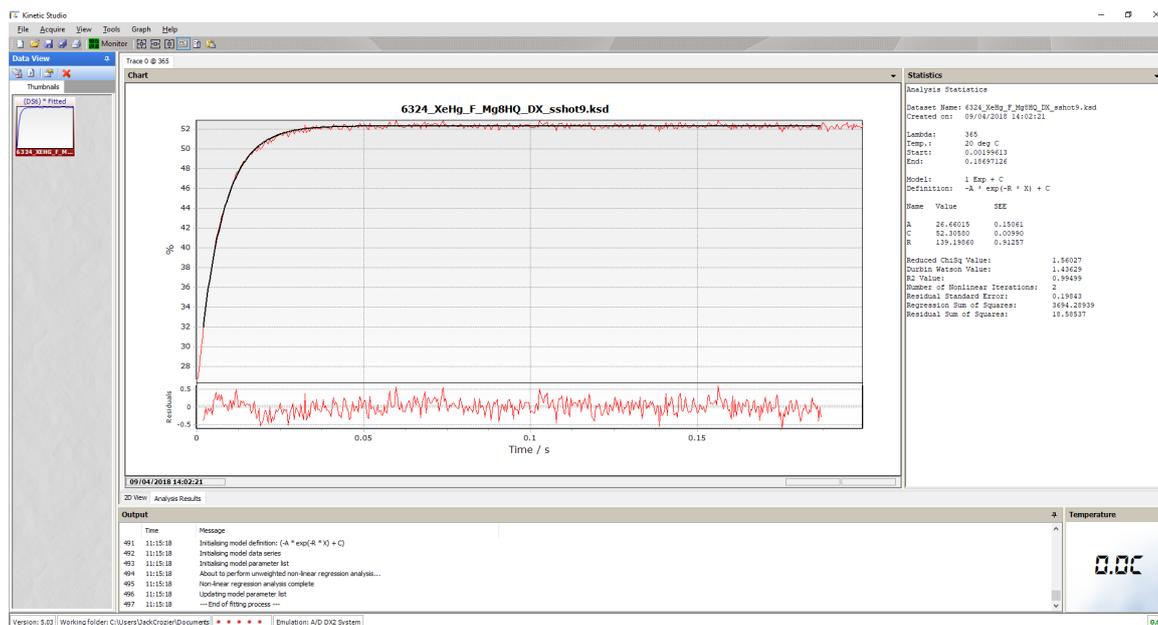
Please note that the estimated fit may or may not appear to be accurate. This will depend on the model and the complexity of the fit.

To perform a fit, please use the following guidelines:

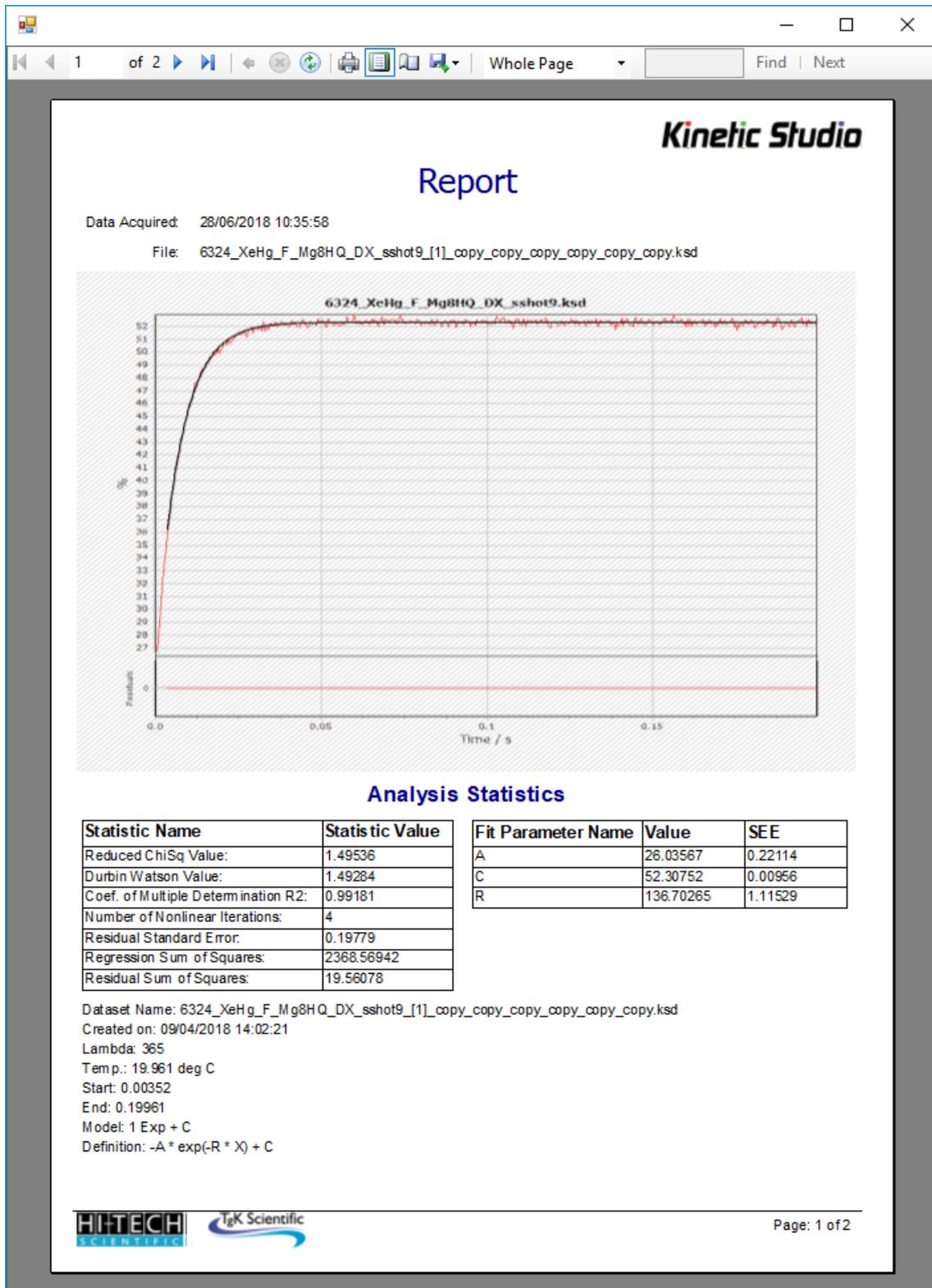
1. Move the blue and green range lines on the graph to indicate the fitting area. Numbers can be manually entered into the blue and green start and end fields.
2. If a number of traces are going to be fitted, the option to '**Fix cursor positions for this session**' can be enabled.
3. Pick the appropriate model from the drop-down equation list. In the example above, a third order model has been selected.
4. The Analysis engine should attempt to estimate the fitting parameters and show an estimated fit. Depending on how good the estimate is the initial parameters can be adjusted.
5. Press either the **[Fit]** or **[Fit and Close]** button to try analyse the data and execute the fitting operation.
6. If the fitting process fails, the '**Options**' dialog can be used to adjust the maximum number of iterations and tolerance of the fit.



7. Once a successful fit has been completed Kinetic Studio will display the result and all related statistics.



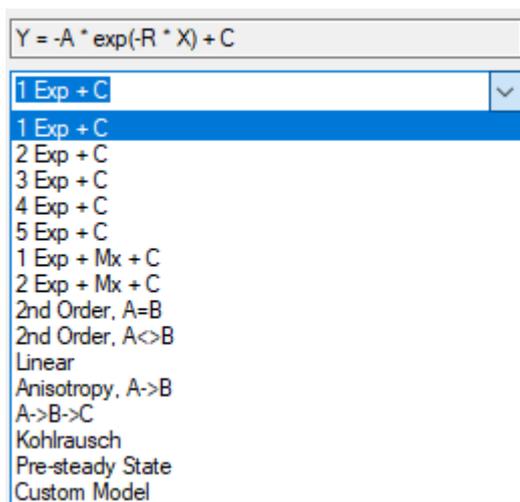
- The fit results may be printed using the built in reporting system using the 'Print' option in the menu from right-clicking on the graph.



## Custom User Definable Equations

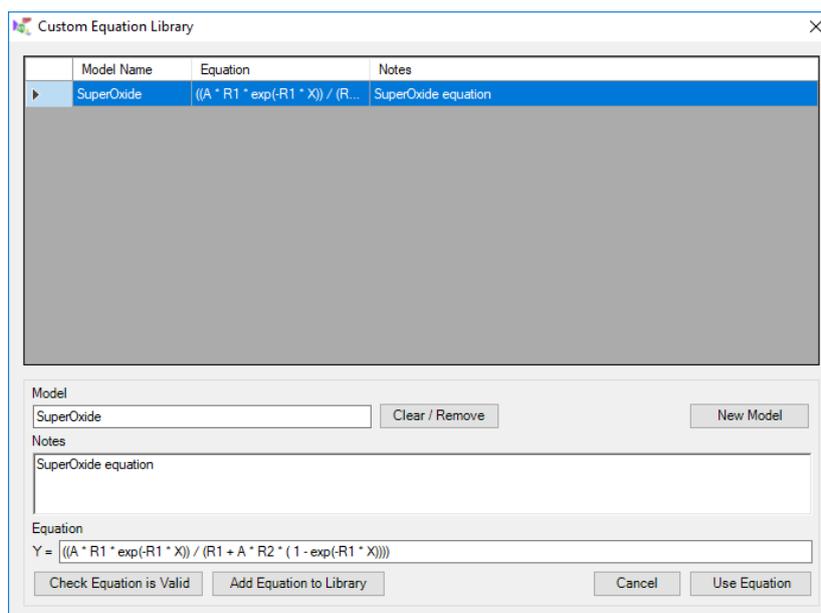
Kinetic Studio supports the ability to define custom fit models. This allows the user to define a mathematical equation for fitting and to use non-linear least squares fitting to try and generate a fit to the experimental data.

To access the custom equation library, the last entry in the built-in equations list is 'Custom Model' as shown in the screenshot below:



This will display the Custom Model editor and library. A model that has been previously defined can be selected, or a new model can be defined. The equation for the SuperOxide reaction is set up as an example.

When generating a new equation model, Kinetic Studio will attempt to validate the model before adding it into the library.



## Entering a New Custom Model

When defining a custom model, parameter names should use the following guidelines:

- A parameter name must begin with an alphabetic character a-z, upper or lower case.
- Parameters are case sensitive. E.g. param1 is not the same as Param1
- The independent variable must be t, T, x or X
- The dependent variable y must not appear in the equation
- An equation can contain up to 100 parameters

Please note that when entering parameters, certain reserved keywords, mathematical function names and internal function names cannot be used.

\* Currently, the typical rate constant variable 'k' cannot be used as a variable name. It is recommended that R is used in its place.

Example parameters:

- A
- MyParam
- MyVar1
- R
- R1
- Rb

Internal Constants:

pi = 3.14159...

## Supported Mathematical Operations:

<b>Mathematical Function</b>	<b>Operator</b>	<b>Usage</b>
Power	^	num1^num2
Multiplication	*	num1*num2
Division	/	num1/num2
Integer Division	\	num1\num2
Modulus	mod	num1 mod num2
Addition	+	num1+num2
Subtraction	-	num1-num2
Negation	-	-num1
Parenthesis	(	(num1)
Parenthesis	)	(num1)

## Supported General Functions

<b>Function</b>	<b>Usage</b>	<b>Description</b>
abs	abs(number)	Absolute Value
exp	exp(number)	Exponential
fix	fix(number)	Similar to int
int	int(number)	Integer
log	log(number)	Natural log
ln	ln(number)	Natural log
log10	log10(number)	Log base 10
rnd	rnd(number)	Random Number
sgn	sng(number)	Sign of a number
sqr	sqr(number)	Square Root
floor	floor(number)	Integer below number
ceil	ceil(number)	Integer above number
max	max(num1,num2)	Maximum of num1 or num2
min	min(num1,num2)	Minimum of num1 or num2
mag	mag(num1,num2)	Vector magnitude of num1 and num2

## Supported Trigonometric Functions:

<b>Function</b>	<b>Usage</b>	<b>Description</b>
cos	cos(number)	Cosine
sin	sin(number)	Sine
tan	tan(number)	Tangent
arcsin	arcsin(number)	Inverse Sine
arccos	arccos(number)	Inverse Cosine
arctan	arctan(number)	Inverse Tangent
sec	sec(number)	Secant
cosec	cosec(number)	Cosecant
cotan	cotan(number)	Cotangent
arcsec	arcsec(number)	Inverse Secant
arccosec	arccosec	Inverse Cosecant
arccotan	arccotan	Inverse Cotangent
hsin	hsin	Hyperbolic Sine
hcos	hcos	Hyperbolic Cosine
htan	htan	Hyperbolic Tangent
hsec	hsec	Hyperbolic Secant
hcosec	hcosec	Hyperbolic Cosecant
hcotan	hcotan	Hyperbolic Cotangent
harcsin	harcsin	Inverse Hyperbolic Sine
harccos	harccos	Inverse Hyperbolic Cosine
harctan	harctan	Inverse Hyperbolic Tangent
harcsec	harcsec	Inverse Hyperbolic Secant
harccosec	harccosec	Inverse Hyperbolic Cosecant
harccotan	harccotan	Inverse Hyperbolic Cotangent

## Helper Functions:

rtod	rtod(radians)	Radians to Degrees
dtor	dtor(degrees)	Degrees to Radians

## Example Equation:

$$((A * R1 * \exp(-R1 * X)) / (R1 + A * R2 * (1 - \exp(-R1 * X))))$$

In this model, X has been used as the independent variable. As X is representing time, it may be more descriptive to use t for time:

$$((A * R1 * \exp(-R1 * t)) / (R1 + A * R2 * (1 - \exp(-R1 * t))))$$

Once a model has been entered, named and any notes added, it can be validated and entered into the equation library. If a backup is ever required, the equation file path is typically:

C:\Documents and Settings\All Users\Application Data\Kinetic Studio\equations.xml

## Selecting a Custom Model

Scroll through the grid to select the required model. Once the line is highlighted, press '**Use Equation**' to return back to the fitting estimator and use the chosen model.

Once selected, the model is entered into the equation drop down list for convenience.

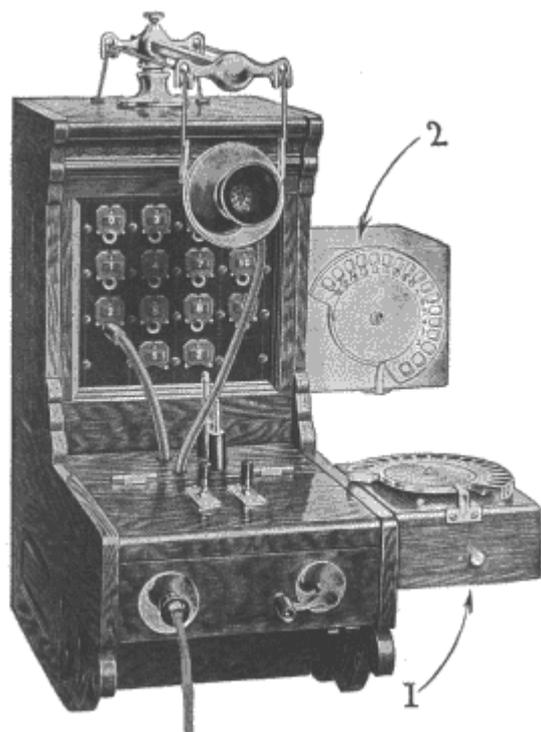
If Kinetic Studio is unable to provide any kind of estimation, it may generate an error message and won't show a preview line in the chart window.

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# Support, Updates and Suggestions

For any support enquiries, update requests or suggestions please contact TgK Scientific Limited.



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When requesting software support that's related to a bug or error message, please provide as much detailed information as possible. Please note down what action was performed prior to the issue and if deemed appropriate, please supply the dataset related to the problem.

The version of Kinetic Studio is indicated in the main starting splash screen and in the status bar at the bottom of the application.

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